

1 **Evaluation of conventional or hydrolyzed stickwater from food-grade skipjack**
2 **tuna by-product in diet for hybrid grouper (*Epinephelus fuscoguttatus* ♀ ×**
3 ***Epinephelus lanceolatus* ♂)**

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17

18 **Abstract**

19 The aim of this study was to investigate how dietary conventional (CSW) or hydrolyzed (HSW)
20 stickwater from skipjack tuna by-product affected growth and feed utilization of hybrid grouper.
21 Nine isonitrogenous (500-505 g crude protein (CP) (kg DM)⁻¹) and iso-energetic (20.3-20.9 MJ
22 gross energy (GE) (kg DM)⁻¹) extruded diets were formulated. A control diet (FMC) was produced
23 with 450 g kg⁻¹ inclusion of fish meal (FM), but without stickwater. The other eight diets were made
24 with a 1:1:1 mixture (dry matter based) of soy protein concentrate (SPC), corn gluten meal (CGM),
25 and CSW or HSW replacing 10, 20, 40 and 60% of FM from the FMC diet. The diets were fed to
26 triplicate groups of hybrid grouper 4 times a day for 8 weeks. The fish were raised in a recirculated
27 aquaculture system (RAS) with 25 ppt salinity and 30°C average temperature. Feed intake (FI) did
28 not significantly ($P > 0.05$) differ among the dietary treatments. Gradually increased replacement
29 with CSW resulted in linear, negative dose response in weight gain (WG), and a linear positive
30 response in FCR. The hepatosomatic index (HSI) decreased linearly. Whole body CP, lipid and ash,
31 and the apparent digestibility (ADC) of energy and the amino acids Ile and Met decreased in a
32 quadratic manner. Linear, negative responses were seen for the ADC of Lys, Thr, Val, and the sum
33 of essential and total amino acids. Replacing with HSW did not significantly affect WG, FCR, HSI,
34 VSI, whole body dry matter, CP, fat contents, protein- or energy efficiencies. Whole-body ash
35 content showed a negative quadratic response. Factorial ANOVA showed that fish fed HSW grew
36 significantly faster and converted the feed more efficiently than the CSW. There was also a tendency
37 ($P=0.061$) that fish fed the HSW diets ate more than the fish receiving the CSW feeds. In conclusion,

38 both feeds with CSW and HSW supported rapid growth and efficient feed conversion of hybrid
39 grouper. Fish fed diets with HSW grew faster and utilized the nutrients more efficiently than did the
40 fish fed diets with CSW.

41

42 **Keywords:** Hydrolysate, Stickwater, Tuna by-product, Hybrid grouper, Growth performance,
43 Digestibility

44

45 **1. Introduction**

46 High market price and demand for fish meal (FM) in the fish feed industry and rising ecological
47 concerns about FM use has prompted research to identify alternative protein sources to minimize
48 the use of FM in fish feeds (Shi et al., 2019, Siddik et al., 2019). High quality protein sources in
49 terms of palatability, essential amino acid profiles and high digestibility are needed to satisfy these
50 demands. Plant protein sources can successfully be used as FM replacer in feeds for omnivorous
51 fish species (Egerton et al., 2020). However, replacing fishmeal with plant proteins may affect
52 growth and health of carnivorous fish (Green et al., 2013). Plant protein sources have been
53 associated with indigestible carbohydrates and antinutritional factors which hinders feed intake,
54 growth, digestion, nutrient absorption, and disease resistance in fish (Shi et al., 2019; Richard et
55 al., 2017). Most of plant protein sources are deficient in one or several essential amino acids and
56 low molecular-weight compounds like taurine and choline which are vital for growth and health of
57 carnivorous fish (Faudzi et al., 2017; Hansen et al., 2020). In this regard, other marine protein
58 sources other than FM are increasingly used in fish feeds. This use of marine feed ingredients has
59 been reported to support growth, digestibility, and immunity like that obtained in fish fed FM
60 (Estruch et al., 2020; Kotzamanis et al., 2007).

61

62 Hybrid grouper (*Epinephelus fuscoguttatus* ♀ × *Epinephelus lanceolatus* ♂) is a species firstly
63 created in 2006 in Malaysia (Ch'ng & Senoo, 2008). Commercial aquaculture of this hybrid has a
64 comparative advantage over its parental species, the brown-marbled grouper (*E. fuscoguttatus*)
65 and giant grouper (*E. lanceolatus*) in the term of growth rate (Dennis et al., 2020), disease
66 resistance, and ability to adapt to a wide salinity range (Ye et al., 2020). This has made hybrid
67 grouper a promising marine cultured species in China. As a carnivorous fish, it has a high protein
68 demand in feed. Yong et al. (2019) recommended 50% dietary protein for the juveniles of this
69 species while Jiang et al. (2016) suggested an optimal protein requirement of 53.5%. A few studies
70 had attempted to replace a fraction of FM in diet of hybrid grouper with different kind of
71 ingredients such as hemoglobin powder (Yao et al., 2018), poultry by-product meal and insect
72 meal (Mohamad-zulkifli et al., 2019), soybean meal (Zhou et al., 2020), peanut meal (Ye et al.,
73 2020) and soy protein concentrate (SPC) (Wang et al., 2020).

74

75 Currently, fractions from FM processing are partially recovered to be used as fish feed ingredients.
76 Stickwater (SW) consists of water solubles from FM processing. This feed ingredient has been
77 used to produce aquatic species with good performance including yellow catfish (*Pelteobagrus*
78 *fulvidraco*) (Wu et al., 2018), giant freshwater prawn (*Macrobrachium rosenbergii*) (Wattanakul
79 et al., 2017), and Atlantic salmon (*Salmo salar* L.) (Berge and Storebakken, 1996; Kousoulaki et
80 al., 2009). The good performance is ascribed to water soluble peptides, free amino acids, taurine,
81 lipid, vitamins, and minerals (Mahdabi & Hosseini Shekarabi, 2018; Wu et al., 2018; Bechtel,

82 2005). SW of high freshness can also be used as feed attractant (Wattanukul et al., 2017). The
83 concentration of water-soluble proteins is high and makes SW a digestible pellet binder. This leads
84 to improved feed physical quality, reducing nutrient leaching and pollution of the environment
85 (del Valle & Aguilera, 1991; Samuelsen et al., 2012). Despite of the potentiality of SW as
86 aquafeed ingredient, little or no information is known about its potential for use in feeds for
87 grouper.

88

89 Hydrolysates of fish protein are characterized by a balanced amino acid profile (Chalamaiah et al.,
90 2012) and antioxidant properties which can boost immunity of fish to fight pathogens (Saadaoui et
91 al., 2019; Murray et al., 2003). Gisbert et al. (2018) reported that a diet containing marine protein
92 hydrolysate enhanced non-specific humoral immunity of European sea bass (*Dicentrarchus*
93 *labrax*), affected by an outbreak of *Vibrio pelagius* infection. Dietary SW and hydrolysates have
94 also been reported to increase appetite, enhance digestible enzyme activities, and improve
95 absorption of nutrients, resulting in boosted fish growth (Aksnes et al., 2006; Kousoulaki et al.,
96 2012; Ospina-Salazar et al., 2016; Khosravi et al., 2015). Protein hydrolysates had also been
97 reported to improve nutrient absorption due to their content of readily digestible free amino acids
98 and small peptides (Olsen & Toppe, 2017). However, excessive use of SW and protein
99 hydrolysates may result in decreased growth performance (Hevrøy et al., 2005).

100

101 It is possible to have good growth performance in carnivorous fish like Atlantic salmon fed a diet
102 completely devoid of marine protein, but with marine lipids (Davidson et al., 2016). However, a
103 combination of marine co-products and terrestrial protein sources may ease formulation and
104 processing of high-quality fish feed. Thus, the current study aimed to: 1) to investigate how
105 conventional (CSW) or hydrolyzed (HSW) stick water from skipjack tuna (*Katsuwonus pelamis*)
106 by-product affected growth and feed utilization of hybrid grouper, 2) to compare the effect of the
107 two types of SW as alternative protein of FM to reveal if hydrolyzation would improve the
108 performance on the above responses and 3) to determine the optimal inclusion level of CSW or
109 HSW in hybrid grouper diet. This study also aims at contributing to the development of cost
110 effective and nutrient-balanced diets for hybrid grouper using SW.

111

112 **2 Materials and methods**

113 2.1 Main ingredients and diets

114 CSW or HSW were produced from the same batch of filleting by-product (heads, guts, trimmings,
115 and frames) from food-grade skipjack tuna. The CSW was made by vacuum concentrating SW
116 from a fishmeal processing line, while the HSW was subject to enzymatic hydrolysis prior to
117 vacuum concentration. The CSW was an experimental batch produced especially for current
118 experiment. The HSW is commercially available from Zhejiang Fengyu Marine Organism
119 Products Co., Ltd., Zhoushan, Zhejiang, China. The characteristics of the two stickwater qualities
120 is presented in Table 1. Compared to the CSW, the dry matter of the HSW contained more acid
121 soluble protein, peptides, and amino acid nitrogen. Histamine was higher in the CSW. The two
122 hydrolysates had similar concentrations of hydrolyzed amino acids and total free amino acids in
123 their dry matter. Peptides in the CSW had the highest contribution to peptides ranging 500 Dalton
124 (Da) and below, and 5 000 Da and above, both when expressed as a part of dry matter, and as a
125 part of acid soluble protein. The hydrolysate had a lower proportion of peptides sized less than

126 180 Da than in the range of 500-180 Da. Except from this, the contribution of peptides declined in
127 a polynomial manner with peptides ranging from 500-180 Da ($337.0 \text{ g (kg soluble protein)}^{-1}$), to
128 more than 10 000 Da ($3.3 \text{ g (kg soluble protein)}^{-1}$).

129

130 The analyzed compositions of the protein-rich ingredients are presented in Table 2. The most
131 notable difference in proximate composition between CSW and HSW was that the CSW contained
132 47% dry matter (DM), while the DM content of the HSW was 5 percent units higher. There also
133 was a high degree of similarity between the two hydrolysates with respect to the amino acid
134 composition.

135

136 Nine isonitrogenous ($500\text{-}505 \text{ g crude protein (CP) (kg DM)}^{-1}$) and iso-energetic ($20.3\text{-}20.9 \text{ MJ}$
137 $\text{gross energy (GE) (kg DM)}^{-1}$) diets were formulated as described in Table 3. The FMC was a
138 control diet with 450 g kg^{-1} high-quality FM, but without CSW or HSW. In the other 8 diets a
139 1:1:1 (dry matter based) mixture of SPC, corn gluten meal (CGM), and stickwater (CSW or HSW)
140 was used to gradually replace FM from the FMC diet. A 2×4 factorial design was adopted with 2
141 stickwater types (CSW or HSW) in combination with 4 levels of FM replacement (10, 20, 40 and
142 60%). The 8 diets in which FM was replaced were named as CSW10 or HSW10, CSW20 or
143 HSW20, CSW40 or HSW40, and CSW60 or HSW60, respectively. Crystalline *L*-lysine and *D*,
144 *L*-methionine were also supplemented to optimize the dietary EAA profile of the diets with 40 and
145 60% replacement. The same diets were also supplemented with mono calcium phosphate (MCP).
146 Yttrium oxide (Y_2O_3) was used as inert marker for digestibility measurement in accordance with
147 Austreng et al. (2000).

148

149 The diets were extruded at the Feed Technology Laboratory of the Sino-European Aquatic
150 Nutrition and Feed Resources Institute, Zhejiang Ocean University. All dry ingredients were
151 ground through a 0.18-mm screen and mixed. Then 40% of water and relevant SW were sprayed
152 into the mash and mixed for 30 min. The moistened mash was kept at room temperature for 12h to
153 facilitate water penetration. The moistened mash was heated to $95\text{-}100 \text{ }^\circ\text{C}$ for 5 min by microwave
154 preconditioning before it was extruded by a laboratory scale twin-screw extruder (Saibainuo,
155 SYSLG30-IV, Jinan, China) with 5.0 mm die. The extrusion parameters that were controlled
156 included feeding rate, screw speed and 4 individual barrel section temperatures, targeting a slowly
157 sinking pellet with bulk density of the extrudate at around 520 g l^{-1} . After forced air drying, the
158 pellets were coated with fish oil using a laboratory scale vacuum coater. The oil had been
159 pre-heated to $50 \text{ }^\circ\text{C}$ in a water bath. After sieving and removal of damaged pellets, the finished
160 diets were stored at $-10 \text{ }^\circ\text{C}$ until used.

161

162 2.2 Fish and feeding

163 A batch of about 3000 hybrid grouper fingerlings with individual weight around 5g were obtained
164 from a local hatchery (Hongsheng Aquaculture Co., Xiangshan, Zhejiang) and kept for one year in
165 the Fish Laboratory of Sino-European Aquatic Nutrition and Feed Resource Institute, Zhejiang
166 Ocean University, where they were fed a commercial feed with 52% CP and 8% fat. Four weeks
167 prior to the start of feeding trial, 300 groupers with uniform size were anaesthetized with MS-222
168 (1 g l^{-1}) and tagged by injecting a Passive Integrated Transponder (PIT, Smartrac N.V., Amsterdam,
169 Netherland) under the superficial skin close to the dorsal fin.

170

171 The feeding trial was conducted in an indoor recirculated aquaculture system (RAS). Before the
172 trial started, the fish were deprived of feed for 48h. Then a total of 540 hybrid grouper with
173 average weight of 0.36 kg were randomly distributed into 27 cylindrical fiberglass tanks with a
174 total volume of 500 l. Ten fish were tagged and the remaining 10 individuals per tank were tagged.
175 Each tank was supplied with seawater at a flow rate of 4-5 l min⁻¹ and aeration 24h day⁻¹. A
176 photoperiod of 14L:10D was maintained. The water temperature ranged from 27.5 to 32.0 °C,
177 with an average at 30.0 °C. Dissolved oxygen levels were above 5.0 mg l⁻¹ in the outlet water,
178 ammonia less than 0.2 mg l⁻¹, and salinity around 25 ppt, based on daily measurements. Each diet
179 was manually fed to fish in 3 parallel tanks, 4 meals per day (08:00, 11:30, 15:00 and 20:00) with
180 45 min per meal. After each feeding, all uneaten pellets were siphoned out immediately and
181 counted. Uneaten feed was quantified by the method of Zhang et al. (2012a), except the average
182 pellet weight for each feed was obtained by counting 3×1000 pellets. The daily feeding rate was
183 tentatively set 10% in excess based on the average feed intake over the last 3 day's feeding, and
184 fish received more feed if they showed signs of feeding at the end of each meal. The feeding trial
185 lasted for 8 weeks. The entire study consisted of a 56-day feeding trial and a simultaneous 30-day
186 digestibility trial.

187

188 2.3 Sampling

189 Before the feeding trial started, 3×5 fish from the acclimation tank were depleted of feed for 24 h,
190 killed by an overdose of MS-222, and kept at -20 °C for whole-body analysis. In the beginning
191 (Day 0) and at Days 14, 28, and 42, all fish were depleted of feed for 24 h, then gently netted out
192 and anaesthetized. The PIT tagged groupers were weighed individually while the other fish were
193 weighed in batch after wipe-drying.

194

195 At the end of feeding trial (Day 57), 3 PIT tagged fish were randomly sampled from each tank for
196 plasma and liver samples. Fish were weighed individually, blood was drawn from the caudal vein
197 with heparinized vacutainers, kept on ice until centrifugation (3000 G for 10 min). The obtained
198 plasma was aliquoted into two separate Eppendorf tubes, frozen in liquid N₂ and kept at -80 °C
199 until analysis. The same fish were dissected to remove whole viscera. The contents in stomach and
200 intestine, liver, and carcass were weighed separately. Another 3 untagged fish were randomly
201 taken from each tank, measured for weight and body length individually, and then killed by a blow
202 to the head. Fish were dissected to remove the contents in stomach and intestine. The whole
203 viscera, liver and carcass were weighed separately, and then stored at -20 °C for whole body
204 analysis.

205

206 Each of the diets was also fed to another triplicate tanks of fish for digestibility assessment, with
207 22 fish (320g in average) in each tank. The same feeding strategy and husbandry condition as
208 growth trial were adopted. Fecal samples were obtained by careful stripping from the last 5 cm of
209 the distal intestine of fish after anaesthetization on Day 5, Day 15, and Day 30, and then pooled by
210 tank and stored at -20 °C prior to analysis.

211

212 2.4 Analyses

213 The initial and final whole-body samples within same tank were homogenized by a meat grinder,

214 autoclaved (YXQ-LS, Xunbo, Shanghai, China) at 120 °C for 30 min, re-homogenized, oven-dried
215 (Jinghong, Zhejiang, China) at 80 °C, and finely ground into powder prior to analysis. Pooled
216 feces samples were freeze-dried and ground with a pestle and mortar. The processed fish samples,
217 feed and ingredient samples were analyzed for dry matter (105 °C to constant weight), Kjeldahl N
218 (Opsis KD-310, Sweden), crude lipid (ether extraction, Opsis SX-360, Sweden), gross energy
219 (Parr, 1271, USA), and ash (550°C, Muffle furnace). Amino acids (except tryptophan) of all
220 samples were analyzed by amino acid analyzer (Hitachi, L-8900, Japan) with 6 M HCl hydrolysis
221 for 22-24 h at 110°C. The sulphur-containing amino acids were determined based on a process of
222 oxidative hydrolysis with performic acid for 30 min at 55°C. Yttrium concentration in feed and
223 feces were determined by inductively coupled plasma mass spectroscopy (Agilent, ICP-MS 7900,
224 USA) after complete digestion of homogenized and dried sample in HNO₃ after cooking in a
225 microwave oven for 1 h (Zhang et al., 2012b). Chemical compositions and peptide contents of
226 CSW and HSW were analyzed from Analysis and Testing Center of Jiangnan University (Wuxi,
227 China) according to the method of GB/T 22729-2008 and previous study (Wu et al., 2018). Briefly,
228 acid soluble protein and peptide molecular weight distribution were analyzed by Kjeldahl method
229 and the HPLC, respectively.

230

231 2.5. Calculations and statistical analysis

232 Feed intake (FI) was quantified by subtracting uneaten feed from the amount of fed on a dry
233 matter basis. Weight gain (WG, g) was calculated as: $WG = FBW - IBW$, where FBW and IBW
234 represent final body weight and initial body weight, respectively. Feed conversion ratio (FCR) was
235 calculated as: $FI \times (FBW - IBW)^{-1}$. Apparent digestibility coefficients of individual nutrient and
236 energy were calculated as: $100 \times (1 - (Y_d \times Y_f^{-1} \times N_f \times N_d^{-1}))$, where Y_d and Y_f represent the
237 concentration of yttrium in diet and faeces, N_d and N_f represent the concentration of individual
238 nutrient or energy in diet and faeces, respectively. Nutrient and energy retentions were calculated
239 as: $100 \times (N_1 \times FBW - N_0 \times IBW) \times (N_d \times FI)^{-1}$, where N_0 and N_1 represent the nutrient or energy
240 concentration in the initial and final whole-body samples, respectively. Hepatosomatic index
241 (HSI, %) or viscerosomatic index (VSI, %) was calculated as: $100 \times (\text{weight of organ}) \times (\text{total fish}$
242 $\text{weight})^{-1}$. Condition factor (CF) was calculated as: $100 \times (\text{fish weight}) \times (\text{body length})^{-3}$, where
243 weight is expressed in g and length is in cm.

244

245 Statistical analysis was done within each type of SW (with or without hydrolysis), and differences
246 were considered significant for $P < 0.05$. The analyses were linear or quadratic regressions based
247 on which model gave best fit to the data, and one-way ANOVA. Quadratic regressions were only
248 presented when the regression coefficient of the 2nd degree component was statistically significant
249 ($P < 0.05$). Maxima or minima in quadratic regressions were calculated by setting the first
250 derivative of the equation to 0. Significant differences in ANOVA were ranked by the Pdiff routine
251 under LSMEANS, and indicated by different superscript letter a, b, c. A two-way factorial
252 ANOVA with the factors SW type (n=2) and replacement level (n=3) was also conducted. Only
253 the effects of SW are reported since the effects of replacement level effects were analyzed more in
254 detail by regression within SW type. The statistical analyses were conducted using the SAS
255 version 9.4 computer software (SAS Institute Inc., Cary, NC, USA).

256

257 3. Results

258 No mortality occurred, and all the fish had high feed intake and grew well. FI, WG, and FCR of
259 grouper fed diets with different levels of CSW or HSW combinations substituting FM are
260 presented in Table 4. Both FBW and FI did not significantly differ among any of the dietary
261 treatments. WG was linearly decreased in response to CSW combination partly replacing FMC.
262 Only fish fed CSW60 had significantly lower WG than fish fed the FMC diet. FCR was linearly
263 increased in response to the increasing dose of CSW combination. The fish fed CSW40 and
264 CSW60 had significantly higher FCR than fish fed the FMC diet. No significant difference in
265 WG or FCR were found in the fish fed the diets containing HSW. Furthermore, none of the
266 diets containing HSW resulted in WG or FCR being significantly different from the results
267 obtained with the FMC diet. The factorial analysis (Table 5) revealed a trend ($P = 0.061$)
268 indicating higher feed intake in the fish fed the HSW than what was seen with the CSW diets.
269 FBW and WG were significantly higher and FCR was significantly lower in fish fed the HSW
270 than in the fish fed the CSW diets. SW inclusion of 10 and 20% resulted in higher WG than 60%,
271 while 10 and 20% inclusion resulted in lowest FCR.

272

273 The only somatic index that came out significant was the HSI of the groupers fed the diets with
274 CSW (Table 6). The regression analysis showed that HSI significantly decreased with increasing
275 dietary concentration of CSW, in a linear manner. The HSI of grouper fed the FMC and CSW10
276 were significantly higher than that of the fish fed CSW40 and CSW60. VSI and CF did not
277 significantly differ among fish fed diets with CSW. Simultaneously, diets containing HSW did
278 not significantly affect HSI, VSI or CF. The factorial ANOVA (Table 7) confirmed that HSI was
279 significantly higher in fish fed HSW than CSW. The significant interaction of SW type and
280 inclusion level is rationalized by HSI of the grouper being affected by dietary level of CSW, but
281 not that of HSW.

282

283 Whole-body dry matter, protein retention (PRE), and energy retention (ERE) efficiencies were not
284 significantly affected by gradually replacing FM in FMC diet with CSW or HSW combinations
285 (Table 8). The response in whole-body crude protein to gradually replacing FMC by CSW was
286 best described by a quadratic curve with maximum at 39.1 % replacement. Whole-body fat
287 content responded in a similar manner, with a maximum at 39.3 %. Fish fed CSW40 also
288 contained significantly more lipid than those fed FMC. Crude protein was significantly higher in
289 grouper fed the CSW 20-60 diets than those fed the FMC. Whole body ash was apparently
290 reduced in response both to increased CSW and HSW. However, SPC was added to the diets
291 proportionally to two SW. The factorial analysis (Table 9) furtherly confirmed the significant
292 differences in ash content only caused by inclusion level not the SW type.

293

294 Apparent digestibility coefficients (ADC) of crude protein, energy, essential and semi-essential
295 amino acids are shown in Table 10. No significant effects of dietary CSW inclusion on the ADC of
296 N, Arg, Cys, His, Leu, Phe, and Tyr were found. The regression of CSW on the ADC of energy
297 was quadratic, with a maximum at 20.0 %, while quadratic responses of ADC of Ile and Met had
298 minimum at 37.3 % and 36.7 %, respectively. The ADC of Lys, Thr, Val, and the ADC of the sum
299 of EAA, and that of the sum of AA responded to increasing concentration of CSW by linear
300 decline. ADC of Met and Val showed quadratic responses to increasing HSW with respective
301 minima at 45.9 and 11.3 %. The ADC of Arg, Ile, Leu, Lys, Thr, Tyr, the sum of EAA, and that of

302 AA were linearly negative, while ADC of N, energy, Cys, His, and Phe were not significantly
303 affected by dietary HSW inclusion. The factorial analysis (Table 11) did not reveal significant
304 differences caused by SW type. Inclusion level significantly affected all the analyzed
305 digestibilities except of N and Phe. Generally, ADC values moderately (1-3%) declined when the
306 inclusion level increased from 10 to 60%, with some exceptions (ADC of Cys 4%, Lys 5%, and
307 Thr 5%).
308

309 **4. Discussion**

310 This study was designed to explore if CSW or HSW from processing of food grade tuna
311 by-product had a potential for partially replacing FM in diets for hybrid grouper. The main criteria
312 used to evaluate these novel feed ingredients were growth rates, feed utilization, body composition
313 and nutrient digestibilities. Throughout the whole experiment period, all groups of hybrid grouper
314 were active and readily accepted the feeds. Feed intake was not significantly influenced by
315 different dietary treatments, but a trend indicates that the HSW was preferred when compared to
316 the CSW. The observation that the palatability of feed was not significantly affected by type of SW,
317 or rate of replacement ranging from 0 to 60% of the dietary FM is contrary to what has been
318 obtained with other ingredients by which feed intake decreased when a high proportion of FM was
319 replaced (Yao et al., 2018; Glencross et al., 2011). One probable explanation to the high feed
320 intake both in grouper fed CSW and HSW, is the high freshness of the food-grade skipjack tuna
321 by-products from which the SW was produced. Histamine concentration is used as an indicator of
322 freshness in fish meal, and 670 mg histidine (kg DM)⁻¹ in the CSW was similar to values obtained
323 from high-quality Chilean fish meal by Anderson et al. (1997).
324

325 WG linearly decreased with an increase in inclusion levels of CSW whereas WG of fish fed HSW
326 were not significantly affected. The highest WG value was observed in fish fed HSW20. Similar
327 results were observed when yellow catfish fed diets with HSW and FM grew faster than those fed
328 CSW (Wu et al., 2018). Feeding rice field eel (*Monopterus albus*) with low inclusion levels of
329 HSW (10-15%) was reported to increase the WG (Shi et al., 2019). Inclusion of HSW, even in
330 highest level (HSW60) did not have significant negative effect in fish growth, defying the
331 decreased growth performance observed when large portion of FM was replaced by fish
332 hydrolysate (Zheng et al., 2012). In this study, negative effect on the FCR was observed as FM
333 replacement with CSW increased in the diets. Poor feed utilization may be a result of the change
334 in amino acid profiles (Wattanakul et al., 2017).
335

336 The lack of significant differences obtained for FCR between fish fed FMC diet and those fed
337 diets with HSW inclusion suggest that HSW can replace a major proportion of FM without
338 negatively affecting growth and feed utilization of hybrid grouper. Likewise, an attempt to replace
339 FM with SPC was done in hybrid grouper but there was a significant negative effect in survival,
340 growth, and feed utilization (Wang et al., 2020). FM was replaced with peanut meal (PNM) up to
341 50 % with no significant effect on growth performance, but negatively affected immunity and
342 intestinal microbiota of hybrid grouper (Ye et al., 2020). WG of fish groups fed HSW was
343 numerically higher than that of corresponding groups fed CSW at the same level. Also, relatively
344 lower FCRs were observed in fish fed FMC and HSW diets, implying that feed was highly utilized
345 by fish fed those diets. Similar results on growth and feed utilization were obtained when FM was

346 replaced by SW in Nile tilapia (*Oreochromis niloticus*) (Wattanakul et al., 2019). The factorial
347 analysis conducted in this experiment also showed that fish fed diets with HSW combination had
348 significantly higher FBW and WG but lower FCR than that of fish fed diets with CSW
349 combination. Similarly, juvenile brown-marbled grouper (*E. fuscoguttatus*) fed milkfish offal
350 hydrolysate had significantly improved feed intake, growth and feed utilization compared to that
351 fed milkfish offal (Mamaug & Ragaza, 2016). The results of present study also agreed in extent
352 with the studies in which the inclusion of low levels of HSW in diets enhanced feed utilization and
353 growth of rice field eel (Shi et al., 2019). Other marine protein hydrolysates have also brought
354 positive effect on growth of Atlantic salmon (Berge and Storebakken, 1996; Refstie et al., 2004).

355

356 Generally, improved growth and feed conversion can be expected due to improved utilization of
357 protein from HSW because the SW being hydrolyzed (Tonheim et al., 2005). Nutrients within the
358 hydrolysates then tend to be rapidly absorbed and assimilate through the intestinal membrane. The
359 reason for rapid absorption of HSW is that the amino acid chains had been hydrolyzed to release
360 shorter peptides and free amino acids (Potier & Tomé, 2008). The factor that contributed most to
361 grouper fed HSW growing faster than those fed the CSW diets seemed to be the tendency
362 ($P=0.061$) for higher feed intake in the fish fed the diets with HSW.

363

364 The lower DM content of CSW than HSW probably reflects higher water binding in conventional
365 than partially hydrolyzed fish proteins (Kristinson & Rasco, 2000). No significant effect was
366 observed on VSI and CF between fish fed FMC diet and diets with CSW. However, fish fed the
367 CSW40 and CSW60 diets had significantly lower HSI than that of fish fed FMC, CSW10 and
368 CSW20 diets. HSI values did not correlate with body lipid content, contrary to previous studies in
369 juvenile silvery-black porgy (*Sparidentex hasta*) (Yaghoubi et al., 2016) and Japanese flounder
370 (*Paralichthys olivaceus*) (Ye et al., 2011). No significant differences were observed on HSI, VSI,
371 and CF between fish fed FMC diet and diets with HSW. Similar results were observed in the same
372 species when FM was replaced to PNM up to 50% (Ye et al., 2020). In contrast, HSI was found to
373 increase when FM was replaced by plant protein (Ye et al., 2019a) and by animal protein blend of
374 poultry by-product meal, spray-dried blood meal and shrimp meal (Ye et al., 2019b).

375

376 Two alternative explanations may explain why fish fed CSW and HSW had lower whole-body ash
377 content than the ones fed the FMC diet. One was the characteristics of the FM and the two SW.
378 This contrasts with the results obtained by Kousoulaki et al. (2009) who found that ash content of
379 Atlantic salmon increased with the increase in SW levels replacing FM. The other explanation was
380 that reduced ash content was due to phytic acid in SPC, and which was added to the diets
381 proportionally to two SW. Dietary concentrations of SPC paralleled that of CSW and HSW (Table
382 3). The regressions of percentwise dietary SPC inclusion on whole-body ash also paralleled the
383 results obtained with CSW ($R^2=0.66$) and HSW ($R^2=0.78$). Dietary phytic acid is known to reduce
384 whole-body ash in Atlantic salmon (Denstadli et al, 2006), both due to poor availability of P from
385 phytic acid, and binding cationic elements such as Mg, Zn, and the Ca: P-ratio. These mineral
386 deficiencies cause vertebrae deformities in Atlantic salmon (Helland et al., 2006). It is difficult to
387 solve this challenge in coldwater fish because phytase does not work at low temperatures
388 (Denstadli et al., 2007). In warmwater fish like grouper, phytase can be carefully coated into the

389 feed after extrusion and drying. Hydrolyses of phytic acid will then occur like other digestive
390 processes.

391

392 Neither CSW nor HSW inclusion affected the retention of protein and energy. Crude protein in
393 fish whole body was significantly increased with an increase in CSW in fish diets. No significant
394 differences in whole body dry matter, crude protein and crude fat were observed between fish fed
395 FMC and HSW diets. The results agree with those of a previous study in juvenile snakehead
396 (*Ophiocephalus argus*) (Yun et al., 2014).

397

398 Determining the digestibility of nutrients is crucial in evaluating the use of novel feed ingredient
399 (de Magalhães et al., 2016; Allan et al., 2000). In this study, the apparent digestibility (ADC) of
400 protein, energy, and individual EAA in diets with SW inclusion ranged from 84.1% to 96.0%,
401 indicating that nutrients were effectively digested by hybrid grouper. The ADC of protein was not
402 affected by HSW or CSW inclusion in diets. Contrary to this study, the ADC of protein decreased
403 with an increase in SPC inclusion in diet for hybrid grouper (Faudzi et al., 2017; Chen et al., 2020).
404 Meanwhile, no significant differences in the ADC of energy were observed between FM and diets
405 with HSW inclusion. CSW60 resulted in significantly lower ADC of energy than CSW10. Apart
406 from amino acid composition, quality of protein source is determined by the bioavailability of
407 essential amino acids which is reflected by digestibility (Wu et al., 2017; Potier & Tomé, 2008).
408 Gut morphology and oxidative stress was reported to be influenced by dietary arginine (Wu et al.,
409 2018) while dietary leucine affected growth hormone (GH) level (Zhou et al., 2019) in hybrid
410 grouper. Therefore, the optimum amount of digested essential amino acids must be examined
411 because of their importance in overall fish performance. Generally, the ADC of amino acids was
412 high in fish fed FMC and diets with low inclusion of SW (10% and 20%). ADC of some amino
413 acids were not affected with FM replacement while there was a correspondingly decrease in ADC
414 of a few amino acids with an increase in SW inclusion levels. Digestibility of the sum of EAA,
415 and that of total AA by fish fed FM, CSW10, HSW10 and HSW20 were significantly higher than
416 other dietary groups. A few studies have touched on the ADC of protein, fat, energy, and
417 phosphorus in fish fed diets with SW inclusion (Kousoulaki et al., 2012; Kousoulaki et al., 2009),
418 but none of them included the ADC of amino acids. Replacing FM with other ingredients has been
419 associated with reduction in digestibility of nutrients, even when the diets meet requirements of
420 specific species (Kousoulaki et al., 2009). However, partial replacement of FM with low levels of
421 SW (10% of FM with SW and up to 20% of FM with HSW) did not generally result in significant
422 negative effects on ADC of nutrients.

423

424 **5. Conclusion**

425 The current study showed that both CSW and HSW from food-grade skipjack tuna co-product are
426 promising protein sources for hybrid grouper feed. The overall performance of HSW was better in
427 both fish growth and feed utilization than CSW. The HSW combined with SPC and CGM with the
428 mixing ratio of 1:1:1 could effectively decrease the inclusion of high-quality FM from 450 g kg⁻¹
429 to 360 g kg⁻¹ in extruded practical diet for hybrid grouper without impairing the feed intake,
430 growth performance, feed utilization, whole-body composition, retentions of protein and energy,
431 nutrient digestibility, and all somatic indices.

432

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438

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673

1 Table 1 Characteristics of conventional (CSW) and hydrolyzed stickwater (HSW)¹.

| | CSW | CSW | HSW | HSW |
|--|-------------------------|---|-------------------------|---|
| Solids, g (kg wet weight) ⁻¹ | 533 | | 580 | |
| Acid soluble protein, g (kg dry matter (DM)) ⁻¹ | 294 | | 674 | |
| Hydrolyzed amino acids, g (kg DM) ⁻¹ | 576 | | 572 | |
| Total free amino acids g (kg DM) ⁻¹ | 65.1 | | 65.0 | |
| Peptides, g (kg DM) ⁻¹ | 229 | | 609 | |
| Amino acid nitrogen, g (kg DM) ⁻¹ | 14.1 | | 15.7 | |
| Histamine, mg (kg DM) ⁻¹ | 670 | | 406 | |
| Peptide size ranges, Da | g (kg DM) ⁻¹ | g (kg acid soluble protein) ⁻¹ | g (kg DM) ⁻¹ | g (kg acid soluble protein) ⁻¹ |
| <180 | 34.9 ² | 118.7 ³ | 130.0 ² | 192.7 ³ |
| 500 - 180 | 43.7 | 148.4 | 227.3 | 337.0 |
| 1 000 - 500 | 8.2 | 27.9 | 143.1 | 212.2 |
| 2 000 -1 000 | 10.2 | 34.5 | 98.9 | 146.7 |
| 3 000 -2 000 | 7.9 | 27.0 | 35.5 | 52.7 |
| 5 000 - 3 000 | 13.2 | 13.2 | 26.3 | 39.0 |
| 10 000 - 5 000 | 31.1 | 105.7 | 11.1 | 16.4 |
| >10 000 | 145.0 | 492.8 | 2.2 | 3.3 |

2

3 ¹Conventional stickwater (CSW) and hydrolyzed stickwater (HSW) from food-grade tuna by-product, Zhejiang

4 Fengyu Marine Organism Products Co., Ltd., Zhoushan, China.

5 ² Calculated by the percentage of the normalized area method x acid soluble protein content (g 100g⁻¹ DM).6 ³ Percentage of the area normalized method (g 100g⁻¹ acid soluble protein)

7

8

9 Table 2 Composition of fish meal (FM), conventional (CSW) or hydrolyzed (HSW) stickwater, soy protein
 10 concentrate (SPC), corn gluten meal (CGM), soybean meal (SBM) and wheat gluten used in experiment (on dry
 11 matter basis)
 12

| Ingredient | FM ¹ | CSW | HSW | SPC ² | CGM ³ | SBM ⁴ | wheat gluten ⁵ |
|--|-----------------|-------------|-------------|------------------|------------------|------------------|---------------------------|
| Composition, kg⁻¹ | | | | | | | |
| Dry matter (DM), g | 933 | 470 | 524 | 928 | 913 | 882 | 926 |
| In DM | | | | | | | |
| Crude protein, g | 750 | 812 | 822 | 683 | 693 | 532 | 815 |
| Crude fat, g | 107 | 40 | 46 | 17 | 5 | 21 | 13 |
| Ash, g | 158 | 192 | 193 | 58 | 19 | 70 | 9 |
| Gross energy, MJ | 21.7 | 18.6 | 18.4 | 20.7 | 23.2 | 19.5 | 23.6 |
| Essential (EAA) ⁶ and semi essential (SEAA) amino acids, g (16 g N)⁻¹ | | | | | | | |
| Arg | 5.22 | 3.98 | 3.97 | 7.21 | 2.93 | 6.99 | 3.27 |
| Cys | 0.63 | 0.35 | 0.36 | 1.47 | 1.35 | 0.98 | 2.14 |
| His | 3.25 | 4.82 | 4.77 | 2.58 | 2.03 | 2.41 | 1.77 |
| Ile | 3.79 | 1.44 | 1.39 | 4.63 | 3.71 | 4.34 | 3.56 |
| Leu | 7.08 | 3.35 | 3.37 | 7.97 | 16.50 | 7.68 | 6.72 |
| Lys | 7.34 | 4.30 | 4.27 | 6.59 | 1.61 | 6.14 | 1.49 |
| Met | 2.45 | 1.24 | 1.21 | 1.57 | 2.06 | 0.70 | 1.90 |
| Phe | 4.05 | 1.91 | 1.87 | 4.97 | 5.84 | 4.82 | 4.66 |
| Thr | 4.14 | 2.32 | 2.27 | 4.36 | 3.34 | 3.90 | 2.45 |
| Val | 4.43 | 2.18 | 2.14 | 5.10 | 4.27 | 4.42 | 4.04 |
| Total EAA and SEAA | 41.8 | 25.5 | 25.3 | 45.0 | 42.3 | 42.4 | 29.9 |
| Total non-essential AA ⁶ | 41.9 | 35.4 | 35.1 | 53.8 | 57.6 | 51.0 | 65.0 |
| Total AA | 83.6 | 60.9 | 60.4 | 98.8 | 99.8 | 93.4 | 94.9 |

13 ¹ Brown fishmeal, Compania Pesquera Del Pacifico Centro S.A, Lima, Peru

14 ² Yihai®, Wilpromil, Glodensea Grain and Oil Industry Co., Ltd, Wilmar, Qinhuangdao, China

15 ³ Lihua Starch Co., Ltd, Wilmar, Qinhuangdao, China

16 ⁴ Zhoushan Good Ocean Grains & Oils Co., Ltd, Zhoushan, China

17 ⁵ Golden Mountain, grade two, Dongguan Yihai Kerry Syral Starch Technology Co., Ltd, Dongguan, China.

18 ⁶ Trp excluded. Cys and Tyr are semi essential (SEAA)

Table 3 Feed formulation and analyzed chemical composition (on dry matter basis).

| Diet | FMC | CSW10 | CSW20 | CSW40 | CSW60 | HSW10 | HSW20 | HSW40 | HSW60 |
|--|-------|-------|-------|-------|-------|-------|-------|-------|-------|
| Ingredients, g kg⁻¹ | | | | | | | | | |
| FM | 450 | 405 | 360 | 270 | 180 | 405 | 360 | 270 | 180 |
| SPC | 100.0 | 114.9 | 129.7 | 159.4 | 189.1 | 114.9 | 129.7 | 159.4 | 189.1 |
| Wheat gluten | 40 | 40 | 40 | 40 | 40 | 40 | 40 | 40 | 40 |
| Wheat flour | 137.7 | 134.6 | 131.7 | 119.7 | 110.6 | 134.6 | 131.7 | 119.7 | 110.6 |
| CSW | 0 | 15.3 | 30.6 | 61.2 | 91.8 | 0 | 0 | 0 | 0 |
| HSW | 0 | 0 | 0 | 0 | 0 | 15.3 | 30.6 | 61.2 | 91.8 |
| Soybean meal | 50 | 50 | 50 | 50 | 50 | 50 | 50 | 50 | 50 |
| CGM | 50.0 | 64.9 | 79.7 | 111.4 | 139.1 | 64.9 | 79.7 | 111.4 | 139.1 |
| Fish oil | 110 | 113 | 116 | 123 | 130 | 113 | 116 | 123 | 130 |
| Premix ¹ | 20 | 20 | 20 | 20 | 20 | 20 | 20 | 20 | 20 |
| L-Lys | 0 | 0 | 0 | 0 | 1.0 | 0 | 0 | 0 | 1.0 |
| DL-Met | 0 | 0 | 0 | 0.5 | 1.1 | 0 | 0 | 0.5 | 1.1 |
| MCP | 10.0 | 10.0 | 10.0 | 12.5 | 15.0 | 10.0 | 10.0 | 12.5 | 15 |
| Y ₂ O ₃ | 10.0 | 10.0 | 10.0 | 10.0 | 10.0 | 10.0 | 10.0 | 10.0 | 10.0 |
| Choline Cl ² | 1.5 | 1.5 | 1.5 | 1.5 | 1.5 | 1.5 | 1.5 | 1.5 | 1.5 |
| Soy lecithin ³ | 20 | 20 | 20 | 20 | 20 | 20 | 20 | 20 | 20 |
| Vitamin C ⁴ | 0.5 | 0.5 | 0.5 | 0.5 | 0.5 | 0.5 | 0.5 | 0.5 | 0.5 |
| Vitamin E ⁵ | 0.3 | 0.3 | 0.3 | 0.3 | 0.3 | 0.3 | 0.3 | 0.3 | 0.3 |
| Analyzed content, kg⁻¹ | | | | | | | | | |
| Dry matter, g | 967 | 943 | 930 | 918 | 953 | 924 | 935 | 926 | 938 |
| Crude protein, g | 502 | 505 | 504 | 503 | 500 | 504 | 503 | 503 | 501 |
| Crude fat, g | 143 | 153 | 156 | 151 | 158 | 146 | 146 | 158 | 148 |
| Ash, g | 116 | 108 | 106 | 103 | 95 | 111 | 106 | 103 | 96 |
| Gross energy, MJ | 20.8 | 20.6 | 20.5 | 20.5 | 20.9 | 20.3 | 20.6 | 20.5 | 20.8 |
| SEAA⁶ and EAA, % | | | | | | | | | |
| Arg | 2.75 | 2.75 | 2.72 | 2.65 | 2.61 | 2.74 | 2.70 | 2.66 | 2.59 |
| Cys | 0.41 | 0.46 | 0.44 | 0.40 | 0.39 | 0.37 | 0.39 | 0.43 | 0.45 |
| His | 1.51 | 1.43 | 1.48 | 1.46 | 1.47 | 1.37 | 1.36 | 1.48 | 1.44 |
| Ile | 1.95 | 2.01 | 1.94 | 1.84 | 1.79 | 2.05 | 2.04 | 1.87 | 1.82 |
| Leu | 4.02 | 4.16 | 4.17 | 4.22 | 4.38 | 4.08 | 4.11 | 4.29 | 4.33 |
| Lys | 3.25 | 3.15 | 2.98 | 2.74 | 2.62 | 3.13 | 2.97 | 2.78 | 2.57 |
| Met | 0.85 | 0.90 | 0.90 | 0.81 | 0.77 | 0.81 | 0.80 | 0.76 | 0.79 |
| Phe | 2.49 | 2.40 | 2.37 | 2.33 | 2.34 | 2.25 | 2.25 | 2.37 | 2.37 |
| Thr | 2.01 | 2.00 | 1.96 | 1.86 | 1.78 | 1.96 | 1.92 | 1.90 | 1.79 |
| Tyr | 1.66 | 1.73 | 1.72 | 1.72 | 1.71 | 1.65 | 1.64 | 1.70 | 1.73 |
| Val | 2.23 | 2.26 | 2.20 | 2.07 | 2.01 | 2.31 | 2.28 | 2.12 | 2.03 |
| ∑EAA | 23.1 | 23.2 | 22.9 | 22.1 | 21.9 | 22.7 | 22.5 | 22.4 | 21.9 |
| ∑AA | 47.0 | 47.2 | 47.0 | 46.1 | 46.3 | 47.1 | 46.8 | 47.0 | 46.3 |

¹ Vitamin premix (mg kg⁻¹ diet): vitamin A 1.95; vitamin B₁ 20; vitamin B₂ 10; vitamin B₆ 30; niacinamide 250; ascorbic acid 5; calcium pantothenate 50; folic acid 20; vitamin E 1200; vitamin K 0.8; vitamin D 0.05; inositol 650; Defatted Rice Bran 150. Mineral premix (mg kg⁻¹ diet): CuSO₄ · 5H₂O 10; FeSO₄ · H₂O 300; ZnSO₄ · H₂O 200; MnSO₄ · H₂O 100; KI (10%) 80; Na₂SeO₃ (10% Se) 67; CoCl₂ · 6H₂O (10% Co) 5; NaCl 100; zeolite 638. Vitamin premix: mineral premix = 2: 1.5.

² 60% Choline Chloride Powder (Vegetable Carrier), Shandong Aocter Group, Shandong, China

³ Youlin[®], powder (> 95%), Beijing Meiyas Phospholipid Technology Co., Ltd, Beijing, China.

⁴ L-Ascorbate-2-Monophosphate, ≥35%, feed grade, Hangzhou Tiannong Bio-nutrition Technology Co., Ltd., Zhejiang, China

⁵ DL-alpha-Tocopherol acetate, 50% feed grade, NHU Co., Ltd., Zhejiang, China

⁶ Trp excluded. Cys and Tyr are semi essential (SEAA)

Table 4 Feed intake, growth performance and feed conversion ratio (FCR) of hybrid grouper fed diets with CSW or HSW combinations

| Parameters | Diet | | | | | <i>P</i> -value | Pooled S.E.M. ¹ | Regression | R ² |
|--|--------------------|---------------------|---------------------|--------------------|--------------------|-----------------|----------------------------|---------------------------------|----------------|
| | FMC | CSW10 | CSW20 | CSW40 | CSW60 | | | | |
| Final body weight (FBW), g fish ⁻¹ | 671 | 667 | 672 | 662 | 632 | 0.07 | 21.3 | | |
| Feed intake (FI), g DM fish ⁻¹ | 213 | 212 | 230 | 216 | 209 | 0.21 | 14.0 | | |
| Weight gain (WG), g fish ⁻¹ | 311 ^a | 301 ^a | 313 ^a | 300 ^a | 276 ^b | 0.026 | 15.9 | - 0.523 x+313 | 0.46 |
| Feed conversion ratio (FCR), g FI (g WG) ⁻¹ | 0.785 ^a | 0.805 ^{ab} | 0.802 ^{ab} | 0.825 ^b | 0.866 ^c | <0.001 | 0.0176 | 1.25 *10 ⁻³ x+ 0.784 | 0.81 |
| Parameters | Diet | | | | | <i>P</i> -value | Pooled S.E.M. ¹ | Regression | R ² |
| | FMC | HSW10 | HSW20 | HSW40 | HSW60 | | | | |
| Final body weight (FBW), g fish ⁻¹ | 671 | 687 | 710 | 669 | 667 | 0.17 | 29.1 | | |
| Feed intake (FI), g DM fish ⁻¹ | 213 | 226 | 236 | 230 | 218 | 0.37 | 18.4 | | |
| Weight gain (WG), g fish ⁻¹ | 311 | 324 | 346 | 310 | 304 | 0.22 | 29.3 | | |
| Feed conversion ratio (FCR), g FI (g WG) ⁻¹ | 0.785 | 0.799 | 0.782 | 0.809 | 0.821 | 0.12 | 0.0247 | | |

For diet codes see Table 2. Different superscript letters ^a, ^b and ^c indicate significant (*P* < 0.05) differences among treatments.

¹ Pooled standard error of means.

Table 5 Factorial analysis of data on feed intake, growth performance and feed utilization of hybrid grouper fed diets with different stickwater types and inclusion levels

| Factor | FBW, g fish ⁻¹ | FI, g DM fish ⁻¹ | WG, g fish ⁻¹ | FCR, g FI (g WG) ⁻¹ |
|--|------------------------------|--------------------------------|-----------------------------|-----------------------------------|
| Stickwater type | | | | |
| CSW | 658 ^b | 217 | 297 ^b | 0.824 ^b |
| HSW | 681 ^a | 228 | 321 ^a | 0.802 ^a |
| Inclusion level, % | | | | |
| 10 | 677 ^{ab} | 219 | 312 ^{ab} | 0.802 ^{ab} |
| 20 | 691 ^a | 233 | 329 ^a | 0.792 ^a |
| 40 | 666 ^{bc} | 223 | 305 ^{bc} | 0.817 ^b |
| 60 | 649 ^c | 214 | 290 ^{cd} | 0.844 ^c |
| Pooled S.E.M ¹ | 15.5 | 21.6 | 26.1 | 0.026 |
| Factorial ANOVA ($P > F$) | | | | |
| Stickwater type | 0.006 | 0.061 | 0.002 | 0.005 |
| Inclusion level | 0.014 | 0.132 | 0.007 | <0.001 |
| Type * level | 0.493 | 0.937 | 0.674 | 0.232 |

Different superscript letters ^{a, b, c,} and ^d indicate significant ($P < 0.05$) differences among treatments.

¹ Pooled standard error of means.

1 Table 6 Somatic indices of hybrid grouper fed diets with CSW or HSW combinations

| Somatic Indices | Diet | | | | | P-value | Pooled S.E.M. ¹ | Regression | R ² |
|--|--------------------|--------------------|--------------------|-------------------|-------------------|---------|----------------------------|----------------------------|----------------|
| | FMC | CSW10 | CSW20 | CSW40 | CSW60 | | | | |
| HSI ² , % | 2.35 ^{ab} | 2.34 ^{ab} | 2.20 ^{bc} | 1.91 ^c | 1.90 ^c | 0.03 | 0.244 | $-8.77 * 10^{-3} x + 2.37$ | 0.56 |
| VSI ³ , % | 12.76 | 12.46 | 16.79 | 12.89 | 14.74 | 0.22 | 3.13 | \ | \ |
| CF ⁴ , g (cm) ⁻³ | 2.86 | 2.96 | 2.83 | 2.68 | 2.66 | 0.16 | 0.195 | \ | \ |
| | Diet | | | | | | | | |
| | FMC | HSW10 | HSW20 | HSW40 | HSW60 | | | | |
| HIS, % | 2.35 | 2.11 | 2.43 | 2.60 | 2.18 | 0.10 | 0.270 | \ | \ |
| VSI, % | 12.76 | 13.75 | 13.62 | 13.24 | 13.1 | 0.18 | 0.64 | \ | \ |
| CF, g (cm) ⁻³ | 2.86 | 2.80 | 2.89 | 2.96 | 2.74 | 0.71 | 0.265 | \ | \ |

2 For diet codes see Table 2. Different superscript letters ^{a,b} and ^c indicate significant ($P < 0.05$) differences among treatments.

3 ¹ Pooled standard error of means.

4 ² Hepatosomatic index.

5 ³ Viscerosomatic index.

6 ⁴ Condition factor.

7

8 Table 7 Factorial analysis of somatic indices of hybrid grouper fed diets with different stickwater types and inclusion level
 9

| Factor | HSI ¹ , % | VSI ² , % | CF ³ , g (cm) ⁻³ |
|-----------------------------------|----------------------|----------------------|--|
| Stickwater type | | | |
| CSW | 2.09 ^a | 14.2 | 2.78 |
| HSW | 2.33 ^b | 13.4 | 2.85 |
| Inclusion level, % | | | |
| 10 | 2.23 | 13.1 | 2.88 |
| 20 | 2.31 | 15.2 | 2.86 |
| 40 | 2.25 | 13.1 | 2.82 |
| 60 | 2.04 | 13.9 | 2.70 |
| Pooled S.E.M. ⁴ | 0.081 | 0.79 | 0.065 |
| Factorial ANOVA (P>F) | | | |
| Stickwater type | <0.01 | 0.33 | 0.33 |
| Inclusion level | 0.14 | 0.23 | 0.24 |
| Type * Level | <0.01 | 0.23 | 0.16 |

10 Different superscript letters ^a and ^b indicate significant ($P < 0.05$) differences among treatments.
 11 ¹ Hepatosomatic index.
 12 ² Viscerosomatic index.
 13 ³ Condition factor.
 14 ⁴ Pooled standard error of means.
 15

16 Table 8 Whole body compositions and nutrient retentions of hybrid grouper fed diets with CSW and HSW combinations
 17

| Items | Diet | | | | | P-value | Pooled S.E.M. ¹ | Regression | R ² |
|-----------------------------------|-------------------|--------------------|--------------------|--------------------|--------------------|---------|----------------------------|--------------------------------------|----------------|
| | FMC | CSW10 | CSW20 | CSW40 | CSW60 | | | | |
| Dry matter, g kg ⁻¹ | 337 | 339 | 332 | 344 | 339 | 0.28 | 7.9 | \ | \ |
| Crude protein, g kg ⁻¹ | 169 ^b | 174 ^{ab} | 176 ^a | 176 ^a | 175 ^a | 0.04 | 3.4 | -0.005x ² +0.391x+169.827 | 0.57 |
| Crude fat, g kg ⁻¹ | 101 ^b | 110 ^{ab} | 109 ^{ab} | 119 ^a | 111 ^{ab} | 0.04 | 7.3 | -0.01x ² +0.786x+100.830 | 0.50 |
| Ash, g kg ⁻¹ | 56.1 ^a | 44.3 ^b | 39.2 ^{bc} | 38.7 ^{bc} | 37.5 ^c | <0.001 | 4.15 | 0.01x ² -0.872x+54.16 | 0.79 |
| Protein retention efficiency, % | 46.7 | 48.9 | 47.4 | 48.8 | 46.0 | 0.47 | 2.95 | \ | \ |
| Energy retention efficiency, % | 52.6 | 57.2 | 52.2 | 57.9 | 56.0 | 0.41 | 5.62 | \ | \ |
| | Diet | | | | | | | | |
| | FMC | HSW10 | HSW20 | HSW40 | HSW60 | | | | |
| Dry matter, g kg ⁻¹ | 337 | 336 | 338 | 334 | 330 | 0.51 | 7.7 | \ | \ |
| Crude protein, g kg ⁻¹ | 169 | 169 | 175 | 173 | 173 | 0.53 | 6.2 | \ | \ |
| Crude fat, g kg ⁻¹ | 101 | 107 | 117 | 113 | 108 | 0.07 | 7.9 | \ | \ |
| Ash, g kg ⁻¹ | 56.1 ^a | 50.1 ^{ab} | 38.6 ^c | 40.5 ^c | 43.0 ^{bc} | <0.01 | 6.31 | 0.012x ² -0.937x+56.17 | 0.64 |
| Protein retention efficiency, % | 46.7 | 45.6 | 52.7 | 45.0 | 47.0 | 0.15 | 3.87 | \ | \ |
| Energy retention efficiency, % | 52.6 | 53.0 | 63.0 | 51.8 | 52.8 | 0.13 | 7.18 | \ | \ |

18 For diet codes see Table 2. Different superscript letters ^a, ^b, and ^c indicate significant ($P < 0.05$) differences among treatments.

19 ¹ Pooled standard error of means.

20

21 Table 9 Factorial analysis of whole-body compositions and nutrient retentions of hybrid grouper fed diets with different stickwater types and inclusion level

| Factor | Dry matter, g kg ⁻¹ | Crude protein, g kg ⁻¹ | Crude fat, g kg ⁻¹ | Ash, g kg ⁻¹ | Nitrogen retention, % | Energy retention, % |
|-----------------------------------|-----------------------------------|--------------------------------------|----------------------------------|----------------------------|--------------------------|------------------------|
| <i>Stickwater type</i> | | | | | | |
| CSW | 338 | 175 | 112 | 39.9 | 47.8 | 56.1 |
| HSW | 336 | 173 | 111 | 42.8 | 47.6 | 55.1 |
| <i>Inclusion level, %</i> | | | | | | |
| 10 | 337 | 171 | 108 | 47.1 ^a | 47.2 | 55.1 |
| 20 | 336 | 176 | 113 | 39.0 ^b | 50.0 | 57.6 |
| 40 | 340 | 175 | 116 | 39.7 ^b | 47.0 | 55.4 |
| 60 | 335 | 174 | 109 | 39.5 ^b | 46.5 | 54.4 |
| Pooled S.E.M. ¹ | 2.6 | 1.4 | 2.7 | 1.50 | 1.11 | 2.24 |
| <i>Factorial ANOVA (P>F)</i> | | | | | | |
| Stickwater type | 0.3 | 0.1 | 0.8 | 0.07 | 0.85 | 0.67 |
| Inclusion level | 0.6 | 0.1 | 0.2 | <0.01 | 0.17 | 0.78 |
| Type * level | 0.1 | 0.8 | 0.3 | 0.51 | 0.04 | 0.07 |

22 For diet codes see Table 2. Different superscript letters ^a and ^b indicate significant ($P < 0.05$) differences among treatments.

23 ¹ Pooled standard error of means.

24

25 Table 10 Apparent digestibility coefficient of nutrients of hybrid grouper fed diets with CSW or HSW combinations

| ADC, % | Diet | | | | | P-value | Pooled S.E.M. ¹ | Regression | R ² |
|-------------------|--------------------|--------------------|---------------------|--------------------|--------------------|---------|-------------------------------|--|----------------|
| | FMC | CSW10 | CSW20 | CSW40 | CSW60 | | | | |
| Nitrogen (N) | 86.4 | 88.5 | 87.5 | 85.9 | 86.4 | 0.14 | 1.58 | \ | \ |
| Energy | 84.1 ^{ab} | 88.0 ^a | 87.3 ^{ab} | 84.6 ^{ab} | 83.9 ^b | 0.01 | 1.81 | -0.003x ² +0.125x+85.313 | 0.34 |
| Arg | 95.5 | 96.0 | 95.4 | 95.0 | 94.8 | 0.18 | 0.74 | \ | \ |
| Cys | 72.2 | 88.6 | 87.2 | 87.9 | 81.7 | 0.05 | 8.34 | \ | \ |
| His | 91.6 | 91.9 | 90.8 | 89.3 | 88.8 | 0.19 | 2.24 | \ | \ |
| Ile | 93.4 ^a | 92.3 ^{ab} | 91.2 ^{bc} | 90.6 ^c | 91.8 ^{bc} | 0.02 | 1.08 | 2.04*10 ⁻³ x ² -0.152x + 93.5 | 0.66 |
| Leu | 95.1 | 95.1 | 94.7 | 94.4 | 94.7 | 0.55 | 0.69 | \ | \ |
| Lys | 91.1 ^a | 91.7 ^a | 89.6 ^{ab} | 87.6 ^{bc} | 86.2 ^c | <0.001 | 1.52 | -0.0938x + 91.7 | 0.77 |
| Met | 92.4 ^a | 88.1 ^b | 87.3 ^b | 86.5 ^b | 88.1 ^b | <0.01 | 1.71 | 4.39*10 ⁻³ x ² - 0.322x + 91.8 | 0.73 |
| Phe | 94.2 | 94.0 | 93.4 | 93.3 | 94.0 | 0.57 | 0.99 | \ | \ |
| Thr | 89.9 ^a | 90.2 ^a | 88.5 ^{ab} | 86.9 ^b | 84.7 ^c | <0.001 | 1.27 | -0.0925x + 90.4 | 0.83 |
| Tyr | 93.7 | 93.3 | 92.5 | 92.5 | 92.9 | 0.27 | 0.93 | \ | \ |
| Val | 92.2 ^{ab} | 92.7 ^a | 91.7 ^{ab} | 90.7 ^b | 90.3 ^b | 0.011 | 0.94 | -0.0394x+92.5 | 0.61 |
| ∑EAA ² | 93.0 ^a | 93.0 ^a | 92.0 ^{ab} | 91.3 ^b | 91.2 ^b | 0.037 | 0.99 | -0.0338 x + 93.0 | 0.53 |
| ∑AA ² | 92.3 ^{ab} | 92.6 ^a | 91.5 ^{abc} | 90.7 ^{bc} | 90.1 ^c | <0.01 | 0.91 | -0.0420 x + 92.6 | 0.67 |
| | FMC | HSW10 | HSW20 | HSW40 | HSW60 | | | | |
| N | 86.4 | 85.5 | 87.9 | 86.3 | 85.9 | 0.40 | 1.95 | \ | \ |
| Energy | 84.1 | 84.2 | 87.2 | 84.8 | 83.8 | 0.11 | 1.95 | \ | \ |
| Arg | 95.5 ^{ab} | 96.0 ^a | 96.0 ^a | 94.8 ^b | 94.8 ^b | 0.02 | 0.60 | -0.0193x + 95.9 | 0.39 |
| Cys | 72.2 | 85.7 | 87.9 | 86.9 | 85.1 | 0.08 | 8.58 | \ | \ |
| His | 91.6 | 93.5 | 91.8 | 89.4 | 90.7 | 0.14 | 2.25 | \ | \ |
| Ile | 93.4 ^a | 92.4 ^a | 92.7 ^a | 90.7 ^b | 89.6 ^b | <0.001 | 0.81 | -0.0636x + 93.4 | 0.84 |
| Leu | 95.1 ^{ab} | 95.3 ^a | 95.5 ^a | 94.4 ^{bc} | 94.0 ^c | <0.01 | 0.51 | -0.0224x + 95.4 | 0.55 |
| Lys | 91.1 ^a | 91.8 ^a | 91.0 ^a | 87.9 ^b | 86.8 ^b | <0.01 | 1.72 | -0.0868x + 92.0 | 0.68 |
| Met | 92.4 ^a | 89.5 ^b | 88.4 ^b | 83.9 ^c | 85.8 ^c | <0.001 | 1.47 | 3.75*10 ⁻³ x ² - 0.344x + 92.7 | 0.86 |
| Phe | 94.2 | 94.0 | 94.1 | 93.0 | 93.2 | 0.20 | 0.90 | \ | \ |
| Thr | 89.9 ^a | 89.7 ^a | 89.8 ^a | 86.4 ^b | 84.7 ^b | <0.001 | 1.31 | -0.0947x + 90.6 | 0.80 |
| Tyr | 93.7 ^a | 93.2 ^a | 93.5 ^a | 91.9 ^b | 91.7 ^b | <0.01 | 0.72 | -0.0356x + 93.7 | 0.66 |
| Val | 92.2 ^a | 92.6 ^a | 93.0 ^a | 90.9 ^b | 89.4 ^c | <0.001 | 0.73 | -1.44*10 ⁻³ x ² + 0.0325x + 92.4 | 0.83 |
| ∑EAA ² | 93.0 ^a | 93.1 ^a | 93.1 ^a | 91.1 ^b | 90.8 ^b | <0.01 | 0.83 | -0.0456 x + 93.4 | 0.69 |
| ∑AA ² | 92.3 ^a | 92.9 ^a | 92.8 ^a | 90.5 ^b | 89.9 ^b | <0.01 | 0.88 | -0.0517 x + 93.0 | 0.67 |

26 For diet codes see Table 2. Different superscript letters ^a, ^b, and ^c indicate significant ($P < 0.05$) differences among treatments.

27 ¹ Pooled standard error of means. ² Trp excluded.

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Table 11 Factorial analysis of data on apparent digestibility coefficient (ADC, %) of hybrid grouper fed diets with different stickwater type and inclusion level

| Factor | N | Energy | Arg | Cys | His | Ile | Leu | Lys | Met | Phe | Thr | Tyr | Val | Σ EAA ¹ | Σ AA ¹ |
|---------------------------------|------|-------------------|-------------------|-------------------|--------------------|-------------------|-------------------|-------------------|--------------------|------|-------------------|--------------------|-------------------|---------------------------|--------------------------|
| Stickwater type | | | | | | | | | | | | | | | |
| CSW | 87.1 | 85.9 | 95.3 | 86.3 | 90.2 | 91.5 | 94.7 | 88.8 | 87.5 | 93.7 | 87.6 | 92.8 | 91.3 | 91.9 | 91.3 |
| HSW | 86.4 | 85.0 | 95.4 | 86.4 | 91.3 | 91.3 | 94.8 | 89.4 | 86.9 | 93.6 | 87.7 | 92.6 | 91.5 | 92.0 | 91.5 |
| Inclusion level, % | | | | | | | | | | | | | | | |
| 10 | 87.0 | 86.1 ^a | 96.0 ^a | 87.2 ^a | 92.7 ^a | 92.3 ^a | 95.2 ^a | 91.8 ^a | 88.8 ^a | 94.0 | 89.9 ^a | 93.2 ^a | 92.7 ^a | 93.1 ^a | 92.8 ^a |
| 20 | 87.7 | 87.2 ^a | 95.7 ^a | 87.5 ^a | 91.3 ^{ab} | 92.0 ^a | 95.1 ^a | 90.3 ^a | 87.8 ^{ab} | 93.8 | 89.1 ^a | 93.0 ^{ac} | 92.3 ^a | 92.5 ^a | 92.2 ^a |
| 40 | 86.1 | 84.7 ^b | 94.9 ^b | 87.4 ^a | 89.3 ^c | 90.6 ^b | 94.4 ^b | 87.8 ^b | 85.2 ^c | 93.1 | 86.7 ^b | 92.2 ^b | 90.8 ^b | 91.2 ^b | 90.6 ^b |
| 60 | 86.1 | 82.9 ^b | 94.8 ^b | 83.4 ^b | 89.7 ^{bc} | 90.7 ^b | 94.4 ^b | 86.5 ^b | 87.0 ^b | 93.6 | 84.7 ^c | 92.3 ^{bc} | 89.8 ^c | 91.0 ^b | 90.0 ^b |
| Pooled S.E.M. ² | 0.54 | 0.46 | 0.17 | 0.98 | 0.62 | 0.29 | 0.14 | 0.52 | 0.5 | 0.23 | 0.38 | 0.25 | 0.27 | 0.26 | 0.24 |
| Factorial ANOVA (P>F) | | | | | | | | | | | | | | | |
| Stickwater type | 0.23 | 0.06 | 0.45 | 0.94 | 0.08 | 0.58 | 0.67 | 0.25 | 0.26 | 0.74 | 0.82 | 0.44 | 0.66 | 0.59 | 0.32 |
| Inclusion level | 0.14 | <0.001 | <0.01 | 0.03 | <0.01 | <0.01 | <0.01 | <0.01 | <0.01 | 0.10 | <0.01 | 0.02 | <0.01 | <0.01 | <0.01 |
| Type * level | 0.13 | 0.02 | 0.28 | 0.18 | 0.75 | <0.01 | 0.011 | 0.80 | 0.020 | 0.17 | 0.34 | 0.037 | 0.05 | 0.20 | 0.14 |

Different superscript letters ^a, ^b, and ^c indicate significant ($P < 0.05$) differences among treatments.¹ Trp excluded.² Pooled standard error of means.30
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Declaration of interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests:

Author contributions

Yuexing Zhang and Lei Zhang contributed to the design of this study, performed most of experiments and statistical analysis, drafted, and revised the manuscript. Liying Huang designed the study, drafted & revised the manuscript, analyzed samples, and provided reagents and experimental facilities. Zhiyong Dong, Qiang Lu, and Yuan Zou participated in the conduction of feeding trial and analysis of experimental samples and data. Feng Tang, and Shuaibing Zhao participated in feed formulation, edited the manuscript, provided reagents, and analyzed samples. Trond Storebakken contributed to the design of this study, edited, and revised the manuscript. All the authors read and approved this version of the final manuscript and confirm the integrity of this work.

Data availability statement

The data that support the findings of this study and not presented in the figures and tables are available from the corresponding author on reasonable request.

Ethics statement

Hybrid grouper (*Epinephelus fuscoguttatus* ♀ × *E. lanceolatus* ♂) is a commercially farmed species in China not the protected species by Chinese law. During the feeding period and sampling procedures, the experimental fish were maintained in compliance with the Laboratory Animal Welfare Guidelines of China (Decree No. 2 of Ministry of Science and Technology, issued in 1988).