1	ENVIRONMENTALLY ASSOCIATED CHROMOSOMAL STRUCTURAL VARIATION
2	INFLUENCES FINE-SCALE POPULATION STRUCTURE OF ATLANTIC SALMON
3	(Salmo salar)
4	Running title: Environmentally mediated chromosomal translocation
5	
6	K. Beth Watson ^{1,2*} , Sarah J. Lehnert ² , Paul Bentzen ¹ , Tony Kess ² , Antony Einfeldt ¹ , Steven
7	Duffy ² , Sigbjørn Lien ³ , Matthew Kent ³ , Ben Perriman ¹ , and Ian R. Bradbury ^{1,2}
8	
9	1: Department of Biology, Dalhousie University, Halifax NS, Canada
10	2: Northwest Atlantic Fisheries Centre, Fisheries and Oceans Canada, St. John's NL, Canada
11	3: Centre for Integrative Genetics, Norwegian University of Life Sciences, Ås Norway
12	
13	*Correspondence: Beth Watson, Department of Biology Life Sciences Centre, Dalhousie
14	University, 1355 Oxford Street, PO Box 15000, Halifax NS, Canada, B3H 4R2
15	Tel: +1 (902-494-1398)
16	Email: beth.watson@dal.ca
17	
18	
19	
20	
21	

22 Abstract. Chromosomal rearrangements (e.g., inversions, fusions, and translocations) have long 23 been associated with environmental variation in wild populations. New genomic tools provide the 24 opportunity to examine the role of these structural variants in shaping adaptive differences within 25 and among wild populations of non-model organisms. In Atlantic Salmon (Salmo salar), 26 variations in chromosomal rearrangements exist across the species natural range, yet the role and 27 importance of these structural variants in maintaining adaptive differences among wild 28 populations remains poorly understood. We genotyped Atlantic Salmon (n = 1429) from 26 29 populations within a highly genetically structured region of southern Newfoundland, Canada with 30 a 220K SNP array. Multivariate analysis, across two independent years, consistently identified 31 variation in a structural variant (translocation between chromosomes Ssa01 and Ssa23), 32 previously associated with evidence of trans-Atlantic secondary contact, as the dominant factor 33 influencing population structure in the region. Redundancy analysis suggested that variation in 34 the Ssa01/Ssa23 chromosomal translocation is strongly correlated with temperature. Our analyses 35 suggest environmentally mediated selection acting on standing genetic variation in genomic 36 architecture introduced through secondary contact may underpin fine-scale local adaptation in 37 Placentia Bay, Newfoundland, Canada, a large and deep embayment, highlighting the importance 38 of chromosomal structural variation as a driver of contemporary adaptive divergence. 39

- 40

41 Key words: Chromosomal structural variation; translocation; secondary contact; local
42 adaptation; Atlantic Salmon

- 43
- 44

45 **1 INTRODUCTION**

46 Elucidating the genetic basis of adaptation is central to the understanding of evolutionary 47 biology (Schluter, 2009). Adaptive differences, accumulated through selection imposed by 48 spatially and temporally heterogenous environments, enable persistence (Felsenstein, 1976; 49 Savolainen, Lascoux, & Merilä, 2013) and drive the diversification of life (Dobzhansky, 1951). 50 Genome-scale data, now available for many non-model organisms, are highlighting the potential 51 of genomic architecture to facilitate adaptive divergence (Campbell et al., 2018; Wellenreuther et 52 al., 2019) with chromosomal structural variants (e.g., inversions, fusions, and translocations) 53 increasingly being identified and associated with environmental and life history variation (see 54 Wellenreuther & Bernatchez, 2018). However, understanding of (i) the origins of chromosomal 55 structural variants (Rougemont & Bernatchez, 2018; Fuller, Koury, Phadnis, & Schaeffer, 2019; 56 Marques, Meier, & SeeHausen, 2019), (ii) associations between structural variants and complex 57 phenotypes (Lee et al., 2017; Fuller et al., 2019; Jay et al., 2019), and (iii) the effectiveness of 58 different types of structural variants as drivers of adaptive divergence (Rieseberg, 2001; Guerrero 59 & Kirkpatrick, 2014) remains limited.

60 Chromosomal structural variants can be caused by changes in copy number (insertion, 61 deletion and duplication), orientation (inversion) or position (translocation and fusion). To date, 62 most work has focused on inversion polymorphisms (Dobigny, Britton-Davidian, & Robinson, 63 2017) due to the potential for strong suppression of recombination and reduced fertility in 64 heterozygous individuals (Sturtevant, 1917; White, 1978; King, 1993). Inversions are 65 taxonomically widespread (Kirkpatrick & Barton, 2006; Wellenreuther & Bernatchez, 2018), 66 frequently polymorphic within and between species and populations (Sturtevant, 1938; 67 Dobzhansky, 1951; White, 1978), and commonly align with environmental gradients (Balanyà,

68	Huey, Gilchrist, & Serra, 2009; Kennington & Hoffmann, 2013; Kapun, Fabian, Goudet, & Flatt,
69	2016), or complex morphological and behavioural phenotypes (Huang, Andrew, Owens, Ostevik,
70	& Rieseberg, 2019; Sinclair-Waters et al., 2018). Unlike inversions that change gene order
71	(Fuller et al., 2019), translocations and fusions physically unlink or link genes and reduce
72	recombination within each newly linked chromosome arm (Dumas & Britton-Davidian, 2002)
73	reducing recombination in both heterozygotes and rearranged homozygotes (Bidau, Giménez,
74	Palmer, & Searle, 2001; Castiglia & Capanna, 2002). The potential of translocations and fusions
75	to facilitate adaptation (Charlesworth, 1985) is supported by recent theoretical modeling work
76	(Guerrero & Kirkpatrick, 2014), and empirical evidence of environmental correlations with
77	fusions and translocations (Drosophila americana, McAllister, 2003; Dichroplus sp., Bidau,
78	Miño, Castillo, & Martí, 2012; Atlantic Salmon, Wellband et al., 2019). However, translocations
79	and fusions remain understudied with little reporting of the distribution or frequency of
80	polymorphisms, and associated phenotypic and environmental variation (Dobigny et al., 2017).
81	Atlantic Salmon (Salmo salar) span the North Atlantic Ocean exhibiting hierarchical
82	spatial structure across their range (King et al., 2007). Genomic differentiation is greatest
83	between continents with eastern (European) and western (North American) populations having
84	diverged more than 600,000 years before present (bp) (Nilsson et al., 2001; King et al., 2007;
85	Rougemont & Bernatchez, 2018). Over the intervening period numerous differences have
86	accumulated including large chromosomal rearrangements, two fusions (Ssa08/Ssa29 and
87	Ssa26/Ssa28) and a translocation (Ssa01/Ssa23) (Brenna-Hansen et al., 2012; Wellband et al.,
88	2019; Lehnert et al., 2019a), which have reduced the number of chromosome pairs from 29 in
89	Europe to 27 in North America (Hartley, 1988; Phillips & Ráb, 2001). Recent findings suggest
90	that variation in the Ssa01/Ssa23 chromosomal translocation exists in North America, and that

91 this variation was likely introduced through trans-Atlantic secondary contact from European 92 Atlantic Salmon near the end of the last glacial maximum (LGM) approximately 18,000 bp 93 (Bradbury et al., 2015; Rougemont & Bernatchez, 2018). Lehnert et al. (2019a) found that the 94 frequency of the Ssa01/Ssa23 translocation changed with latitude across Atlantic Canada with 95 more northern locations, such as sites in Newfoundland and Labrador, exhibiting the highest 96 frequency of the European karyotype (no Ssa01/Ssa23 translocation) relative to southern 97 populations which were almost entirely fixed for the North American karyotype (Ssa01/Ssa23 98 translocation). The importance of this structural variation resulting from secondary contact, and 99 the mechanisms acting to maintain high levels of polymorphism within and across Atlantic 100 Salmon populations in Atlantic Canada remain unknown.

101 Here we explore fine-scale spatial variation in southern Newfoundland, Canada, an area 102 with pronounced regional spatial structure (Bradbury et al., 2014), and evidence of trans-Atlantic 103 secondary contact (King et al., 2007; Bradbury et al., 2015). We first examine genomic variation 104 within Atlantic Salmon among 26 rivers using two discrete years of sampling and identify 105 variation in the Ssa01/Ssa23 translocation as a major driver of population structure. We examine 106 temporal stability of population structure as well as the frequency of this structural variation 107 within and among rivers. Next, we identify environmental associations with population structure 108 and consequently the Ssa01/Ssa23 translocation. We build directly on previous work that 109 identified range-wide polymorphism of a translocation between chromosomes Ssa01 and Ssa23 110 across Atlantic Canada (Lehnert et al., 2019a) and mitochondrial DNA evidence of trans-Atlantic 111 secondary contact along southeastern Newfoundland (King et al., 2007; Bradbury et al., 2015) 112 and highlight the potential role of chromosomal structural variation in fine-scale local adaptation

and the importance of secondary contact in generating standing genetic variation and drivingcontemporary adaptive divergence.

115

116 **2 METHODS**

117 **2.1 Sampling and genotyping**

118 Juvenile Atlantic Salmon, young-of-the-year (YOY) and parr (ages 0 to 2+), were 119 collected by electrofishing during the period July to September of 2017 and 2018 from 26 rivers 120 around Placentia Bay, a large (145 km wide at the mouth by 125 km long), deep bay (240 m) 121 separating the Avalon and Burin Peninsulas on the south coast of Newfoundland, Canada (Figure 122 1 and Table 1). Two rivers, Lance (LAN) and Little Barasway (LBB), were excluded from 123 sampling in 2018 due to small sample size in 2017. Cohorts were assigned based on age-length 124 relationships validated by scale ageing (Sylvester et al., 2019). Fin clips were collected and 125 preserved in 95% ethanol. DNA was extracted using DNeasy Blood and Tissue or DNeasy 96 126 Blood and Tissue kits (Oiagen, Toronto, ON, Canada) following manufacturer's protocols. 127 Concentration of extracted DNA was assessed using a Nanodrop spectrophotometer and by 128 agarose gel visualization. DNA was standardized to a concentration of 15 ng/ μ l. A total of 1429 129 (2017: 745 and 2018: 684) individuals were genotyped by Centre for Integrative Genetics 130 (CIGENE, Ås, Norway) using a 220K bi-allelic single nucleotide polymorphism (SNP) 131 Affymetrix Axiom array developed for Atlantic Salmon as described in Barson et al. (2015). 132 These SNPs were a subset of those in the 930K XHD *Ssa*l array (dbSNP accession numbers 133 ss1867919552–ss1868858426) designed using Norwegian aquaculture salmon. Genotype data 134 were filtered for high quality SNPs based on their clustering patterns and subsequent filtering was 135 performed using PLINK v 1.9 (Purcell et al., 2007; Chang et al., 2015). SNPs were filtered for a

minor allele frequency (MAF) cut-off of 0.01, a missingness threshold of 0.05 across samples,
and non-biallelic loci within each year sampled. SNPs were retained only if they passed filtering
in both years resulting in a total of 139,038 SNPs. In addition, samples were filtered to retain
only individuals with <5% missing genotypes, resulting in a total of 662 and 611 individuals in
2017 and 2018, respectively.

141

142 **2.2 Detection of population structure**

143 For population genetic analyses that require a panel of neutral and unlinked loci we first 144 used PLINK version 1.9 (Chang et al., 2015) to identify outlier loci ($F_{ST} > 95^{th}$ percentile) among 145 sample sites. Loci identified as outliers in both 2017 and 2018 were removed. We then removed 146 SNPs with high physical linkage using a sliding-window approach in PLINK version 1.9. SNPs 147 with a variance inflation factor (VIF) greater than 2 were removed from 50 SNP windows shifted 148 by five SNPs each iteration. We further removed all SNPs on chromosomes with known 149 structural variants Ssa01/Ssa23 and Ssa08/Ssa29 (Lehnert et al., 2019a). This neutral, unlinked 150 dataset was then thinned, keeping one SNP per 200,000 bases, using PLINK version 1.9 (Chang et 151 al., 2015).

Indices of genetic diversity including observed and expected heterozygosity (H_0 and H_e) and F_{1S} (Nei, 1987), were calculated per river and year to assess deviation from Hardy-Weinberg equilibrium. Indices were calculated using the *basic.stats* function in the R package *hierfstat* (Goudet, 2005) with the neutral, unlinked dataset (n = 6,302 SNPs). Confidence intervals for river-specific F_{1S} were calculated using the *boot.ppfis* function in *hierfstat* (Goudet, 2005) with 1,000 bootstrap replicates. Genetic differentiation was assessed by calculating pairwise F_{ST} (Weir and Cockerham, 1984) between rivers using the *stamp.fst* function, with 100 bootstrap replicates, in the R package *StAMPP* (Pembleton, Cogan, & Forster, 2013) and visualized using *gplots*(Warnes et al., 2016).

161 To estimate the number of distinct genetic clusters in each year, ADMIXTURE version 1.3 162 (Alexander, Novembre & Lange, 2009), which calculates individual ancestry proportions using 163 maximum likelihood estimates in a parametric model, was run for K (genetic clusters) 1 to 27 164 with three different random number seeds with the neutral, unlinked dataset (n = 6,302 SNPs). 165 From ADMIXTURE runs, standard deviation of cross-validation (CV) error was used to select a 166 reasonable range of K as in McCartney-Melstad, Vu & Shaffer (2018). Bar plots of estimated 167 individual ancestry proportions given by the Q-values were generated using R version 3.6.1 (R 168 Core Team, 2019).

169 The R package *pcadapt* (Luu et al., 2017) was then used to detect genomic regions 170 associated with population-based differences in genomic architecture across all 26 rivers in each 171 year. Multiple values of K (number of principal components; PCs), ranging from 1 to 100, were 172 explored. The R package *qvalue* (Storey et al., 2015) was used to correct for false-discovery rate 173 by transforming *p*-values for all SNPs into *q*-values which were plotted using the Manhattan plot 174 function in the R package *qqman* (Turner, 2014). The final number of PC axes retained was K =175 2, as we were primarily interested in large scale patterns of population differentiation. The 176 inclusion of additional values of K highlighted inter-individual differences rather than population 177 level differences. Further, upon visual inspection of the Manhattan plots, we found that two 178 strongly divergent genomic regions localized on chromosomes Ssa01 and Ssa23, a known 179 chromosomal translocation (Brenna-Hansen et al., 2012; Lehnert et al., 2019a), were the 180 dominant source of variation across multiple values of K (see Results). These regions associated 181 with the chromosomal translocation were important drivers of differentiation along the first PC

182 axis and were thus a primary focus of our study. We note that physical genomic positions 183 presented in our study are based on the European Atlantic Salmon genome, where Ssa01 and 184 Ssa23 are separate chromosomes (Lien et al., 2016), which differs from the standard North 185 American karyotype where the p arm of Ssa01 has fused to Ssa23 (Brenna-Hansen et al., 2012). 186

- 187

2.3 Environmental association analysis

188 Spatial patterns of association between genomic variation and climatic (i.e., temperature 189 and precipitation) and habitat (i.e., axial river length, basin relief, number of obstructions, and 190 human population density) variables were investigated using partial redundancy analysis (RDA) 191 as implemented in the R package vegan (Oksanen et al., 2017). Environmental variables were 192 collected from publicly available sources (see Table S1). GPS coordinates of sample sites were 193 used to extract 19 BIOCLIM variables, interpolated monthly climate data at a spatial resolution of 194 30 arc-seconds averaged for the years 1970 – 2000, from the WorldClim 2.0 database (Fick & 195 Hijmans, 2017). Human population density, a proxy for habitat disturbance, was calculated as in 196 Lehnert et al. (2019b). Axial river length, number of obstructions, and basin relief were obtained 197 from Porter et al. (1974). Axial river length and number of obstructions were approximated using 198 GOOGLE EARTH and maximum elevation from HydroSHEDS digital elevation model (Lehner, 199 Verdin, & Jarvis, 2008) was substituted for basin relief for missing sites (Fair Haven Brook, Red 200 Harbour East, and Piercey's Brook). All climatic and habitat data were standardized using the 201 scale function in R (R Core Team, 2019). 202 Given that many of the climatic variables were highly correlated (r > 0.8), we first 203 performed a variable reduction step using PCA to summarize climatic variation, for temperature

204 and precipitation. The first two PCs of each PCA were retained to reduce dimensionality and

205 covariance of loadings. Next, latitude and longitude of each river mouth were determined using 206 GOOGLE EARTH. Geographic distance between each river mouth was calculated using the least-207 cost distance function, constrained to a maximum depth of 100 m below sea-level, in the R 208 package marmap (Pante & Simon-Bouhet, 2013). Multivariate associations between genomic and 209 environmental data were tested using redundancy analysis (RDA), conditioned on geographic 210 distance from the most easterly site (Branch River), with four climatic summary variables (i.e. 211 PC1 and PC2 temperature, and PC1 and PC2 precipitation) and four habitat variables (i.e. basin 212 relief, number of obstructions, axial river length, and human population density) as predictors and 213 individual genotypes as dependent variables. Variance inflation factors were below 5 for all 214 variables indicating no multicollinearity between predictors. The RDA was visualized using the 215 plot function in R version 3.6.1 (R Core Team, 2019). The final number of RDA axes retained (n 216 = 3) was determined by visual inspection of the scree plot. To identify SNPs influenced by 217 climatic and habitat variation, we scaled and centred the raw scores on each constrained RDA 218 axis. We then identified outlier SNPs based on the loadings on each RDA axes, defined here as 219 SNPs more than three standard deviations from the mean. The Manhattan plot function in the R 220 package ggman (Rajagopal, 2020) was used to visualize each of the three retained RDA axes. 221 Visual inspection of the first RDA axis identified outlier SNPs localized on chromosomes Ssa01 222 and Ssa23, a known chromosomal translocation (Brenna-Hansen et al., 2012; Lehnert et al., 223 2019a). The correlation between frequency of the Ssa01/Ssa23 chromosomal translocation and 224 environmental variation was further explored in subsequent analyses. 225

226 2.4 Assignment of translocation karyotype

227	Variation in the Ssa01/Ssa23 chromosomal translocation was found to be both a major
228	source of population level differentiation and associated with environmental variation. Therefore,
229	we next examined variation in translocation frequency. Using the results of <i>pcadapt</i> , outlier SNPs
230	(q-value < 0.05 in both 2017 and 2018, $n = 887$) in the outlier block regions, Ssa01 (44,000,000 -
231	53,000,000 bp) and Ssa23 (0 - 9,500,000 bp), were combined as in Lehnert et al. (2019a). Spatial
232	genetic structure of the Ssa01/Ssa23 chromosomal translocation was explored with principal
233	component analyses (PCA) performed using the R package <i>pcadapt</i> (Luu et al., 2017) with $K = 3$.
234	Based on clustering patterns on the first PC axis, on which individuals were separated into three
235	clusters consistent with Lehnert et al. (2019a), individuals were assigned a karyotype using the
236	kmeans function in R version 3.6.1 (R Core Team, 2019). The three clusters corresponded to
237	three karyotypes: 1) standard North American (homozygous translocated; Ssa01p/Ssa23 and
238	Ssa01q), 2) standard European (homozygous non-translocated; Ssa01p/q and Ssa23), and 3)
239	heterozygous (carrying a translocated and non-translocated copy of the chromosomes).
240	Karyotype assignment followed Lehnert et al. (2019a), which incorporated European samples;
241	greater genetic variation was observed on PC1 and PC2 for the standard European karyotype
242	relative to the standard North American karyotype. This pattern of variation in genetic diversity
243	was consistent in our analysis (see Results). Using these genotype assignments, genetic
244	differentiation (observed heterozygosity) between karyotypes was assessed by calculating
245	pairwise F_{ST} (Weir and Cockerham, 1984) between groups in PLINK version 1.9 (Chang et al.,
246	2015).
247	Neighbor-joining (NJ) trees based on Nei's D (Nei, 1972) were generated using outlier

loci (q-value < 0.05) within the outlier block regions on Ssa01 and Ssa23 and the R package

248

249 StAMPP (Pembleton et al., 2013). Trees were visualized using FIGTREE v1.4 (Rambaut, 2012).

- Linkage disequilibrium (LD) was calculated among outlier SNPs (q < 0.05) on chromosomes
- 251 Ssa01 and Ssa23 between karyotypes. Pairwise LD (R^2) values were calculated using PLINK v 1.9
- 252 (Chang et al., 2015) and visualized using the R package *gplots* (Warnes et al., 2016).
- 253

254 **2.5 Frequency of translocation karyotype**

255 **Population variation and temporal stability**

Heterogeneity in translocation and karyotype frequencies between rivers was tested for each year sampled (2017 and 2018) using an analysis of deviance in a generalized linear model (GLM) with a binomial logistic transformation, followed by a comparison of contrasts as in

259 Mérot et al. (2018), and a pairwise Fisher's exact test adjusted for multiple comparisons.

260 Temporal stability of translocation and karyotype frequency within each river between years was

then tested using a pairwise Fisher's exact test. Two rivers, Lance (LAN), and Little Barasway

262 (LBB), were excluded from analysis due to limited sample size. To calculate translocation

- 263 frequency, we used the equation:
- 264 ((# homozygous translocated x 2) + (# heterozygotes)) / (# total individuals x 2)

265 which provides the frequency of the standard North American allele per river.

266

267 Environmental associations

Given the association between the Ssa01/Ssa23 chromosomal translocation and environmental variables (see RDA above), we next tested for significant correlations between the identified climatic variables (temperature and precipitation) and translocation frequency using linear regression. In the model, the response variable was the frequency of the North American

allele (Ssa01p/Ssa23) or the standard North American karyotype (homozygous translocated) and
the explanatory variable being the climatic summary variables (see Results).

274

275 **2.6 Gene ontology**

276 We examined functional enrichment of genes associated with the Ssa01/Ssa23 277 chromosomal translocation and identified as significant environmental outliers (q < 0.05) on 278 RDA1. We conducted gene ontology (GO) enrichment analysis using GO annotations in the 279 Atlantic Salmon genome from SalmoBase (Samy et al., 2020). A reference set of genes, genes 280 within 10 kb of all 138,451 SNPs from the array, was identified and extracted using BEDTOOLS 281 (Quinlan & Hall, 2010). Outlier sets of genes, genes within 10 Kb of outlier SNPs on RDA1 and 282 located within the translocation were then extracted for 2017 (n = 914) and 2018 (n = 700). The 283 R package topGO version 2.38.1 (Alexa, Rahnenführer, & Lengauer, 2006) was then used to test 284 for over-representation of GO biological processes using a node size of 5 and the 'weight01' 285 algorithm to account for structural relationships among GO terms. An alpha level of 0.01 was 286 used to determine significance. Using the same outlier sets of genes we then tested for 287 enrichment of gene profiles using the R package CLUSTERPROFILER (Yu, Wang, Han, & He, 288 2012). NCBI gene ID numbers were used as search criteria in the Kyoto Encyclopedia of Genes 289 and Genomes (KEGG) (Kanehisa, Goto, Sato, Furumichi, & Tanabe, 2012). An alpha level of 290 0.01 was used to determine significance.

291

3 RESULTS

293 **3.1 Sampling and genotyping**

294 In total, 662 individuals sampled in 2017 and 611 individuals sampled in 2018 were 295 genotyped and passed quality control thresholds. Exploratory analysis using principal component 296 analysis (PCA) found genetic variation separated Red Harbour East (RHA) from all other rivers 297 along the first two principal component (PC) axes in both 2017 and 2018 (Figure S1). Due to the 298 prevalence of non-anadromous Atlantic Salmon along the south coast of Newfoundland 299 (Verspoor, McGinnity, Bradbury, & Glebe, 2015) and the verification of a significant waterfall at 300 the mouth of RHA, this river was excluded following analyses of neutral genetic structure as 301 these individuals most likely represent a highly divergent landlocked population. A total of 302 138,451 SNPs, with a high overall genotyping rate (> 99%), and 632 individuals in 2017 and 585 303 individuals in 2018 were used in downstream analyses.

304

305 3.2 Detection of population structure

306 All rivers exhibited significant genetic differentiation (p < 0). Patterns of pairwise F_{ST} 307 clearly indicated strong regional structure within Placentia Bay consistent across both discrete 308 years sampled (Figure S2). Pairwise F_{ST} , which ranged from 0.0062 to 0.11 in 2017 and 0.0077 309 to 0.083 in 2018, was greatest between rivers along the Burin (Bay de l'Eau (BDL) – Piercey's 310 Brook (PBR)) and Avalon (Branch (BRA) – Ship Harbour (SHI)) Peninsulas indicative of an 311 east-west divide. Rivers along the Avalon Peninsula exhibited a higher degree of genetic 312 differentiation relative to each other as compared to rivers along the Burin Peninsula or head of 313 the bay where neighboring rivers exhibited little genetic differentiation. Interestingly, Cuslett 314 River (CUS) was found to be genetically similar to more geographically distant rivers along the 315 Burin Peninsula than neighboring rivers on the Avalon Peninsula. Observed (H_0) and expected 316 (H_e) heterozygosity ranged from 0.16 to 0.20 (mean = 0.19) in both 2017 and 2018. Inbreeding

317 coefficient (F_{IS}) ranged from -0.067 to -0.018 (mean = -0.026) in 2017 and -0.036 to 0.00 (mean 318 = -0.023) in 2018 (Table 2).

319	Fine-scale population structure was observed using a panel of putatively neutral loci
320	(6,302 SNPs with $F_{ST} < 0.05$ globally in both 2017 and 2018; known chromosomal
321	rearrangements excluded). The value of K with the lowest mean CV error in ADMIXTURE was $K =$
322	11 in 2017, and $K = 9$ in 2018. The reasonable range of K , values that had standard deviations
323	that overlapped with the lowest mean CV error were $K = 10$ and $12 - 14$ in 2017, and $K = 10 - 11$
324	in 2018 (Figure S3). Red Harbour East (RHA) appeared distinct across all values of K in both
325	years sampled. Although the majority of rivers formed river-specific clusters in both years
326	sampled, the head of the bay (CBC - SHA) appeared to form an admixed cluster, most similar to
327	rivers along southern Burin Peninsula, across all values of <i>K</i> in both 2017 and 2018 (Figure S4).
328	We also explored spatial structure within Placentia Bay by performing a principal
329	component analysis (PCA) on the full data set ($n = 138,451$ SNPs). This analysis similarly found
330	genetic variation separated populations by geographic region along the first two PC axes, with
331	PC1 (variance explained in 2017: 1.8% and 2018: 2.4%) highlighting an east-west divide within
332	Placentia Bay and PC2 (variance explained in 2017: 1.2% and 2018: 1.6%) a north-south divide
333	along the Avalon Peninsula (Figure 2a, b). This pattern of spatial structure was found to be
334	temporally stable with the exception of Northwest Mortier Bay (NMB), which in 2017 clustered
335	more closely with the Avalon Peninsula than the Burin Peninsula on which it is located.
336	Divergence among clusters identified by PC1 was found to be driven by large outlier block
337	regions (> 8 Mbp; Table S2) on both chromosomes Ssa01 and Ssa23 (Figure 2c, d). Each peak of
338	genetic differentiation (Ssa01 and Ssa23) contained > 400 SNPs that were statistical outliers ($q <$
339	0.05) in both years. An increasing number of PC axes tested ($K = 2 - 100$; see Supplementary

340 Information Figure S5) supported Ssa01 and Ssa23 as the dominant factor driving genomic 341 divergence among Atlantic Salmon within Placentia Bay. In a range-wide study, Lehnert et al. 342 (2019a) found genomic divergence driven by Ssa01 and Ssa23 indicative of polymorphism in a 343 known chromosomal translocation that differentiates European and North American salmon 344 (Brenna-Hansen et al., 2012). As in Lehnert et al., (2019a), outlier block regions on Ssa01 and 345 Ssa23 were combined and analyzed together in downstream analyses (see below).

- 346
- 347

3.3 Environmental association analysis

348 PCA-based reduction of climatic variables was used to construct summary variables for 349 temperature and precipitation. For the PCA of temperature variables, the first PC axis explained 350 47.1% of site environmental variation and was positively associated with temperature seasonality 351 (BIO4) and temperature annual range (BIO7) but negatively associated with minimum 352 temperature of the coldest month (BIO 6), while the second PC axis explained 2.0% of site 353 environmental variation was most strongly negatively associated with annual mean temperature 354 (BIO1) (Table S3). For the PCA of precipitation variables, the first PC axis of the summary 355 variable for precipitation explained 65.1% of site environmental variation and was most strongly 356 negatively associated with annual precipitation (BIO12), while the second PC axis explained 357 2.3% of site environmental variation and was strongly positively associated with precipitation 358 seasonality (BIO15).

359 In the RDA, total proportion of genetic variance explained by the constraining 360 (environmental) variables was 4.3% and 4.4% with 0.85% and 1.3% of the total proportion of 361 variance explained by the conditioning variable (distance from the most easterly site; BRA) in 362 2017 and 2018 respectively. We found temperature and precipitation explained the greatest

363 proportion of genetic variance on RDA1 based on vector length and number of significant SNPs. 364 A total of 422 and 222 SNPs were significantly associated with temperature PC1 and 114 and 365 440 SNPs were significantly associated with precipitation PC1 on RDA1 in 2017 and 2018 366 respectively. In both discrete years sampled, SNP loadings on RDA1 indicated a strong 367 association with the Ssa01/Ssa23 outlier block regions previously identified (Figure 3c, d), 368 suggesting an association between the translocation and environmental variation. Genetic 369 variance associated with temperature was most strongly driven by rivers at the head of Placentia 370 Bay and along the Burin Peninsula (Figure 3a, b), a pattern that was consistent across years. 371 Whereas genetic variance associated with precipitation was most strongly driven by rivers along 372 southern Burin and Avalon Peninsulas (Figure 3a, b).

- 373
- 374 **3.4** Assignment of translocation karyotype

375 Given that variation in the Ssa01/Ssa23 chromosomal translocation was identified as a 376 major source of genetic structure, and found to be associated with environmental variation, we 377 next examined translocation and karyotype frequency. Analysis of outlier SNPs (n = 887) within 378 the outlier block regions on Ssa01/Ssa23 found three distinct clusters on PC1 (variance explained 379 in 2017: 54.3% and 2018: 53.6%) (Figure 4a, b), consistent with a chromosomal rearrangement. 380 Similar clustering patterns were found using neighbor-joining (NJ) trees (Figure 4c, d). 381 Karyotype was assigned to each of the three clusters (based on PCA) with individuals assigned as 382 either: 1) standard North American (homozygous translocated; Ssa01p/Ssa23 and Ssa01q), 2) 383 standard European (homozygous non-translocated; Ssa01p/q and Ssa23), and 3) heterozygous 384 (carrying a translocated and non-translocated copy of the chromosomes). The heterokaryotype 385 was found to be intermediate to the homokaryotypes along PC1 (Figure 4). Individuals from

386 throughout Placentia Bay were found in each of the three clusters suggesting karyotype clusters 387 were not completely driven by the geography of the bay. Interestingly, the cluster of individuals 388 found to have the standard European karyotype (homozygous non-translocated; Ssa01p/q and 389 Ssa23) exhibited greater genetic variation along the first two PC axes than the clusters of 390 heterozygous or standard North American karyotype (homozygous translocated; (Ssa01p/Ssa23 391 and Ssa01q) individuals which exhibited the least amount of genetic variation along the first two 392 PC axes (Figure 4a, b). 393 Genetic differentiation (F_{sT}) between homokaryotypes was significantly greater (p < 0.001) 394 within the Ssa01/Ssa23 outlier block regions (Ssa01: $F_{ST} = 0.55$ and $F_{ST} = 0.52$, and Ssa23: $F_{ST} =$ 0.71 and $F_{\rm ST} = 0.70$ in 2017 and 2018 respectively) relative to that observed genome wide ($F_{\rm ST} =$ 395 396 0. 0094 and $F_{sT} = 0.0096$ in 2017 and 2018 respectively). Heatmaps of linkage disequilibrium 397 (LD) between outlier SNPs (q < 0.05) on chromosomes Ssa01 and Ssa23 revealed regions of high 398 LD, in all pairwise comparisons of translocation karyotype (Figure S6). As expected, linkage 399 disequilibrium (R^2) was highest between homokaryotypes. Interestingly, LD between the 400 heterokaryotype and standard European karyotype was found to be lower than that observed 401 between the heterokaryotype and standard North American karyotype. Heterozygosity was found 402 to be four times higher for the standard European karyotype relative to the standard North 403 American karyotype within the outlier block regions (Figure S7).

- 404
- 405 **3.5 Frequency of translocation karyotype**

406 Population variation and temporal stability

407 All rivers were polymorphic for the Ssa01 and Ssa23 chromosomal translocation (Figure 408 S8). Frequency of the translocation did not significantly differ between years (p = 0.263) but did

409 differ significantly between rivers within years (p < 0.01) (Figure S9). Average translocation 410 frequency (standard North American 'allele') was 61.3% (range: 5.5 – 86.0%) in 2017 and 59.6% 411 (range: 6.7 - 94.5%) in 2018. Frequency of the standard North American karyotype (homozygous 412 translocated; Ssa01p/Ssa23 and Ssa01q) was 41.9% (range: 0 - 72.0%) in 2017 and 41.0%413 (range: 0 - 89.9%) in 2018 (Table S4). Frequency of the translocation was temporally stable 414 within rivers across the two discrete years sampled with the exception of Northwest Mortier Bay 415 (NMB), Cuslett (CUS), and Big Salmonier (BSA) which differed significantly in both karyotype 416 and translocation frequency between 2017 and 2018 (Table S5). Translocation frequency 417 significantly increased in NMB and significantly decreased in CUS and BSA from 2017 to 2018 418 (Figure S10).

419

420 Environmental associations

421 The translocation appeared spatially distributed along a longitudinal gradient with the 422 highest frequency of the standard North American karyotype found along central Burin Peninsula 423 and the head of Placentia Bay, and absent or occurring at low frequency along the Avalon 424 Peninsula (Figure 5). A pronounced transition in frequency of the translocation homokaryotype, 425 consistent across both years sampled, was observed between Ship Harbour Brook (SHI) and Fair 426 Haven Brook (FHB). SHI and FHB had an average of 2.3% and 42.7% standard North American 427 karyotype (homozygous translocated) individuals across years, respectively, suggesting an 428 increase in European ancestry (in this genomic region) between sites. Interestingly, variation in 429 translocation frequency between rivers was significantly correlated with temperature PC1 (2017: 430 p = 0.067 and 2018: p = 0.0087) (Figure 6) but not precipitation PC1 (Figure S11). The 431 translocation (standard North American allele) occurred more frequently in rivers that have a

lower minimum temperature in the coldest month and exhibited greater variability in temperatureboth seasonally and annually.

434

435 **3.6 Gene Ontology**

436 We searched the Salmo salar genome for annotated genes within 10kb on either side of 437 each SNP within the outlier block regions on Ssa01 and Ssa23 identified as an environmentally 438 associated outlier on RDA1. We identified 260 and 241 unique genes in proximity to the 914 and 439 700 SNPs identified as RDA1 outliers within the Ssa01/Ssa23 chromosomal translocation in 440 2017 and 2018 respectively. These genes represent putative targets of selection. We found 441 functional enrichment of 26 and 35 gene ontology (GO) biological processes (p < 0.01, Table S6) 442 in 2017 and 2018 respectively. Of these, 21 GO biological processes were significantly enriched 443 (p < 0.01) in both 2017 and 2018. Of particular interest were processes related to immunity, 444 growth, and oxidative stress. Further, using the outlier SNPs identified above, enrichment tests in 445 KEGG indicated over-representation of the insulin signaling pathway in both 2017 (p = 0.00763) 446 and 2018 (p = 0.00439).

447

448 **4 DISCUSSION**

Chromosomal structural variation is a significant, yet poorly understood source of genetic variation (Wellenreuther et al., 2019) which may underpin complex phenotype and life history variation across a wide range of taxa (Dobigny et al., 2017). In contrast with chromosomal inversions, which have been increasingly associated with adaptive variation (Lamichhaney et al., 2016; Jay et al., 2018; Huang et al., 2019), few well-documented occurrences of fusion or translocation polymorphisms in wild populations have been reported (see Bidau & Martí, 2002;

455 Dobigny et al., 2017; Wellband et al., 2019; Cayuela et al., 2020). Our study is among the first to 456 report evidence of an adaptive chromosomal translocation influencing spatial structure in the 457 wild. We found the Ssa01/Ssa23 chromosomal translocation, previously found to reflect trans-458 Atlantic differences and secondary contact (Brenna-Hansen et al., 2012; Lehnert et al., 2019a), to 459 be polymorphic and associated with fine-scale spatial structure of Atlantic Salmon in Placentia 460 Bay, Newfoundland, Canada. Moreover, we found translocation frequency is significantly 461 correlated with environmental variation in the region. This work extends previous analyses 462 (Bradbury et al., 2015; Lehnert et al., 2019a) providing a high-resolution examination of trans-463 Atlantic secondary contact in Atlantic Salmon in southern Newfoundland, and highlights the 464 importance of secondary contact, introgression, and chromosomal structural variation as drivers 465 of adaptive divergence.

466

467 **4.1 Chromosomal translocation drives fine-scale spatial structure**

468 In our study, Atlantic Salmon populations exhibited hierarchical spatial genetic structure 469 within Placentia Bay, with the greatest genetic differentiation occurring between the Avalon and 470 Burin Peninsulas. This is consistent with previous work by Bradbury et al. (2015) which 471 identified two discrete genetic clusters along southern Newfoundland, east and west, with a 472 boundary near the Burin Peninsula. Trans-Atlantic secondary contact has been suggested to be a 473 significant factor structuring this region. Gene flow following secondary contact has been 474 documented for many temperate species that experienced periods of range expansion and 475 contraction throughout the Quaternary (Hewitt, 2000; Tigano & Friesen, 2016). In Atlantic 476 Salmon, secondary contact between European and North American salmon occurred, most 477 recently, during the Pleistocene at the end of the last glacial maximum (King et al., 2007;

Rougemont & Bernatchez, 2018) and is supported by evidence of European mitochondrial DNA 478 479 (King et al., 2007; Bradbury et al., 2015), and recently the identification of a chromosomal 480 polymorphism associated with European ancestry in northern Canada (Lehnert et al., 2019a). 481 The Ssa01/Ssa23 chromosomal translocation, previously associated with introgression 482 from European Atlantic Salmon into northern Canada (Lehnert et al. 2019a), was found to be 483 polymorphic within Placentia Bay and appeared to be a significant factor contributing to genetic 484 structuring in this region. Genetic variation of outlier loci within the outlier block regions 485 identified on chromosomes Ssa01 and Ssa23 showed three distinct clusters, a pattern observed by 486 Lehnert et al. (2019a) and consistent with a chromosomal rearrangement. Karyotype frequencies 487 showed a longitudinal clinal pattern. Populations on the Avalon Peninsula were predominately 488 composed of individuals with the standard European karyotype (homozygous non-translocated; 489 Ssa01p/q and Ssa23) whereas populations at the head of the bay and on the Burin Peninsula were 490 predominately composed of individuals with the standard North American karyotype 491 (homozygous translocated; Ssa01g and Ssa01p/Ssa23). Interestingly, Lehnert et al. (2019a) found 492 no individuals with the standard European karyotype south of Labrador, although sampling was 493 limited to only four populations in Newfoundland located west of the Burin Peninsula. While 494 long read nanopore sequencing is required to confirm the presence of a translocation in these 495 populations, this finding strongly supports a hypothesis of trans-Atlantic secondary contact in 496 Placentia Bay.

497 Observed patterns of genetic diversity indicated the standard North American karyotype 498 has reduced genetic diversity relative to the standard European karyotype. Furthermore, pairwise 499 comparisons of linkage disequilibrium between karyotypes suggested greater suppression of 500 recombination between the standard North American karyotype and heterokaryotype than

501	between the standard European karyotype and heterokaryotype. This finding is consistent with
502	reports of reduced frequency of recombination and a shift in the distribution of recombination
503	towards the distal ends of chromosomes in fused homokaryotypes and, to a lesser extent,
504	heterokaryotypes (Bidau et al., 2001; Castiglia & Capanna, 2002; Dumas & Britton-Davidian,
505	2002; Guerrero, & Kirkpatrick, 2014). While North American salmon generally have low levels
506	of European ancestry (~ 3% genome-wide), individuals with the standard European karyotype
507	have been reported to have high levels of European ancestry (> 50%) within the outlier block
508	regions on Ssa01 and Ssa23 (Lehnert et al., 2019a). Given that the SNP array was developed
509	using European Atlantic Salmon (Barson et al., 2015), it is possible inferences about diversity
510	may be influenced by ascertainment bias. However, the dataset used here was based on a subset
511	of polymorphic loci and using a similar array and methodology Bradbury et al. (2015) concluded
512	the observed pattern in genetic diversity along southern Newfoundland was the result of
513	historical processes with minimal influence of ascertainment bias.
514	Interestingly, the outlier block region identified on chromosome Ssa23 $(0 - 9.5 \text{ Mbp})$ was
515	larger in size than that reported by Lehnert et al. (2019a) perhaps suggesting multiple secondary
516	contact events may have occurred during the colonization of North America following the last
517	glacial maximum, a hypothesis for which Rougemont & Bernatchez (2018) found some support.
518	Moreover, the larger outlier blocks found here may suggest a more recent secondary contact
519	event in Newfoundland compared to northern regions (Labrador). This highlights the need for
520	future studies on the demographic and evolutionary history of Atlantic Salmon in Canada. While
521	other studies have identified evidence that secondary contact influences differentiation range-
522	wide (Lehnert et al., 2019a), we found variation in the Ssa01/Ssa23 chromosomal translocation is
523	significantly associated with genome-wide population structure demonstrating the importance of

524 the Ssa01/Ssa23 chromosomal translocation to fine-scale structuring within Placentia Bay and 525 suggesting the translocation may influence gene flow through incompatibilities or adaptive 526 differences between karyotypes.

527

528 **4.2** Environment correlated with chromosomal translocation

529 Geographic regions of post-glacial secondary contact can provide opportunities to 530 investigate the evolution and maintenance of chromosomal structural variation and its role in 531 adaptive divergence (Tigano & Friesen, 2016; Lee et al., 2017). Secondary contact events can 532 generate polymorphism in standing variation, which can then be selected upon in a heterogenous 533 environment (Alcala & Vuilleumier, 2014; Marques et al., 2019). Genomic architecture is an 534 underappreciated source of variation on which selection can act (Wellenreuther et al., 2019). 535 Chromosomal structural variants may be adaptive due to spatially and/or temporally varying 536 selection on: i) breakpoints or position effects which cause gene disruption or alter expression 537 (Corbett-Detig, 2016; Puerma, Orengo, & Aguadé, 2016), ii) recombination rate (McDonald, 538 Rice, & Desai, 2016), or iii) alleles captured or accumulated (Wright, 1931; Coyne & Orr, 2004; 539 Fuller et al., 2019). As such, chromosomal structural variants are expected to clearly delineate the 540 genetic boundaries between parapatric populations that straddle an ecotone (Slatkin, 1975; 541 Kirkpatrick & Barton, 2006). Using a fine-spatial scale approach we identified a cline in 542 translocation karyotype that aligned with an environmental gradient consistent with a hypothesis 543 of adaptive significance. 544 Placentia Bay, a long (125 km) and deep (240 m) embayment in southeastern

545 Newfoundland, spans 145 km at the mouth and narrows towards the head of the bay where

546 summer temperatures are warmer relative to the mouth of the bay (Fisheries and Oceans Canada,

547 2007). Genotype-environment analysis (redundancy analysis; RDA) indicated temperature range 548 and seasonality best explained the observed spatial genetic structure and highlighted a strong 549 association with the Ssa01/Ssa23 chromosomal translocation. Although the use of air temperature 550 as a proxy for freshwater temperature may not be accurate, particularly across small spatial scales 551 (Hansen, Read, Hansen, & Winslow, 2016), previous work in the region has reported a strong 552 correspondence between air and water temperatures (Bradbury et al., 2014). Furthermore, a 553 significant correlation between translocation frequency and temperature was found, indicating, 554 that like inversions, translocations have the potential to be adaptive, a finding supported by 555 evidence of signatures of positive selection acting on both the standard North American and 556 standard European karyotypes (Lehnert et al., 2019a). Taken together, these findings suggest the 557 standard North American karyotype is adaptive in North America, however, within secondary 558 contact zone(s) where introgression has occurred, the standard European karyotype also confers a 559 locally adaptive advantage, presumably by adding to standing variation on which selection acts 560 within a heterogenous landscape over a fine-spatial scale. Alternatively, temperature-aligned 561 population structure could be the result of demographic history. Chromosomal structural variants 562 may have been pre-adapted to differing temperature regimes and aligned themselves along the 563 currently observed cline in temperature during post-glacial colonization; consistent with a 564 hypothesis of niche coupling (Knowles, Carstens, & Keat, 2007). Regardless of the influence of 565 colonization history, a significant correlation between contemporary fine-scale population 566 structure and temperature was observed.

Local adaptation has been recognized as an important evolutionary process in salmonids
(reviewed in Taylor, 1991; Garcia de Leaniz et al., 2007) with temperature often identified as the
dominant factor structuring populations (Larson, Lisi, Seeb, Seeb, & Schindler, 2016).

570	Temperature has been shown to directly influence metabolic and growth rate (Burgerhout et al.,
571	2017; Stehfest et al., 2017; Vikeså, Nankervis, & Hevrøy, 2017), age at smoltification and
572	maturation (Mangel, 1994; Minns et al., 1995; Friedland, 1998), proportion of precocial parr in
573	Atlantic Salmon populations (Valiente et al., 2005; Yates et al. 2015), migration timing (Jonsson
574	& Jonsson, 2018), and fitness and survival through oxidative stress (Birnie-Gauvin, Costantini,
575	Cooke, & Willmore, 2017). While some evidence suggests a difference in proportion of precocial
576	males (Dalley, Andrew, & Green, 1983) between the mouth and head of Placentia Bay, little is
577	known about the life history and ecology of salmon at this spatial scale in this region.
578	While our results suggest an association between translocation frequency and
579	temperature, we acknowledge that other unmeasured variables that covary with temperature may
580	contribute to the genetic structure observed (Storfer et al., 2006). Inclusion of additional
581	environmental parameters, such as pH and/or geological characteristics (Bourret et al., 2013;
582	Bradbury et al., 2014), and pathogen or parasite diversity (Dionne, Miller, Dodson, &
583	Bernatchez, 2009), may provide further insight into the mechanisms influencing spatial structure
584	in the region. Although, notably, in regions such as Labrador, previous work has found a higher
585	frequency of the standard European karyotype within populations within a large marine
586	embayment (Lake Meville) where temperatures are warmer compared to coastal populations with
587	a lower frequency of the standard European karyotype (Sylvester et al., 2018; Lehnert et al.,
588	2019a). While this association has not been formally tested, it highlights another region in North
589	America where polymorphism of the Ssa01/Ssa23 chromosomal translocation may align with
590	clinal variation in temperature. Although these relationships appear to operate in different
591	directions, this may further support the role of multiple secondary contact events in parts of
592	Canada from different regions of Europe.

594

4.3 Chromosomal translocation exhibits temporal stability across ecotone

595 Translocation frequency was found to be temporally stable across the two discrete years 596 sampled. A pronounced transition in translocation frequency was observed between Ship Harbour 597 (SHI) and Fair Haven Brook (FHB) on the Avalon Peninsula, however, this transition was less 598 apparent on the Burin Peninsula, where Northwest Mortier (NMB) and Big Salmonier (BSA), 599 rivers located in an intermediary zone of clinal variation, were found to significantly differ in 600 translocation frequency between 2017 and 2018. While maintenance of polymorphism and 601 temporal stability suggest the Ssa01/Ssa23 chromosomal translocation is adaptive, it is plausible 602 that the Ssa01/Ssa23 chromosomal translocation is neutral or near-neutral and polymorphism 603 persists due to demographic factors such as low gene flow and low effective population size in 604 region (Palstra et al., 2007; Bradbury et al., 2015) and drift. Alternatively, polymorphism of the 605 Ssa01/Ssa23 chromosomal translocation may be maintained, not because the variants represent 606 adaptations to divergent habitats, but because the homozygous translocated karvotype carries 607 harmful recessive mutations as suggested by Jay et al. (2019) in *Heliconious numata*. Future 608 work should examine transposable element (TE) dynamics and the rate of non-synonymous to 609 synonymous substitution (dN/dS) within the translocated region.

610

611 **4.4 Functional significance and gene annotations**

We identified biological processes which were over-represented using genes within the outlier block regions on Ssa01 and Ssa23 that were located near environmentally associated SNPs. KEGG pathway analysis found enrichment of the insulin signalling pathway in both 2017 and 2018. In addition, analysis of gene ontology found over-representation of biological 616 processes primarily related to regulation of metabolic processes, and immune response. The

617 insulin signalling pathway may relate to both metabolic processes (Babbitt, Warner, Fedrigo,

618 Wall, & Wray, 2010; Zhang et al. 2018) and immunity (Yada & Tort, 2016; Cheng et al., 2017;

619 Wang et al., 2019).

620 In salmon, genes and/or biological processes related to metabolic processes and immunity 621 can be influenced by environmental factors, such as temperature (Dionne, Miller, Dodson, Caron, 622 & Bernatchez, 2007; Beauregard et al., 2013). Metabolic differences in salmon populations under 623 different temperature regimes have been reported; salmon in warmer conditions grow faster and 624 migrate to sea at a younger age (Power, 1981; Metcalfe & Thorpe, 1990) while salmon in cooler 625 environments exhibit higher growth rate and more efficient metabolic processes (Nicieza et al., 626 1994). In Atlantic Salmon, precocial male parr are common in Newfoundland (Dalley et al. 627 1983), and their occurrence can be influenced by temperature (Valiente et al., 2005; Yates et al. 628 2015).

629 Temperature regime has also been correlated with bacterial diversity and as such genetic 630 diversity for immune-related genes (Dionne et al., 2007, 2008) and is thought to be involved in 631 local adaptation to different pathogen communities (Bourret et al., 2013). Parasites have played a 632 major role in mortality of wild fish in Newfoundland (Khan, 2009) with outbreaks and mortality 633 of proliferative kidney disease (PKD) being seasonal and temperature dependent in salmonids 634 (Sterud et al., 2007). Overall, these processes point to potential adaptive associations with 635 temperature but remain speculative, and experimental work is needed to better understand these 636 relationships.

637

638 **4.5 Conservation and management implications**

639 Atlantic Salmon in southern Newfoundland have undergone significant declines in 640 abundance over the last few decades (Chaput et al., 2012; Lehnert et al., 2019b) and are currently 641 managed as a single designatable or evolutionarily significant unit (COSEWIC, 2010). Our 642 results clearly demonstrate two genetic clusters and provide strong evidence that an adaptive 643 chromosomal translocation associated with trans-Atlantic secondary contact drives fine-scale 644 population structure in the region. Placentia Bay is of particular interest in that it is a geographic 645 region of post-glacial secondary contact and as such provides an opportunity to investigate the 646 evolution and maintenance of chromosomal structural variation and its role in adaptive 647 divergence because evolutionary dynamics of chromosomal structural variants differ from other 648 parts of the genome, (Tigano & Friesen, 2016; Lee et al., 2017; Wellenreuther & Bernatchez, 649 2018; Mérot, Oomen, Tigano, & Wellenreuther, 2020). While polymorphism in chromosomal 650 structural variation, such as the Ssa01/Ssa23 chromosomal translocation characterized here, may 651 complicate the process of delineating evolutionarily significant units, particularly when the 652 relative fitness consequences are unclear, it can be important to consider genomic architecture 653 underlying adaptive phenotype or life history variation when predicting the consequences of 654 environmental disturbance and climatic change (Oomen, Kuparinen, & Hutchings, 2020).

- 655
- 656

657 **5 CONCLUSIONS**

Species and populations adapt through selection imposed by spatially and temporally
heterogenous environments on new mutation or standing genetic variation (Wright, 1931; Coyne
& Orr, 2004). Gene flow is an important source of standing genetic variation (Tigano & Friesen,
2016) promoting adaptation through the re-introduction of previously lost variation (Rieseberg,

662 2009) and the introgression of novel genetic variants and allelic combinations among meta-663 populations (Poelstra, Richards, & Martin, 2018). The Ssa01/Ssa23 chromosomal translocation 664 (Brenna-Hansen et al., 2012) has recently been found to be polymorphic within secondary contact 665 zones in North America (Lehnert et al., 2019a). This study further supports secondary contact 666 with European introgression into Atlantic Salmon populations along southeastern Newfoundland 667 and highlights the importance of secondary contact in shaping population genetic structure. 668 Effects of chromosomal structural variants are expected to be most pronounced when fixed in 669 populations prior to secondary contact, with subsequent reproductive isolation maintained by 670 adaptive change involving many genes with small fitness effects (Feder, Nosil, & Flaxman, 671 2014). Here, we found evidence of an adaptive chromosomal rearrangement and a cline in 672 translocation frequency aligned with a cline in temperature. These findings suggest the standard 673 North American karyotype is broadly adaptive in North America, however, within secondary 674 contact zone(s) where introgression of the standard European karyotype has occurred, the 675 standard European karyotype also confers an adaptive advantage in local populations. Future 676 work should explore the roles of demography and drift, monitor clinal stability of translocation 677 frequency over an extended period of time to investigate the evolution and maintenance of 678 putatively adaptive translocation in the wild, and use direct temperature measurements and 679 common-garden experiments to investigate differential gene expression between 680 homokaryotypes. This study highlights the importance of chromosomal structural variation as a 681 source of standing variation on which selection can act. 682

683

684

685	ACKNOWLEDGEMENTS	(ensure all relevant	grant numbers are listed	1)
-----	------------------	----------------------	--------------------------	------------

686	The authors thank the staff of the Newfoundland DFO Salmonids section for juvenile sampling,
687	NRC for use of the plate reader, the Aquatic Biotechnology Laboratory of the Bedford Institute
688	of Oceanography for sample preparation, CIGENE for SNP genotyping and data processing, and
689	C. Mérot for providing R scripts and guidance on GLM analyses. This study was funded through
690	the Program for Aquaculture Regulatory Research of Fisheries and Oceans Canada, and an
691	NSERC Discovery Grant to IRB.
692	
693	
694	
695	
696	
697	
698	
699	
700	
701	
702	
703	
704	
705	
706	
707	

708 **REFERENCES**

709 Alcala, N., & Vuilleumier, S. (2014). Turnover and accumulation of genetic diversity across large 710 time-scale cycles of isolation and connection of populations. *Proceedings of the Royal Society B*, 711 **281**(1704), https://doi.org/10.1098/rspb.2014.1369. 712 713 Alexa, A., Rahnenführer, J., & Lengauer, T. (2006). Improved scoring of functional groups from 714 gene expression data by decorrelating GO graph structure. *Bioinformatics*, 22(13), 1600-1607. 715 716 Alexander, D. H., Novmbre, J., & Lange, K. (2009). Fast model-based estimation of ancestry in 717 unrelated individuals. Genome Research, 19, 1655 – 1664. 718 719 Babbitt, C. C., Warner, L. R., Fedrigo, O., Wall, C. E., & Wray, G. A. (2010). Genomic 720 signatures of diet-related shifts during human origins. Proceedings of the Royal Society B, 721 278(1708), https://doi.org/10.1098.rspb/2010.2433. 722 723 Balanyà, J., Huey, R. B., Gilchrist, G. W., & Serra, L. (2009). The chromosomal polymorphism 724 of Drosophila subobscura: a microevolutionary weapon to monitor global change. Heredity. 103, 725 364 – 367. 726 727 Barson, N. J., Aykanat, T., Hindar, K., Baranski, M., Bolstad, G. H., Fiske, P., ... & Primmer, C. 728 R. (2015). Sex-dependent dominance at a single locus maintains variation in age at maturity in 729 salmon. Nature, 528, 405-408. 730 731 Beauregard, D., Enders, E., & Boisclair, D. (2013). Consequences of circadian fluctuations in 732 water temperature on the standard metabolic rate of Atlantic salmon parr (Salmo salar). 733 *Canadian Journal of Fisheries and Aquatic Sciences*, **70**(7), 1072 – 1081. 734 735 Bidau, C. J., Giménez, M. D., Palmer, C. L., & Searle, J. B. (2001). The effects of Robertsonian 736 fusion on chiasma frequency and distribution in the house mouse (*Mus musculus domesticus*) 737 from a hybrid zone in northern Scotland. Heredity, 87, 305 – 313. 738 739 Bidau, C. J., & Martí, D. A. (2002). Geographic distribution of Robertsonian fusions in 740 Dichroplus pratensis, (Melanoplinae, Acrididae): the central-marginal hypothesis reanalysed. 741 *Cytogenetic and Genome Research*, **96**, 66 – 74. 742 743 Bidau, C. J., Miño, C. I., Castillo, E. R., & Martí, D. A. (2012). Effects of abiotic factors on the 744 geographic distribution of body size variation and chromosomal polymorphisms in two 745 neotropical grasshopper species (Dichroplus: Melanoplinae: Acrididae). Psyche, 746 https://doi.org/10.1155/2012/863947. 747 748 Birnie-Gauvin, K., Costantini, D., Cooke, S. J., & Willmore, W. G. (2017). A comparative and 749 evolutionary approach to oxidative stress in fish : a review. Fish and Fisheries, 18(5), 928 - 942. 750 https://doi.org/10.1111/faf.12215.

752 Bourret, V., Dionne, M., Kent, M. P., Lien, S., & Bernatchez, L. (2013). Landscape genomics in 753 Atlantic salmon (Salmo salar) searching for gene-environment interactions driving local 754 adaptation. *Evolution*, **67**(12), 3469 – 3487. 755 756 Bradbury, I. R., Hamilton, L. C., Robertson, M. J., Bourgeois, E., Mansour, A., & Dempson, B., 757 (2014). Landscape structure and climatic variation determine Atlantic salmon genetic 758 connectivity in the Northwest Atlantic. Canadian Journal of Fisheries and Aquatic Sciences, 759 71(2), 246 - 258.760 761 Bradbury, I. R., Hamilton, L. C., Dempson, B., Robertson, M. J., Bourret, V., Bernatchez, L., & 762 Verspoor, E. (2015). Transatlantic secondary contact in Atlantic salmon, comparing 763 microsatellites, a single nucleotide polymorphism array and restriction-site associated DNA 764 sequencing for the resolution of complex spatial structure. *Molecular Ecology*, 24(20), 5130 -765 5144. 766 767 Brenna-Hansen, S., Li, J., Kent, M. P., Boulding, E. G., Dominik, S., Davidson, W. S. & Lien, S. 768 (2012). Chromosomal differences between European and North American Atlantic salmon 769 discovered by linkage mapping and supported by fluorescence *in situ* hybridization analysis. 770 BMC Genomics, 13, 432. 771 772 Burgerhout, E. (2017). Genetic background and embryonic temperature affect DNA methylation 773 and expression of *myogenin* and muscle development in Atlantic salmon (Salmo salar). *PLoS* 774 *One*, **12**(6): e0179918. 775 776 Campbell, C. R., Poelstra, J. W., & Yoder, A. D. (2018). What is speciation genomics? The roles 777 of ecology, gene flow, and genomic architecture in the formation of species. *Biological Journal* 778 of the Linnean Society, 124(4), 1-23. 779 780 Castiglia, R., & Capanna, E. (2002). Chiasma repatterning across a chromosomal hybrid zone 781 between chromosomal races of Mus musculus domesticus. Genetica, 114, 35 – 40. 782 783 Cayuela, H., Dorant, Y., Merot, C., Laporte, M., Normandeau, E., Gagnon-Harvey, S., Sirois, P., 784 & Bernatchez, L. (2020). Thermal adaptation rather than demographic history drives genetic 785 structure inferred by copy number variants in a marine fish. *BioRkiv*, 786 https://doi.org/10.1101/2020.04.05.026443. 787 788 Chang, C. C., Carson, C. C., Tellier, L. C. A. M., Vattikuti, S., Purcell, S. M., & Lee, J. J. (2015). 789 Second-generation PLINK: rising to the challenge of larger and richer datasets. *GigaScience*, 790 4(7), doi:10.1186/s13742-015-0047-8. 791 792 Chaput, G. (2012). Overview of the status of Atlantic salmon (Salmo salar) in the North Atlantic 793 and trends in marine mortality. ICES Journal of Marine Science, 69, 1538-1548. 794 795 Charlesworth, B. (1985). Recombination, genome size and chromosome number. In T. Cavalier-796 Smith (Ed.), The evolution of genome size (pp 489 – 513). John Wiley & Sons, Chichester.

- 797
- Cheng, C-H., Ye, C-X., Guo, Z-X., & Wang, A-L. (2017). Immune and physiological responses of pufferfish (*Takifugu obscurus*) under cold stress. *Fish & Shellfish Immunology*, **64**, 137 – 145.
- 800
- 801 Corbett-Detig, R. B. (2016). Selection on inversion breakpoints favors proximity to pairing
- sensitive sites in *Drosophila melanogaster*, *Genetics*, **204**(1), 259 265.
- 803

804 COSEWIC. (2010). COSEWIC assessment and status report on the Atlantic Salmon Salmo salar

- 805 (Nunavik population, Labrador population, Northeast Newfoundland population, South
- 806 Newfoundland population, Southwest Newfoundland population, Northwest Newfoundland
- 807 population, Quebec Eastern North Shore population, Quebec Western North Shore population,
- 808 Anticosti Island population, Inner St. Lawrence population, Lake Ontario population, Gaspé-
- 809 Southern Gulf of St. Lawrence population, Eastern Cape Breton population, Nova Scotia
- 810 Southern Upland population, Inner Bay of Fundy population, Outer Bay of Fundy population) in
- 811 *Canada*. Committee on the Status of Endangered Wildlife in Canada. Ottawa. xlvii + 136 pp.
- 812
- 813 Coyne, J. A., & Orr, H. A. (2004). Speciation. Sinauer, Sunderland, MA.
- 814

815 Dalley, E. L., Andrews, C. W., & Green, J. M. (1983). Precocious male Atlantic salmon parr

- 816 (Salmo salar) in insular Newfoundland. Canadian Journal of Fisheries and Aquatic Science,
- **40**(5), 647 652.
- 818
- Bionne, M., Miller, K. M., Dodson, J. J., Caron, F., & Bernatchez, L. (2007). Clinal variation in
 MHC diversity with temperature: evidence for the role of host-pathogen interaction on local
- adaptation in Atlantic salmon. *Evolution*, **61**(9), 2154 2164.
- 822
- Bionne, M., Caron, F., Dodson, J. J., & Bernatchez, L. (2008). Landscape genetics and
 hierarchical genetic structure in Atlantic salmon: the interaction of gene flow and local
- 825 adaptation. *Molecular Ecology*, **17**(10), 2382 2396.
- 826

27 Dionne, M., Miller, K. M., Dodson, J. J., & Bernatchez, L. (2009). MHC standing genetic

- 828 variation and pathogen resistance in wild Atlantic salmon. *Philosophical Transactions of the*
- 829 *Royal Society B*, **364**(1523), https://doi.org/10.1098/rstb.2009.0011.
- 830
- B31 Dobigny, G., Britton-Davidian, J., & Robinson, T. J. (2017). Chromosomal polymorphism in
- 832 mammals: an evolutionary perspective. *Biological Reviews*, 92(1), 1 21.
- 833
- Bobzhansky, T. (1951). Genetics and the origin of species (3rd ed.) Columbia University Press.
- 836 Dumas, D., & Britton-Davidian, J. (2002). Chromosomal rearrangements and evolution of
- 837 recombination: comparison of chiasma distribution patterns in standard and Robertsonian
- 838 populations of the house mouse. *Genetics*, **162**(3), 1355 1366.
- 839

840 Feder, J. L., Nosil, P., & Flaxman, S. M. (2014). Assessing when chromosomal rearrangements 841 affect the dynamics of speciation: implications from computer simulations. Frontiers in Genetics, 842 https://doi.org/10.3389/fgene.2014.00295. 843 844 Felsentein, J. (1976). The theoretical population genetics of variable selection and migration. 845 Annual Review of Genetics, 10, 253 – 280. 846 847 Fick, S. E., & Hijmans, R. J. (2017). WorldClim 2: New 1-km spatial resolution climate surfaces 848 for global land areas. International Journal of Climatology, 37(12), 4302-4315. 849 850 Fisheries and Ocean Canada (2007). Placentia Bay integrated management plan. Fisheries and 851 Oceans Canada. Retrieved from the Fisheries and Marine Institute of Memorial University of 852 Newfoundland website: https://www.mi.mun.ca/media/marineinstitutewwwmimunca/mi/programsandcourses/marinespati 853 854 alplanning/files/PBIMCIntegratedManagementPlan.pdf 855 856 Friedland, K. D. (1998). Ocean climate influences on critical Atlantic salmon (Salmo salar) life 857 history events. Canadian Journal of Fisheries and Aquatic Science, 55 (Supplement 1), 119 – 858 130. 859 860 Fuller, Z. L., Koury, S. A., Phadnis, N., & Schaeffer, S. W. (2019). How chromosomal 861 rearrangements shape adaptation and speciation: case studies in Drosophila pseudoobscura and 862 its sibling species Drosophila persimilis. Molecular Ecology, **28**(6), 1238 – 1301. 863 864 Garcia de Leaniz, C., Fleming, I. A., Einum, S., Verspoor, E., Jordan, W. C., Consuegra, S., & ... 865 Quinn, T. P. (2007). A critical review of adaptive genetic variation in Atlantic salmon: 866 implications for conservation. *Biological Reviews*, **82**(2), 173 – 211. 867 868 Goudet, J. (2005). HIERFSTAT, a package for R to compute and test hierarchical F-statistics. 869 Molecular Ecology Notes, 5(1), 184 - 186. 870 871 Guerrero, R. F., & Kirkpatrick, M. (2014). Local adaptation and the evolution of chromosomes 872 fusions. *Evolution*, **68**(10), 2747 – 2756. 873 874 Hansen, G. J. A., Read, J. S., Hansen, J. F., & Winslow, L. A. (2016). Projected shifts in fish 875 species dominance in Wisconsin lakes under climate change. Global Change Biology, 23(4), 876 1463 - 1476. 877 878 Hartley, S. (1988). Cytogenetic studies of Atlantic salmon, Salmo salar L., in Scotland. Journal 879 of Fish Biology, 33, 735–740. 880 881 Hewitt, G. (2000). The genetic legacy of the Quaternary ice ages. *Nature*, **405**, 907 – 913. 882

- 883 Huang, K., Andrew, R. L., Owens, G. L., Ostevik, K., L., Rieseberg, L. H., (2019). Multiple 884 chromosomal inversions contribute to adaptive divergence of a dune sunflower ecotype. *BioRkiv*, 885 https://doi.org/10.1101/829622. 886 887 Jay, P., Whibley, A., Frézal, L., Rodríguez de Cara M. A., Nowell, R. W., Mallet, J., ... & Joron, 888 M. (2018). Supergene evolution triggered by the introgression of a chromosomal inversion. 889 *Current Biology*, https://doi.org/10.1016/j.cub.2018.04.072. 890 891 Jay, P., Chouteau, M., Whibley, A., Bastide, H., Llaurens, V., Parrinello, H., & Joron, M. (2019). 892 Mutation accumulation in chromosomal inversions maintains wing pattern polymorphism in a 893 butterfly. BioRxiv, https://doi.org/10.1101/736504. 894 895 Jonsson, B., & Jonsson, N. (2018). Egg incubation temperature affects the timing of the Atlantic 896 salmon Salmo salar homing migration. Journal of Fish Biology, 93(5), 1016 – 1020. 897 898 Kanehisa, M., Goto, S., Sato, Y., Furumichi, M., & Tanabe, M. (2012). KEGG for integration 899 and interpretation of large-scale molecular data sets. Nucleic Acids Research, 40(1), 109-114. 900 901 Kapun, M., Fabian, D. K., Goudet, J., & Flatt, T. (2016). Genomic evidence for adaptive 902 inversion clines in Drosophila melanogaster. Molecular Biology and Evolution, 33(5), 1317 -903 1336. 904 905 Kennington, W. J., & Hoffmann, A. A. (2013). Patterns of genetic variation across inversions: 906 geographic variation in the In(2L)t inversion in populations of *Drosophilia melanogaster* from 907 eastern Australia. BMC Evolutionary Biology, 13, 100. 908 909 Khan, R. A. (2009). Parasites causing disease in wild and cultured fish in Newfoundland. 910 *Icelandic Agricultural Sciences*, **22**, 29 – 35. 911 912 King, M. (1993). Species evolution: the role of chromosomal change. Cambridge University 913 Press, Cambridge, U. K. 914 915 King, T. L., Verspoor, E., Spidle, A. P., Gross, R., Phillips, R. B., Koljonen, M. L., & ... 916 Morrison, C. L. (2007). Biodiversity and population structure. In E. Verspoor, L. Stradmeyer, & 917 J. Nielsen (Eds.), The Atlantic Salmon: Genetics, conservation and management (pp. 117–166). 918 Oxford, UK: Blackwell Publishing Ltd. 919 920 Kirkpatrick, M., & Barton, N. (2006). Chromosome inversions, local adaptation and speciation. 921 *Genetics*, **173**, 419 – 434. 922 923 Knowles, L. L., Carstens, B. C., & Keat, M. L. (2007). Coupling genetic and ecological-niche 924 models to examine how past population distributions contribute to divergence. Current Biology, 925 17(11), 940–946.
- 926

- 927 Lamichhaney, S., Fan, G., Widemo, F., Gunnarsson, U., Schwochow Thalmann, D., Hoeppner,
- M. P., & ... Andersson, L. (2016). Structural genomic changes underlie alternative reproductive strategies in the ruff (*Philomachus pugnax*). *Nature Genetics*, **48**, 84 88.
- 930
- 931 Larson, W. A., Lisi, P. J., Seeb, J. E., Seeb, L. W., & Schindler, D. E. (2016). Major
- histocompatibility complex diversity is positively associated with stream water temperatures in
- proximate populations of sockeye salmon. *Journal of Evolutionary Biology*, **29**(9), 1846 1859.
 934
- Lee, C-R., Wang, B., Mojica, J. P., Mandáková, T., Prasad, K. V. S. K., Goicoechea, J. L., ... &
 Mitchell-Olds, T. (2017). Young inversion with multiple linked QTLs under selection in a hybrid
 zone. *Nature Ecology and Evolution*, 1:0119. doi:10.1038/s41559-017-0019
- 938
- Lehner, B., Verdin, K., & Jarvis, A. (2008). New global hydrography derived from spaceborne
 elevation data. *EOS*, 89(10), 93 94.
- 941
- Lehnert, S. J., Bentzen, P., Kess, T., Lien, S., Horne, J. B., Clement, M., & Bradbury, I. R.
- 943 (2019a). Chromosome polymorphisms track trans-Atlantic divergence and secondary contact in
 944 Atlantic salmon. *Molecular Ecology*, 28(8), 2074 2087.
 945
- Lehnert, S. J., Kess, T., Bentzen, P., Kent, M. P., Lien, S., Gilbey, J., Bradbury, I. R. (2019b).
 Genomic signatures and correlates of widespread population declines in salmon. *Nature Communications*. https://doi.org/10.1038/s41467-019-10972-w
- 949
- Luu, K., Bazin, E., & Blum, M. G. (2017). pcadapt: An R package to perform genome scans for
 selection based on principal component analysis. *Molecular Ecology Resources*, 17, 67–77.
- 952
- McAllister, B. F. (2003). Sequence differentiation associated with an inversion on the neo-X
 chromosome of *Drosophila americana*. *Genetics*, 165(3), 1317 1328.
- 955
- McCartney-Melstad, E., Vu, J.K., & Shaffer, H.B. (2018). Genomic data recover previously
 undetectable fragmentation effects in an endangered amphibian. *Molecular Ecology*, 27(22),
 4430 4443.
- McDonald, M. J., Rice, D. P., & Desai, M. M. (2016). Sex speeds adaptation by altering the dynamics of molecular evolution. *Nature*, **531**(7593), 233–236.
- 962

- Mangel, M. (1994). Climate change and salmonid life history variation. *Deep-Sea Research*, 41,
 75 106.
- 965
- Marques, D. A., Meier, J. I., & Seehausen, O. (2019). A combinatorial view on speciation and
 adaptive radiation. *Trends in Ecology and Evolution*, 34(6), 531 544.
- 968
- 969
- 970

971 Mérot, C., Berdan, E. L., Babin, C., Normandeau, E., Wellenreuther, M., & Bernatchez, L.

972 (2018). Intercontinental karyotype-environment parallelism supports a role for a chromosomal

- 973 inversion in local adaptation in a seaweed fly. *Proceedings of the Royal Society B*, 285(1881),
- 974 https://doi.org/10.1098/rspb.2018.0519
- 975
- Mérot, C., Oomen, R. A., Tigano, A., & Wellenreuther, M. (2020). A roadmap for understanding
 the evolutionary significance of structural genomic variation. *Trends in Ecology & Evolution*, (in
- 978 press). https://doi.org/10.1016/j.tree.2020.03.002.
- 979

986

- Metcalfe, N. B., & Thorpe, J. E. (1990). Determinants of geographical variation in the age of
 seaward-migrating salmon, *Salmo salar. Journal of Animal Ecology*, **59**(1), 135 145.
- Minns, C. K., Randall, R. G., Chadwick, E. M. P., Moore, J. E., & Green, R. (1995). In Climate
 change and northern fish populations. Edited by R. J. Beamish, *Canadian Special Publication of Fisheries and Aquatic Sciences*, 121, 699 708.
- Nei, M. (1972). Genetic distance between populations. *The American Naturalist*, **106**, 283 292.
- Nei, M. (1987). *Molecular evolutionary genetics*. Columbia University Press, New York.

Nicieza, A G., Reyes-Gavilán, F. G., & Braña, F. (1994). Differentiation in juvenile growth and
bimodality patterns between northern and southern populations of Atlantic salmon (*Salmo salar*L.). *Canadian Journal of Fisheries and Aquatic Sciences*, 72(9), 1603 – 1610.

- L.). Canadian Journal of Fisheries and Aquatic Sciences, 12(9), 1603 1610.
 994
- 995 Nilsson, J., Gross, R., Asplund, T., Dove, O., Jansson, H., Kellonieni, J. & Lumme, J.
- 996 (2001). Matrilineal phylogeography of Atlantic salmon (*Salmo salar* L.) in Europe and
 997 postglacial colonization of the Baltic Sea area. *Molecular Ecology*, **10**(1): 89 92.
- 998
- Oksanen, J., Guillaume Blanchet, F., Friendly, M., Kindt, R., Legendre, P., McGlinn, D., ... &
 Wagner, H. (2017). vegan: Community ecology package. *R package version 2.4-5*. Retrieved
- 1001 from https://github.com/vegandevs/vegan
- 1002
- 1003 Oomen, R. A., Kuparinen, A., & Hutchings, J. A. (2020). Consequences of single-locus and
 1004 tightly linked genomic architectures for evolutionary responses to environmental change.
 1005 *BioRxiv*, https://doi.org/10.1101/2020.01.31.928770.
- 1006
- Palstra, F. P., O'Connell, M. F., & Ruzzante, D. E. (2007). Population structure and gene flow
 reversals in Atlantic salmon (*Salmo salar*) over contemporary and long-term temporal scales:
- 1009 effects of population size and life history. *Molecular Ecology*, **16**(21), 4504 4522.
- 1010
- 1011 Pante, E., & Simon-Bouhet, B. (2013). marmap: a package for importing, plotting and analyzing
- 1012 bathymetric and topographic data in R. *PLoS One*, **8**(9): e73051.
- 1013

- 1014 Pembleton, L. W., Cogan, N. O. I., & Forster, J. W. (2013). StAMPP: an R package for
- calculation of genetic differentiation and structure of mixed-ploidy level populations. *Molecular Ecology Resources*, 13(5), 946 952.
- 1017
- 1018 Phillips, R., & Ráb, P. (2001). Chromosome evolution in the Salmonidae (Pisces): an update.
- 1019 *Biological Reviews.* **76**, 1 25.
- 1020
- 1021 Poelstra, J. W., Richards, E. J., & Martin, C. H. (2018). Speciation in sympatry with ongoing
- secondary gene flow and a potential olfactory trigger in a radiation of Cameroon cichlids.
 Molecular Ecology, 27(21), 4270 4288.
- 1024

- Porter, T. R., Riche, L. G., & Traverse, G. R. (1974). *Catalogue of rivers in insular Newfoundland, vol. A.*, Department of the environment, St. John's, NL.
- Power, G. (1981). Stock characteristics and catches of Atlantic salmon (Salmo salar) in Quebec,
 and Newfoundland and Labrador in relation to environmental variables. *Canadian Journal of Fisheries and Aquatic Sciences*, 38(12), 1601 1611.
- 10311032 Puerma, E., Orengo, D. J., & Aguadé, M. (2016). Multiple and diverse structural changes affect
- the breakpoint regions of polymorphic inversions across the *Drosophila* genus. *Scientific Reports*, 6, 36248. https://doi.org/10.1038/srep36248
- 1035
- Purcell, S., Neale, B., Todd-Brown, K., Thomas, L., Ferreira, M. A. R., Bender, D., ... & Sham,
 P. C. (2007). PLINK: A tool set for whole-genome association and population-based linkage
 and produces The American Learning of Human Compting **91**, 550, 575
- analyses. *The American Journal of Human Genetics*, **81**, 559–575.
- 1040 R Core Team. (2019). R: a language and environment for statistical computing. *R Foundation for* 1041 Statistical Computing, Vienna, Austria. http://www.R-project.org/
- 1042
- 1043 Quinlan, A. R., & Hall, I. M. (2010). BEDTools: A flexible suite of utilities for comparing
 1044 genomic features. *Bioinformatics*, 26(6), 841–842
- 1045
- 1046 Rajagopal, V. M. (2020). *ggman*: R package for Manhattan plots, GitHub repository 1047 https://github.com/drveera/ggman/
- 1048
- 1049 Rambaut, A. (2012). FigTree (Version 1.4). Retrieved from
- 1050 http://tree.bio.ed.ac.uk/software/figtree/
- 1051
- 1052 Rieseberg, L. H. (2001). Chromosomal rearrangements and speciation. *Trends in Ecology and* 1053 *Evolution*, **16**(7), 351 – 358.
- 1054
- 1055 Rieseberg, L. H. (2009). Evolution: replacing genes and traits through hybridization. Current
- 1056 *Biology*, **19**(3), R119-R122.
- 1057

1058 Rougemont, Q., & Bernatchez, L. (2018). The demographic history of Atlantic salmon (Salmo 1059 salar) across its distribution range reconstructed from approximate Bayesian computations. 1060 *Evolution*, **72**(6), 1261 – 1277. 1061 1062 Samy, J. K. A., Mulugeta, T. D., Nome, T., Sandve, S. R., Grammes, F., Kent, M. P., ... & Våge, 1063 D. I. (2020). SalmoBase: an integrated molecular data resource for Salmonid species. BMC 1064 Genomics, 18, 482. Doi:10.1186/s12864-017-3877-1. 1065 1066 Savolainen, O., Lascoux, M., & Merilä, J. (2013). Ecological genomics of local adaptation. *Nature Reviews Genetics*, **14**, 807 – 820. 1067 1068 1069 Schulter, D. (2009). Evidence for ecological speciation and its alternative. Science, **323**(5915), 1070 737 – 741. 1071 1072 Sinclair-Waters, M., Bradbury, I. R., Morris, C. J., Lien, S., Kent, M. P. & Bentzen, P. (2018). 1073 Ancient chromosomal rearrangement associated with local adaptation of a postglacially colonized 1074 population of Atlantic cod in the northwest Atlantic. *Molecular Ecology*, **27**(2), 339 – 351. 1075 1076 Slatkin, M. (1975). Gene flow and selection in a two-locus system. *Genetics*, **81**(4), 787 – 802. 1077 1078 Stehfest, K. M., Carter, C. G., McAllister, J. D., Ross, J. D., & Semmens, J. M. (2017). Response 1079 of Atlantic salmon Salmo salar to temperature and dissolved oxygen extremes established using 1080 animal-borne environmental sensors. Scientific Reports, 7, 4545, https://doi.org/10.1038/s41598-1081 017-04806-2 1082 1083 Sterud, E., Forseth, T., Ugedal, O., Poppe, T. T., Jorgensen, A., Bruheim, T., ... & Mo, T. A. 1084 (2007). Severe mortality in wild Atlantic salmon Salmo salar due to proliferative kidney disease 1085 (PKD) caused by Tetracapsuloides bryosalmonae (Myxozoa). Diseases of Aquatic Organisms, 1086 77, 191 – 198. 1087 1088 Storey, J. D., Bass, A. J., Dabney, A., & Robinson, D. (2015). qvalue: Q-value estimation for 1089 false discovery rate control. (R package 2.10.0). Vienna, Austria: R Foundation for Statistical 1090 Computing. Retrieved from http://github.com/jdstorey/qvalue 1091 1092 Storfer, A., Murphy, M. A., Evans, J. S., Goldberg, C. S., Robinson, S., Spear, S. F., & ... Waits, 1093 L. P. (2006). Putting the 'landscape' in landscape genetics. *Heredity*, 98(3), 128-142. 1094 1095 Sturtevant, A. H. (1917). Genetic factors affecting the strength of linkage in *Drosophilia*. 1096 Proceedings of the National Academy of Science USA, 3:555. 1097 1098 Sturtevant, A. H. (1938). The interrelations of inversions, heterosis and recombination. The 1099 American Naturalist, 72:742, 447 - 452. 1100

- 1101 Sylvester, E.V.A., Beiko, R. G., Bentzen, P., Paterson, I., Horne, J. B., Watson, B., ... &
- 1102 Bradbury, I.R. (2018). Environmental extremes drive population structure at the northern range
- 1103 limit of Atlantic salmon in North America. *Molecular Ecology*, **27**(20), 4026 4040.
- 1104
- 1105 Sylvester, E.V.A., Wringe, B.F., Duffy, S.J., Hamilton, L.C., Fleming, I.A., Castellani, M., ... &
- 1106 Bradbury, I.R. (2019). Estimating the relative fitness of escaped farmed salmon offspring in the
- wild and modelling the consequences of invasion for wild populations. *Evolutionary Applications*, 12(4), 705 717.
- 1108 1109
- 1110 Taylor, E. B. (1991). A review of local adaptation in Salmonidae, with particular references to 1111 Pacific and Atlantic salmon. *Aquaculture*, 98(1-3), 185-207.
- Tigano, A., & Friesen, V. L. (2016). Genomics of local adaptation with gene flow. *Molecular Ecology*, 25(10), 2144 2164.
- 1115

- Turner, S. D. (2014). Qqman: An R package for visualizing GWAS results using QQ and
 manhattan plots. *BioRxiv*, 005165.
- 1119 Valiente, A. G., Juanes, F., & Garcia-Vazquez, E. (2005). Reproductive strategies explain genetic
 1120 diversity in Atlantic salmon, *Salmo salar. Environmental Biology of Fishes*, 74, 323 334.
- 1121
- 1122 Verspoor, E., McGinnity, P., Bradbury, I., & Glebe, B. (2015). The potential direct and indirect
 1123 genetic consequences for native Newfoundland Atlantic salmon from interbreeding with
 1124 European-origin farm escapes. *DFO Can. Sci. Advis. Sec. Res. Doc.* 2015/030. vii + 36 p.
- European-origin farm escapes. DFO Can. Sci. Aavis. Sec. Res. Doc. 2015/050. VII + 50 p 1125
- Vikeså, V., Nankervis, L., & Hevrøy, E. M. (2017). Appetite, metabolism and growth regulation
 in Atlantic salmon (*Salmo salar* L.) exposed to hypoxia at elevated seawater temperature. *Aquaculture Research*, 48(8), 4086 4101.
- 1129
- Wang, Y., Li, C., Pan, C., Liu, E., Zhao, X., & Ling, Q. (2019). Alterations to transcriptomic
 profile, histopathology, and oxidative stress in liver of pikeperch (*Sander lucioperca*) under heat
 stress. *Fish & Shellfish Immunology*, **95**, 659 669.
- 1133 1134 Warnes, G. R., Bolker, B., Bonebakker, L., Gentleman, R., Liaw, W. H. A., Lumley, T., ...
- 1135 Venables, B. (2016). gplots: Various R programming tools for plotting data (R package 3.0.1).
- 1136 Vienna, Austria: *R Foundation for Statistical Computing*.
- 1137
- 1138 Weir, B. S., & Cockerham, C. C. (1984). Estimating *F*-statistics for the analysis of population 1139 structure. *Evolution*, 38(6), 1358 – 1370.
- 1140
- 1141 Wellband, K., Mérot, C., Linnansaari, T., Elliott, J. A, K., Curry, A., Bernatchez, L. (2019).
- 1142 Chromosomal fusion and life history-associated genomic variation contribute to within-river
- 1143 local adaptation of Atlantic salmon. *Molecular Ecology*, **28**(6), 1439 1459.
- 1144

- 1145 Wellenreuther, M., & Bernatchez, L. (2018). Eco-evolutionary genomics of chromosomal
- 1146 inversions. Trends in Ecology & Evolution, **33**(6), 427 440.
- 1147
- 1148 Wellenreuther, M., Mérot, C., Berdan, E., & Bernatchez, L. (2019). Going beyond SNPs: the role
- of structural genomic variants in adaptive evolution and species diversification. *Molecular Ecology*, 28(6), 1203 1209.
- 1151
- 1152 White, M. J. D. (1978). *Modes of speciation*. W. H. Freeman, San Francisco, CA.
- 1153

- Wright, S. (1931). *Evolution in Mendelian populations*. University of Chicago, Chicago, Illinois.
- Yada, T., & Tort, L. (2016). Stress and disease resistance: Immune system and immunoendocrine
 interactions. *Fish Physiology*, 35, 365 403.
- Yates, M. C., Debes, P. V., Fraser, D. J., Hutchings, J. A. (2015). The influence of hybridization
 with domesticated conspecifics on alternate reproductive phenotypes in male Atlantic salmon in
 multiple temperature regimes. *Canadian Journal of Fisheries and Aquatic Sciences*, 72(8), 1138
 1145.
- 1163
- Yu, G., Wang, L.-G., Han, Y., & He, Q.-Y. (2012). clusterProfiler: An R package for comparing
 biological themes among gene clusters. *OMICS: A Journal of Integrative Biology*, 16(5), 284–
 287.
- Zhang, Y., Qin, C., Yang., L., Lu, R., Zhao, X., & Nie, G. (2018). A comparative genomics study
 of carbohydrate/glucose metabolic genes: from fish to mammals. *BMC Genomics*, 246.
 https://doi.org/10.1186/s12864-018-4647-4.
- 1171
- 1172

1173 AUTHOR CONTRIBUTIONS

- 1174 K.B.W., S.J.L., I.R.B., and P.B. contributed to the conception and design of the study. M.K., and
- 1175 S.L. K.B.W., S.J.L., A.E., B.P., and T.K. performed statistical analyses. I.R.B., S.L., M.K., and
- 1176 S.D. provided molecular data and metadata for the study. K.B.W. drafted the manuscript and all
- 1177 authors contributed to the writing and approved the final draft of the manuscript.
- 1178
- 1179 **DATA ACCESSIBILITY** (data must be archived on a publicly accessible repository)
- 1180

1182	Table	e 1	Sampling	locations	of Atla	ntic	Salmon	(Salmo	salar)) in	Placent	ia Bay	, Newfoun	dlan	d,
1100	\sim		n '	1 1							~	4	11		

1183 Canada. Rivers ordered geographically, east to west around the bay. Samples were collected in 1184 2017 and 2018. Number of samples genotyped (*N*) per site per year.

River	RiverID	Longitude (°W)	Latitude (°N)	N2017	N2018	
Branch River	BRA	-53.97	46.89	30	32	
Lance River	LAN	-54.07	46.82	9	-	
Cuslett Brook	CUS	-54.16	46.96	30	30	
Great Barasway Brook	GBW	-54.06	47.13	30	41	
Little Barasway Brook	LBB	-54.04	47.18	16	-	
Southeast Placentia River	SPR	-53.88	47.23	30	31	
Northeast Placentia River	NPR	-53.84	47.27	30	24	
Ship Harbour Brook	SHI	-53.87	47.35	30	32	
Fair Haven Brook	FHB	-53.89	47.54	30	30	
Come By Chance River	CBC	-53.98	47.86	30	31	
North Harbour River	NHR	-54.03	47.92	30	32	
Black River	BLA	-54.16	47.89	30	30	
Pipers Hole Brook	PHR	-54.27	47.93	30	33	
Sandy Harbour River	SHA	-54.36	47.71	30	9	
Nonsuch River	NON	-54.65	47.45	30	20	
Cape Roger Brook	CRB	-54.69	47.44	30	32	
Bay de l'Eau River	BDL	-54.73	47.51	30	32	
Rushoon River	RUS	-54.92	47.37	30	37	
Red Harbour East River	RHA	-54.99	47.33	30	26	
Red Harbour West River	RHW	-55.01	47.29	30	30	
Northwest Mortier Bay Brook	NMB	-55.31	47.17	30	32	
Tides Brook	TDS	-55.26	47.13	30	28	
Big Salmonier Brook	BSA	-55.22	47.06	30	31	
Lawn River	LWN	-55.54	46.95	30	30	
Taylor Bay Brook	TBR	-55.71	46.88	30	10	
Piercey's Brook	PBR	-55.86	46.88	30	21	

1190	Table 2 Summary	of genetic	diversity	for Atlantic S	Salmon from	26 rivers in Pla	acentia Bay,
------	-----------------	------------	-----------	----------------	-------------	------------------	--------------

Newfoundland, Canada. Rivers are ordered geographically, east to west around the bay. Number of samples (N), mean observed (H_o) and expected (H_e) heterozygosity and median F_{IS} calculated

|--|

DivorID			2017				2018	
KIVEIID	Ν	Ho	He	FIS	Ν	Ho	He	FIS
BRA	30	0.184	0.182	-0.025	32	0.187	0.183	-0.025
LAN	9	0.181	0.170	-0.067	-	-	-	-
CUS	30	0.184	0.185	-0.018	30	0.189	0.185	-0.036
GBW	18	0.188	0.186	-0.030	41	0.189	0.186	-0.026
LBB	15	0.191	0.184	-0.037	-	-	-	-
SPR	27	0.192	0.185	-0.040	30	0.194	0.188	-0.036
NPR	29	0.193	0.192	-0.018	17	0.192	0.191	-0.032
SHI	22	0.202	0.200	-0.024	32	0.204	0.201	-0.018
FHB	30	0.188	0.185	-0.036	25	0.193	0.188	-0.029
CBC	30	0.192	0.194	-0.018	19	0.194	0.192	-0.029
NHR	24	0.188	0.188	-0.022	29	0.192	0.189	-0.018
BLA	24	0.196	0.195	-0.022	29	0.198	0.198	-0.018
PHR	29	0.193	0.193	-0.018	30	0.196	0.194	-0.018
SHA	30	0.198	0.196	-0.018	9	0.197	0.198	0.000
NON	27	0.181	0.177	-0.020	20	0.181	0.180	-0.027
CRB	29	0.190	0.190	-0.018	27	0.183	0.182	-0.020
BDL	30	0.181	0.181	-0.018	32	0.188	0.187	-0.016
RUS	25	0.182	0.183	-0.021	31	0.182	0.182	-0.017
RHA	30	0.158	0.158	-0.018	26	0.159	0.158	-0.020
RHW	27	0.196	0.196	-0.020	24	0.196	0.194	-0.022
NMB	27	0.185	0.181	-0.040	31	0.193	0.189	-0.035
TDS	15	0.179	0.177	-0.037	24	0.186	0.185	-0.022
BSA	30	0.188	0.187	-0.018	24	0.187	0.184	-0.022
LWN	27	0.193	0.190	-0.020	22	0.195	0.194	-0.024
TBR	18	0.193	0.194	-0.030	10	0.192	0.199	0.000
PBR	30	0.194	0.194	-0.018	17	0.194	0.192	-0.032





Figure 1 Sampling locations (n = 26, red circles) of juvenile Atlantic Salmon (*Salmo salar*) in

Placentia Bay, Newfoundland, Canada. Samples collected in 2017 and 2018; see Table 1 for details.



Figure 2 Genomic outlier blocks drive spatial structure of Atlantic Salmon. (a, b) Genetic
structure across Placentia Bay, Newfoundland, Canada based on the first two principal
component (PC) axes from *pcadapt* (Luu et al., 2017) using 138,451 SNPs. (c, d) Manhattan
plots showing genomic regions of variation based on PC1. Samples collected in (a, c) 2017 (b, d)
2018. Rivers coloured east (yellow-red) to west (green-blue) with head of the bay (purple); see
Figure 1 and Table 1 for location details. Red line represents a genome-wide significance
threshold of 5.0e-8.





- 1212 (WorldClim) temperature and precipitation variables, and habitat variables. Manhattan plots,
- 1213 showing absolute loadings, of the distribution of outlier SNPs (blue) associated with the first
- 1214 RDA axis of (c) 2017 and (d) 2018. See Figure 1 and Table 1 for location details.
- 1215





1217 Figure 4 Genetic relationships between individual Atlantic Salmon based on outlier SNPs (q <



- 1219 2018. Neighbor-joining (NJ) tree for (c) 2017 and (d) 2018. Homozygous European non-
- 1220 translocated (Ssa01p/q and Ssa23) karyotype (yellow), heterozygous (red), and homozygous
- 1221 North American translocated (Ssa01q and Ssa01p/23) karyotype (blue-green). See Figure 1 and
- 1222 Table 1 for location details.
- 1223



Figure 5 Frequency of Ssa01 and Ssa23 chromosomal translocation exhibits fine-scale spatial
variability in Placentia Bay, Newfoundland, Canada. (a, b) Karyotype frequency within river in
(a) 2017 and (b) 2018. Asterisk (*) indicates karyotype frequency differed significantly between
years. (c) Translocation frequency between rivers across years. Homozygous European nontranslocated (Ssa01p/q and Ssa23) karyotype (yellow), heterozygous (red), and homozygous
North American translocated (Ssa01q and Ssa01p/23) karyotype (blue-green). See Table 1 for
location and sample details.



Figure 6 Correlation between temperature and the Ssa01/Ssa23 chromosomal translocation in Atlantic Salmon within Placentia Bay, Newfoundland, Canada. (a, c) Linear regression of the first principal component (PC) of a PCA based on 11 temperature variables (BIOCLIM) and proportion of non-translocated Ssa01p/q sampled in (a) 2017 and (c) 2018. (b, d) Temperature annual range (BIO7), the highest loading variable on temperature PC1. Size of point indicates frequency of non-translocated Ssa01p/q in (b) 2017 and (d) 2018. See Table 1 for location details.

1245 Supplemental Information

1246

- 1247 Table S1 Climatic and habitat variables used to identify potential drivers of translocation
- 1248 frequency using a partial redundancy analysis (RDA). Description of each measure is provided
- 1249 with the years measured, unit, and data source.

Variable	Description	Unit	Year	Source
BIO1	Annual mean temperature	۰C	1970-2000	WorldClim
BIO2	Mean diurnal range	۰C	1970-2000	WorldClim
BIO3	Isothermality	۰C	1970-2000	WorldClim
BIO4	Temperature seasonality	۰C	1970-2000	WorldClim
BIO5	Max temperature of warmest month	۰C	1970-2000	WorldClim
BIO6	Min temperature of coldest month	۰C	1970-2000	WorldClim
BIO7	Temperature annual range	۰C	1970-2000	WorldClim
BIO8	Mean temperature wettest quarter	۰C	1970-2000	WorldClim
BIO9	Mean temperature of driest quarter	۰C	1970-2000	WorldClim
BIO10	Mean temperature warmest quarter	۰C	1970-2000	WorldClim
BIO11	Mean temperature coldest quarter	۰C	1970-2000	WorldClim
BIO12	Annual precipitation	mm	1970-2000	WorldClim
BIO13	Precipitation of wettest month	mm	1970-2000	WorldClim
BIO14	Precipitation of driest month	mm	1970-2000	WorldClim
BIO15	Precipitation seasonality	mm	1970-2000	WorldClim
BIO16	Precipitation of wettest quarter	mm	1970-2000	WorldClim
BIO17	Precipitation of driest quarter	mm	1970-2000	WorldClim
BIO18	Precipitation of warmest quarter	mm	1970-2000	WorldClim
BIO19	Precipitation of coldest quarter	mm	1970-2000	WorldClim
Human density	Human population density		2000	NASA NEO
Axial length	Length of river along down-valley axis	km	1974	DFO
Obstructions	Number of obstructions		1974	DFO
	Difference in elevation between the			
Relief	highest and lowest point of the basin	m	1974	DFO
FW_dem_max	Maximum elevation	m	2000	HydroSHEDS

1250

- 1253 Table S2 Outlier block regions associated with a known chromosomal translocation
- 1254 (Ssa01p/Ssa23) in Atlantic salmon identified here using *pcadapt* across rivers in Placentia Bay,
- 1255 Newfoundland, Canada. Outlier block regions were approximated by visual inspection of q-
- 1256 values. Boundaries of and number of significant (q-value < 0.05) single nucleotide
- 1257 polymorphisms (SNPs) in each region reported.

	Chromosome	Approximate bound	aries of outlier blocks	Number of
		Start (bp)	End (bp)	outlier SNPs
	Ssa01	44,000,000	53,000,000	480
	Ssa23	0	9,500,000	407
1258				
1259				
1260				
1261				
1262				
1202				
1263				
1264				
12(5				
1265				
1266				
1267				
1207				
1268				
1269				
1050				
12/0				
1271				

Climatia waniahla	Lo	ading
Climatic variable	PC1	PC2
Temperature		
BIO1	-0.19	-0.62
BIO2	0.34	-0.11
BIO3	0.20	-0.26
BIO4	0.36	-0.013
BIO5	0.34	-0.078
BIO6	-0.36	-0.036
BIO7	0.36	0.001
BIO8	-0.17	-0.54
BIO9	0.25	-0.23
BIO10	0.30	-0.38
BIO11	-0.35	-0.2
Precipitation		
BIO12	-0.39	0.048
BIO13	-0.38	0.18
BIO14	-0.37	-0.20
BIO15	-0.17	0.81
BIO16	-0.38	0.19
BIO17	-0.35	-0.42
BIO18	-0.37	-0.24
BIO19	-0.37	0.01

1273 Table S3 Principal component, PC1 and PC2, loadings for each categorical PCA used in

1274 r	redundancy	analyses	(RDA)	with	code o	of clir	natic	variable.
--------	------------	----------	-------	------	--------	---------	-------	-----------

1282Table S4 Proportion of karyotypes for the Ssa01p/Ssa23 chromosomal translocation in Placentia1283Bay, Newfoundland, Canada. Karyotype assigned based on outlier SNPs (n = 887) from the1284outlier block regions on Ssa01p and Ssa23. Rivers are ordered geographically, east to west1285around the bay. See Figure 1 and Table 1 for location details.

		2017			2018	
RiverID	Homo NA (Trans)	Hetero	Homo EU (No Trans)	Homo NA (Trans)	Hetero	Homo EU (No Trans)
BRA	0.000	0.233	0.767	0.000	0.250	0.750
CUS	0.667	0.267	0.067	0.267	0.533	0.200
GBW	0.111	0.556	0.333	0.073	0.341	0.585
SPR	0.000	0.111	0.889	0.000	0.133	0.867
NPR	0.138	0.517	0.345	0.118	0.294	0.588
SHI	0.045	0.409	0.545	0.000	0.438	0.563
FHB	0.533	0.400	0.067	0.320	0.600	0.080
CBC	0.533	0.400	0.067	0.421	0.526	0.053
NHR	0.667	0.292	0.042	0.310	0.552	0.138
BLA	0.542	0.458	0.000	0.655	0.276	0.069
PHR	0.310	0.655	0.034	0.567	0.367	0.067
SHA	0.667	0.333	0.000	0.889	0.111	0.000
NON	0.704	0.222	0.074	0.700	0.300	0.000
CRB	0.517	0.414	0.069	0.778	0.222	0.000
BDL	0.433	0.467	0.100	0.313	0.500	0.188
RUS	0.720	0.280	0.000	0.742	0.258	0.000
RHW	0.630	0.333	0.037	0.625	0.250	0.125
NMB	0.222	0.444	0.333	0.645	0.355	0.000
TDS	0.200	0.533	0.267	0.375	0.333	0.292
BSA	0.633	0.367	0.000	0.250	0.625	0.125
LWN	0.667	0.333	0.000	0.591	0.318	0.091
TBR	0.556	0.333	0.111	0.500	0.400	0.100
PBR	0.133	0.600	0.267	0.294	0.588	0.118

1289 Table S5 Pairwise comparison of translocation and karyotype frequencies within river between

1290 years (2017 and 2018) sampled. Bold *p*-values indicate significantly different frequencies,

1291 calculated using a Fisher Exact Test. Rivers ordered geographically, east to west, around

1292 Placentia Bay, Newfoundland; see Figure 1 and Table 1 for location details.

RiverID	Tr	anslocation	frequency		F	Karyotype fre	equency	
RIVEIID	Estimate	lower CI	upper CI	р	Estimate	lower CI	upper CI	р
BRA	1.08	0.32	3.77	1.00	0.00	0.00	Inf	1.00
CUS	0.29	0.12	0.69	0.00	7.04	0.99	85.86	0.04
GBW	0.51	0.20	1.29	0.13	2.58	0.18	28.50	0.57
SPR	1.21	0.19	8.68	1.00	0.00	0.00	Inf	1.00
NPR	0.55	0.19	1.50	0.26	1.95	0.22	26.24	0.65
SHI	0.84	0.31	2.32	0.82	Inf	0.04	Inf	0.42
FHB	0.60	0.24	1.44	0.22	1.95	0.12	31.65	0.60
CBC	0.79	0.30	2.14	0.65	1.00	0.02	22.06	1.00
NHR	0.33	0.12	0.86	0.02	6.66	0.55	370.64	0.14
BLA	1.14	0.40	3.18	0.82	Inf	0.12	Inf	0.51
PHR	1.69	0.72	4.08	0.23	1.06	0.05	69.29	1.00
SHA	3.36	0.42	156.00	0.44	0.00	0.00	Inf	1.00
BDL	0.65	0.29	1.42	0.27	2.52	0.41	19.57	0.43
NON	1.28	0.38	4.75	0.78	0.00	0.00	8.01	0.51
CRB	3.02	1.01	10.31	0.03	0.00	0.00	4.25	0.19
RUS	1.10	0.31	3.78	1.00	0.00	0.00	Inf	1.00
RHW	0.77	0.27	2.16	0.64	3.29	0.24	188.57	0.60
NMB	5.70	2.32	14.89	0.00	0.00	0.00	0.22	0.00
TDS	1.35	0.49	3.73	0.64	0.60	0.06	4.89	0.67
BSA	0.29	0.11	0.74	0.01	Inf	0.99	Inf	0.03
LWN	0.60	0.20	1.81	0.33	Inf	0.23	Inf	0.20
TBR	0.90	0.23	3.68	1.00	1.00	0.01	23.98	1.00
PBR	1.86	0.73	4.80	0.20	0.22	0.01	2.11	0.17

1288

1293

1294

1297 Table S6 Gene ontology (GO) enrichment, using topGO, for single nucleotide polymorphisms

1298 (SNPs) identified as outliers in a genotype-environment analysis (redundancy analysis; RDA)

1299 and located within the outlier block regions on chromosomes Ssa01 and Ssa23. GO IDs in bold

1300 indicate the term was significant (p-value < 0.01) in both discrete years sampled.

CO ID				2017	p-value N_{anno} N_{sig} 0.000319300.00076200.0016747900.00169200.00228300.002812921800.002912200.002912200.0034132	2018			
GOID	Description	$N_{ m anno}$	$N_{ m sig}$	Nexp	<i>p</i> -value	$N_{ m anno}$	$N_{ m sig}$	Nexp	<i>p</i> -value
GO:0045887	positive regulation of synaptic growth at neuromuscular junction	19	3	0.13	0.0003	19	3	0.12	0.00
GO:2000541	protein geranylgeranylation	6	2	0.040	0.0007	6	2	0.04	0.00
GO:0008380	RNA splicing	747	9	5.05	0.0016	747	9	4.8	0.0013
GO:0006729	tetrahydrobiopterin biosynthetic process	9	2	0.060	0.0016	9	2	0.06	0.0014
GO:0045075	regulation of interleukin-12 biosynthetic process	28	4	0.19	0.0002	28	3	0.18	0.0046
GO:0007346	regulation of mitotic cell cycle	1292	18	8.74	0.0028	1292	18	8.31	0.0023
GO:0031622	positive regulation of fever generation	12	2	0.080	0.0029	12	2	0.08	0.0026
GO:0031394	positive regulation of prostaglandin biosynthetic process	12	2	0.080	0.0029	12	2	0.08	0.0026
GO:0021912	regulation of transcription from RNA polymerase II promoter involved in spinal cord motor neuron fate specification	13	2	0.090	0.0034	13	2	0.08	0.0031

GO:0006979	response to oxidative stress	1072	13	7.25	0.0043	1072	13	6.89	0.0032
GO:0002091	negative regulation of receptor internalization	15	2	0.10	0.0045	15	2	0.1	0.0041
GO:1900025	negative regulation of substrate adhesion- dependent cell spreading	16	2	0.11	0.0051	16	2	0.1	0.0047
GO:0021913	regulation of transcription from RNA polymerase II promoter involved in ventral spinal cord interneuron specification	19	2	0.13	0.0072	13	2	0.08	0.0031
GO:0008152	metabolic process	19535	141	132.10	0.0022	19535	134	125.64	0.0084
GO:0014850	response to muscle activity	53	3	0.36	0.0056	53	3	0.34	0.0048
GO:0048515	spermatid differentiation	330	4	2.23	0.0007	330	3	2.12	0.0122
GO:0090076	relaxation of skeletal muscle	18	2	0.12	0.0065	18	2	0.12	0.0059
GO:0034612	response to tumor necrosis factor	321	7	2.17	0.0079	321	7	2.06	0.0069
GO:0071603	endothelial cell-cell adhesion	20	2	0.14	0.0080	20	2	0.13	0.0072
GO:0046326	positive regulation of glucose import	140	5	0.95	0.0027	140	4	0.9	0.0129
GO:0043201	response to leucine	20	2	0.14	0.0080	20	2	0.13	0.0072
GO:1900028	negative regulation of ruffle assembly	21	2	0.14	0.0088	21	2	0.14	0.008
GO:0010629	negative regulation of gene expression	4286	35	28.98	0.0104	4286	35	27.57	0.0079
GO:0032496	response to lipopolysaccharide	727	10	4.92	0.0053	727	9	4.68	0.0135
GO:0048535	lymph node development	65	3	0.44	0.0098	65	3	0.42	0.0085

GO:0071340	skeletal muscle acetylcholine-gated channel clustering	23	2	0.16	0.0105	23	2	0.15	0.0095
GO:0021918	regulation of transcription from RNA polymerase II promoter involved in somatic motor neuron fate commitment	7	1	0.050	0.0464	13	2	0.08	0.0031
GO:0048378	regulation of lateral mesodermal cell fate specification	12	2	0.080	0.0029	12	1	0.08	0.0745
GO:0003256	regulation of transcription from RNA polymerase II promoter involved in myocardial precursor cell differentiation	39	1	0.26	1.0000	13	2	0.08	0.0031
GO:1901213	regulation of transcription from RNA polymerase II promoter involved in heart development	62	1	0.42	1.0000	13	2	0.08	0.0031
GO:0000430	regulation of transcription from RNA polymerase II promoter by glucose	8	0	0.050	1.0000	13	2	0.08	0.0031
GO:0000431	regulation of transcription from RNA polymerase II promoter by galactose	8	0	0.050	1.0000	13	2	0.08	0.0031
GO:0010767	regulation of transcription from RNA polymerase II promoter in response to UV-induced DNA damage	5	0	0.030	1.0000	13	2	0.08	0.0031

GO:0046060	process	19	2	0.13	0.0072	19	0	0.12	1
GO:1900094	transcription from RNA polymerase II promoter involved in determination of left/right symmetry	38	0	0.26	1.0000	13	2	0.08	0.0031
GO:0061418	regulation of transcription from RNA polymerase II promoter in response to hypoxia regulation of	74	0	0.50	1.0000	13	2	0.08	0.0031
GO:0043619	regulation of transcription from RNA polymerase II promoter in response to oxidative stress	36	0	0.24	1.0000	13	2	0.08	0.0031
GO:0043618	regulation of transcription from RNA polymerase II promoter in response to stress	115	0	0.78	1.0000	13	2	0.08	0.0031
GO:0021920	regulation of transcription from RNA polymerase II promoter involved in spinal cord association neuron specification	5	0	0.030	1.0000	13	2	0.08	0.0031
GO:0021882	regulation of transcription from RNA polymerase II promoter involved in forebrain neuron fate commitment	9	0	0.060	1.0000	13	2	0.08	0.0031



Figure S1 Population structure of Atlantic Salmon in Placentia Bay, Newfoundland in (a) 2017
and (b) 2018. Red Harbour East (RHA) was found to be genetically differentiated from all other
rivers sampled based on the first two principal component (PC) axes from *pcadapt* (Luu et al.,
2017) using 139,038 SNPs. Rivers coloured east (yellow-red) to west (green-blue) with head of
the bay (purple). See Figure 1 and Table 1 for location details.





1313

1314 Figure S2 Heatmap of pairwise F_{ST} for Atlantic Salmon rivers (n = 25) in Placentia Bay,

1315 Newfoundland, Canada. The upper matrix represents 2017 and the lower matrix 2018. Rivers are

1316 ordered geographically from east to west. Lance (LAN) and Little Barasway (LBB) were not

1317 sampled in 2018. See Figure 1 and Table 1 for location details.

1318



1321Figure S3 Cross-validation (CV) error means and standard deviations from three ADMIXTURE1322runs using different random number seeds. Standard deviations which overlap the dashed red1323line, indicating lowest mean CV error represent a reasonable range of K. (a) 2017, K = 101324through K = 14 and (b) 2018, K = 9 through K = 11.





1327 Figure S4 ADMIXTURE (K = 9 - 11) results for Atlantic Salmon (*Salmo salar*) sampled in

1328 Placentia Bay, Newfoundland, Canada in (a) 2017 (n = 662) and (b) 2018 (n = 611). Rivers

1329 ordered geographically, east to west, around the bay; see Figure 1 and Table 1. Lance (LAN) and

1330 Little Barasway (LBB) were not sampled in 2018 due to limited sample size in 2017.







 \mathbb{R}^2 0 0.2 0.4 0.6 0.8 1.0 b) Homozgous Translocated vs. Heterozygous c) Homozgous Non-translocated vs. Homozygous Translocated

a) Homozgous Non-translocated vs. Heterozygous



- 1340 outlier block regions on chromosomes (Ssa01p and Ssa23). Pairwise LD (R^2) for (a) homozygous
- 1341 non-translocated vs. heterozygous, (b) homozygous translocated vs. heterozygous, and (c)
- 1342 homozygous non-translocated vs. homozygous translocated in 2017.





1345 Figure S7 Observed heterozygosity (Ho) for each karyotype. Karyotypes assigned using *kmeans*



- 1347 standard North American (homozygous translocated; Ssa01p/Ssa23 and Ssa01q), and
- 1348 heterozygous. All values calculated in plink v1.9. Lines represent smoothed values for a span of
- 1349 0.1 using ggplot2.
- 1350
- 1351
- 1352



Figure S8 Heterogeneity of (a, b) translocation (c - h) and karyotype frequencies between rivers
within year, (a, c, e, and g) 2017 and (b, d, f, and h) 2018 sampled. Bars represent confidence
intervals. Rivers ordered geographically, east to west, around Placentia Bay, Newfoundland; see
Figure 1 and Table 1 for location details.. Colours indicate karyotype; homozygous non-

1359 translocated (yellow), heterozygous (red), and homozygous translocated (blue-green).





Figure S9 Pairwise comparison of (a, b) translocation (c, d) and karyotype frequencies between
rivers within year, (a, c) 2017 (b, d) and 2018 sampled. Stars indicate significantly different
frequencies, calculated using a Fisher Exact Test adjusted for multiple comparisons. Rivers
ordered geographically, east to west, around Placentia Bay, Newfoundland; see Figure 1 and
Table 1 for location details.







1368 Figure S10 Temporal stability of (a) translocation (b) and karyotype frequencies within river

between years sampled (2017 and 2018). Rivers that exhibited a significant change in

1370 translocation or karyotype frequency between 2017 and 2018 highlighted. See Figure 1 and Table

1371 1 for location details.



Figure S11 Correlation between precipitation and the Ssa01p/Ssa23 chromosomal translocation
in Atlantic Salmon within Placentia Bay, Newfoundland, Canada. Linear regression of the first
principal component (PC) of a PCA based on 9 precipitation variables (BIOCLIM) and proportion
of non-translocated Ssa01p/q sampled in (a) 2017 and (b) 2018.