The aerial environment modulates plant responses to blue light

3

4 S. N. Innes, S. B. Jakobsen, A. Niday, H. Ali, L.E. Arve and S. Torre

5

6 Norwegian University of Life Sciences 7 Faculty of Biosciences 8 **Department of Plant Sciences** 9 P.O.BOX 5003 1432 Ås 10 11 Norway 12 Keywords: air humidity, blue light, chlorophyll, nitrogen, stomata, transpiration 13 14 15 Abstract The optimal amount of BL in the light spectrum varies dependent on plant species, 16 17 plant process, and the background environment. The aim of this study was to investigate plant responses to BL in different aerial environments. In controlled production systems, such as 18 greenhouses with reduced ventilation and air movement, high relative air humidity 19

(RH>85%) is common. Such an environment inhibits plant transpiration and nutrient uptake 20 and may have a negative impact on stomatal function and plant quality. In a number of 21 experiments, we investigated the response to BL in different air humidity regimes. The results 22 show that plants grown under high RH (90%) use BL more efficiently compared with those 23 grown under moderate RH (60%). At high RH, plant growth and leaf quality of basil (Ocimum 24 basilica), cucumber (Cucumis sativus) and tomato (Lycopersicon esculentum) improved with 25 increased amounts of BL ($5 \rightarrow 30\%$). We conclude that manipulation of BL can be used as a 26 27 cultivation strategy to improve plant productivity and quality in an environment with high RH. 28

29

30 INTRODUCTION

Blue light (BL) controls many processes important for plant productivity, such as morphology, stomatal function and patterning, stimulation of chlorophyll synthesis and photosynthetic capacity (Hogewoning et al. 2010; Islam et al. 2012; Terfa et al. 2012; Terfa et al. 2013). In Northern Europe, supplementary lighting is common in greenhouses during periods with
low natural radiation, and the dominating lamp type is high pressure sodium (HPS) with a low
amount of BL (5-8%). Increased BL (>20%) has been shown to improve plant quality and stomatal
function in different plant species (Islam et al. 2012; Terfa et al. 2012). Manipulating the amount of
BL could therefore be a useful strategy for controlling transpiration, growth and morphology of
plants.

In controlled production systems, such as greenhouses with reduced ventilation and air 40 41 movement, a high relative air humidity (RH>85%) is common. High RH inhibits plant transpiration 42 and nutrient uptake and can induce leaf yellowing and suppress growth and dry mass (DM) accumulation in some plants species (Gislerød et al. 1987; Gislerød & Mortensen 1990; Lihavainen et 43 44 al. 2016). Stomata of plants developed under high RH show reduced ability to respond to closing 45 signals such as darkness and drought, and are usually wide open during day and night (Arve et al. 46 2014; Torre et al. 2003). Several approaches to counteract the negative effect of high RH on stomatal 47 function have been tested, and daily temperature and/or RH variation, application of abscisic acid, high wind speed, longer periods with darkness and BL have been shown to improve stomatal 48 49 responsiveness to darkness (Fanourakis et al. 2016).

50 The aim of this study was to evaluate whether additional BL could be used as a cultivation 51 strategy to improve growth and quality of herbaceous species produced under high RH. Thus, we 52 tested the response to additional BL on transpiration. chlorophyll content, morphology and growth 53 in three common greenhouse species: basil (*Ocimum basilica*), cucumber (*Cucumis sativus*) and 54 tomato (*Lycopersicon esculentum*).

55

56 MATERIALS AND METHODS

57

58 Experimental set-up and plant material

59 Seeds of basil (Ocimum basilica 'Marian') from LOG A/S (Oslo, Norway) were sown in 12-cm pots in fertilized peat (Norway) in a greenhouse compartment with 20°C and 70% RH. After 14 days, 60 pots with germinated seedlings were placed in controlled growth chambers at 20°C, RH of either 60% 61 62 or 90% and ambient CO₂. Tomato (Lysopersicon esculentum 'Ailsa Craig') and cucumber (Cucumis sativus 'Quarto F1') were seeded in 12-cm pots as described above. When the first true leaves were 63 expanding, plants were placed in controlled growth chambers at 23°C with either 60% or 90% RH. 64 65 In the growth chambers, all plants received 20 h of light at a total of 200 μ mol m⁻² s⁻¹ 66 photosynthetically active radiation (PAR) from either HPS or a combination of HPS + BL (Figure 1), 67 and 4 h of darkness per day. Chambers with BL received 50 µmol m⁻² s⁻¹ from light emitting diodes 68 (LEDs) (400-500 nm, peak at 460 nm; Philips GreenPower LED module HF Blue) and 150 µmol m⁻² s⁻ ¹ from 400W HPS lamps (Gavita Superagro. Norway), as measured with a Li-COR LI 190 SA quantum 69 70 sensor (LI-COR Inc., USA). The amount of BL in the HPS+BL treatment was 30%, calculated by adding 71 the intensity of all wavelengths between 400 and 500 nm and then calculating the percentage of total 72 intensity between 400 and 700 nm (Figure 1). The plants were watered when needed with 50/50 73 mixture of YaraLiva® Calcinit[™] calcium nitrate solution (14.4% NO₃. 1.1% NH₄. 19.0% Ca. Yara Norge 74 AS. Oslo. Norway) and Kristalon[™] Indigo (7.5% NO₃. 1% NH₄. 4.9% P. 24.7% K. 4.2% Mg. 5.7% S. 75 0.027% B. 0.004% Cu. 0.06% Mn. 0.2% Fe. 0.004% Mo. 0.027% Zn. Yara Norge AS. Oslo. Norway). 76 Electrical conductivity was 1.5 mS cm⁻¹.





78

Figure 1. Light spectra of the lamps used in the experiments. High pressure sodium (HPS) lamps
(Osram NAV T-400W), green line; HPS + Blue Light (BL) emitting diodes (Philips GreenPower LED
module HF Blue), blue line.

82

83 Growth analysis, leaf color, chlorophyll and nutrient content

The growth and leaf color of basil were evaluated after 4 weeks of growth according to a scale from 3 to 0, where 3 = no visible yellowing, 2 = yellow spots in-between the veins and 1 = severe leaf yellowing. Growth of tomato and cucumber was evaluated after 3 weeks. The number of leaves (>1cm) was counted, and the total length was measured from the base of the shoot to the shoot apical meristem. Dry weight was determined after drying for 5 d at 70°C. Leaf area was measured with a leaf area meter (Li-3100, Li-Cor Inc.) The relative chlorophyll content was measured with a handheld
chlorophyll content meter (model CL-0.1, Hansatech Instruments Ltd, UK).

91

92 Transpiration measurements

93 Leaf transpiration was measured at the end of the experimental period on fully expanded 94 leaves using a porometer (AP4, Delta-T Devices Ltd., Cambridge, UK). The measurements were 95 conducted in the middle of the dark period and 1-2 h after the light was turned on. Epidermal 96 impressions were made of fresh, intact, fully expanded leaves from tomato and cucumber by Suzuki's 97 universal micro-printing (SUMP) method using SUMP liquid and SUMP plate B (SUMP Laboratory, Tokyo, Japan) as described previously (Tanaka et al. 2005). All samples were taken interveinally 98 99 close to the midrib on the abaxial side (tomato) or both abaxial and adaxial sides (cucumber). The 100 copied SUMP images were observed under a light microscope, and the number of stomata was 101 counted with UTHSCSA ImageTool for Windows version 3.00 (University of Texas Health Science 102 Centre, San Antonio, TX, USA).

103

104 Statistics

Significant differences between means were tested for normally distributed data using
 general linear models (GLM) and Tukey's test. Differences with p<0.05 were considered significantly
 different. All statistical tests were performed in Minitab 16.1.1 (Windows version, State College, PA,
 USA).

109

110 **RESULTS**

111 At moderate RH, no signs of chlorosis or black spots on the basil leaves were observed. However, the plants grown in high RH with HPS developed severe leaf yellowing (Table 1). The 112 113 symptoms first appeared as yellow spots in-between the veins, and on some plants the entire leaf 114 turned yellow and small black spots appeared on the leaf surface. Additional BL improved the leaf quality of basil in high RH; the yellowing was less severe and the chlorophyll content increased 115 significantly (Table 1). At moderate RH, rather small effects were observed when the plants received 116 117 more BL compared to HPS alone, although the plants were slightly shorter and had more chlorophyll (Table 1). At moderate RH, BL reduced internode lengths but plants developed the same number of 118 119 leaves as with HPS alone. However, under high RH the stem length increased with HPS+BL but the 120 plants had a higher number of leaves compared to those grown with HPS alne (Table 1).

Table1. Effects of additional blue light (BL) on growth and leaf quality of basil grown at high (90%)
and moderate (60%) RH. Visible leaf quality was evaluated according to a scale from 3 to 0 where 3
= no visible yellowing, 2 = yellow spots in-between the veins and 1 = severe leaf yellowing (n=10).
HPS, High-pressure sodium lamps.

	Moderate RH (60 %)		High RH (90 %)	
	HPS + BL	HPS	HPS + BL	HPS
Plant height (cm)	25.00±1.21 ^{ab}	27.55±0.91ª	23.10±0.65b	20.12±0.33c
Number of leaves	12.80±0.27 ^a	12.10±0.22 ^a	12.00±0.25ª	10.80±0.23b
Relative chlorophyll content	14.12±0.42ª	13.00±0.43ª	11.03±0.33 ^b	7.74±0.41 ^c
Leaf quality (0-3)	3.0	3.0	2.8	1.6

127

128 129 The suppressed growth and leaf unfolding rate observed in basil at high RH with HPS was not observed in tomato or cucumber but a tendency of leaf yellowing and a lower chlorophyll content 130 131 were found (Tables 2 and 3). Furthermore, BL inhibited plant height and growth under both RH 132 regimes but the effect was much stronger at moderate RH compared to high RH (Tables 2 and 3). In tomato grown at moderate RH, additional BL reduced plant height and total dry weight by 39 and 133 35%, respectively. However, with high RH the reduction in height and dry weight was only 22 and 134 13% (Table 2). Similarly, in cucumber grown at moderate RH, additional BL reduced plant height and 135 total dry weight by 40 and 20%, respectively, while at high RH the reduction was only 20 and 2% 136 137 (Table 3). Number of leaves followed a similar trend in both species (Tables 2 and 3). The number of fruits per cucumber plant and average fruit length were significantly larger with moderate RH and 138 HPS alone and with high RH and HPS + BL than with moderate RH and HPS + BL and with high RH 139 140 with HPS alone (Table 3). 141

142

Table 2. Growth and morphology of tomato grown under high (90%) and moderate (60%) RH with

the traditional high-pressure sodium (HPS) lamp (200 μ molm⁻²s⁻¹) and HPS + blue LED (BL) (150 +

145 50 μ molm⁻²s⁻¹).

	Moderate RH (60 %)		High RH (90 %)	
	HPS + BL	HPS	HPS + BL	HPS
Plant height (cm)	16.50±0.58°	27.0±1.03ª	20.94±0.65 ^b	27.38±0.79ª
Number of leaves	7.50±0.27 ^b	8.63±0.32ª	8.25 ± 0.25^{ab}	9.13±0.23ª
Total dry weight (g)	2.82±0.24 ^b	4.3 ± 0.47^{a}	3.84 ± 0.23^{ab}	4.45±0.20ª
Relative chlorophyll content	23.12 ± 1.07^{a}	17.90±0.63 ^b	23.89±1.22ª	17.44±0.92 ^b
Stomata number (50 μm²)	14.00 a	14.92 a	13.51 ab	12.81 b
Different letters in the same ro	ow indicate sig	nificant differen	ces at p<0.05 (n	=8).
Table 3. Growth and morphol	ogy of cucum	oer grown unde	er high (90%) a	nd moderate
with the traditional high-pres	sure sodium (HPS) lamp (200) µmolm ⁻² s ⁻¹) an	nd HPS + blue
(150 + 50 μmolm ⁻² s ⁻¹).				

	Moderate RH (60 %)		High RH (90 %)	
	HPS + BL	HPS	HPS + BL	HPS
Plant height (cm)	35.0±1.16°	59.38±1.27ª	47.81±1.84b	62.81±1.86 ^a
Number of leaves	10.63±0.18 ^c	11.25±0.16 ^b	11.88±0.35ª	11.38±0.26 ^b
Dry weight (g)	6.31±0.12 ^c	7.84±0.26ª	6.86±0.21 ^{bc}	7.02±0.29 ^b
Fruit number per plant	7.40±0.25 ^b	9.00±0.32ª	9.00±0.32ª	7.20±0.59 ^b
Fruit length (cm)	2.45±0.07 ^b	2.60 ± 0.15 ab	3.10±0.19ª	1.30±0.04 ^c
Relative chlorophyll content	28.26±1.99ª	20.55±0.75 ^b	25.93±0.96ª	17.55±0.80 ^b
¹ Stomata number (50 μm²)	19.73 c	12.75 a	15.45 b	13.72 a

154 Different letters in the same row indicate significant differences at p<0.05 (n=8).

- 155 ¹merged stomata count adaxial and abaxial side
- 156
- 157



- . . .
- 164



Figure 2. Transpiration rate (μ molm⁻²s⁻¹) of basil leaves developed under 60% and 90% RH with HPS (200 μ mol m⁻²s⁻¹) or with HPS + blue LED lamps (150 + 50 μ molm⁻²s⁻¹). Different letters within day and night indicate significantly different values. N=5. Mean ± SE.

- 169
- 170

The transpiration rate of the three species was affected differently by BL dependent on the RH background. BL increased transpiration significantly at both moderate and high RH (p<0.05) in cucumber (results not shown).

174 The ratio between transpiration rates during day and night was calculated to compare the 175 responsiveness to darkness as a signal for closure (Table 4). In basil, the main difference in day/night transpiration rate ratio was found between moderate and high RH, but no significant difference was 176 177 found between any of the treatments (Table 4). However, in cucumber and tomato a trend towards 178 an increased day/night transpiration was observed at high RH when additional BL was added during 179 the day, but the data was not statistically different (Table 4). Cucumber and tomato grown under high 180 RH with HPS alone had a day/night transpiration rate ratio close to 1, which indicates almost no 181 stomatal movement in response to darkness (Table 4).

- 182
- 183

Table 4. Ratio between day and night transpiration rate for basil, tomato and cucumber grown under
high (90%) and moderate (60%) RH grown with the traditional high-pressure sodium (HPS) lamp
(200 μmolm⁻²s⁻¹) and HPS + blue LED (BL) (150 + 50 μmolm⁻²s⁻¹). Transpiration was measured with
a porometer (see Materials and Methods for details).

188

	Moderate RH (60 %)		High RH (90 %)		
	HPS + BL	HPS	HPS + BL	HPS	
Basil	1.45±0.12ª	1.48±0.19ª	2.07±0.47ª	2.01±0.50ª	
Tomato	1.51 ± 0.14^{a}	1.54±0.12ª	1.27 ± 0.15^{ab}	1.15±0.05 ^b	
Cucumber	1.54±0.05 ^{ab}	1.82±0.15ª	1.24 ± 0.05^{bc}	1.05±0.17°	

189 Different letters in the same row indicate significant differences at p<0.05 (n=8-10).

190

191

192 DISCUSSION

193

194 Additional blue light improves leaf quality and growth in high RH

Plant production under high RH (> 85%) is common during periods when ventilation isavoided to save energy. This is usually also the time when supplementary lighting is required to

improve growth and yield of greenhouse crops at northern latitudes. In this study, we demonstrated
that addition of BL increased chlorophyll content under both high and moderate RH in all three
species, basil, cucumber and tomato (Tables 1-3), and improved growth and leaf quality under high
RH. Thus, additional BL is a useful cultivation strategy for improving leaf quality and productivity
under high RH.

202 The reason for leaf yellowing at high RH is proposed to be related to reduced transpiration 203 and nutrient deficiencies (Gislerød et al. 1987; Gislerød & Mortensen 1990; Mortensen & Gislerød 204 1989). Reduced chlorophyll content is often connected to deficiencies in Mg, Fe or N (Engels et al. 205 2012). It is likely that the leaf yellowing observed in plants produced under high RH and HPS is due 206 to insufficient N uptake and that the BL improves N uptake by increasing transpiration. However, 207 different plant species may respond differently to BL and/or the BL may work via different 208 mechanisms to increase chlorophyll content and improve leaf quality. The increased chlorophyll 209 content found in leaves exposed to additional BL could also be due to a direct effect on chlorophyll 210 biosynthesis. Senger and Bauer (1987) showed that plants grown under supplementary BL 211 fluorescent lamps had higher Chl *a/b* ratios and more sun-like type chloroplasts than plants exposed to less BL. Furthermore, higher Chl content was reported in cucumber and roses produced with an 212 213 increased proportion of BL and points towards a photosynthetic apparatus better adapted to high 214 light levels (Evans 1987; Hogewoning et al. 2010; Terfa et al. 2013).

215 The effect of interaction between temperature and light quality on growth and morphology 216 has been the subject in many studies (Bergstrand et al. 2016; Moe et al. 2002). However, less 217 attention has been paid to the aerial environment and its interaction with light quality. In this study, additional BL reduced stem elongation and DM accumulation more strongly under moderate RH than 218 219 under high RH (Tables 2 and 3). BL is involved in inhibition of growth of internodes and cell expansion or division (Dougher & Bugbee 2004; Folta et al. 2003). Furthermore, dry air (large vapor 220 221 pressure deficit) is also known to be an abiotic stressor that induces stomatal closure and reduces 222 growth and stem elongation in herbaceous plant species (Zhang et al. 2015). It has been well 223 described in other growth studies that exposure to more than one stressor at the same time can have a synergistic effect on the growth response (Murali & Teramura 1985). However, the reason why BL 224 225 confers stronger growth inhibition at moderate RH compared to high RH is not clear.

226

227 Additional BL increases transpiration but dependent on RH and plant species

Increased transpiration could be due to a higher number of stomata or an increased stomatalaperture. BL is known to promote both stomatal opening and stomata number (Terfa et al. 2013). In

230 the present study. BL increased transpiration in basil, tomato and cucumber but the strength of the 231 response varied with species and RH regime (Figure 2; Table 4). However, cucumber showed the 232 strongest response to BL, and a significant increase in transpiration was found under both moderate 233 and high RH when more BL was added (data not shown). Furthermore, a significantly larger number 234 of stomata was found in cucumber on the upper and lower sides of the leaves when exposed to additional BL, as described earlier by Hogewoning et al. (2010). The increased number of stomata in 235 236 cucumber in response to BL may explain the stronger effect on transpiration in this species compared 237 to tomato. However, the day/night transpiration ratio increased in cucumber and tomato produced under high RH when BL was added, indicating an improved stomatal closure in darkness (Table 4). 238 Previous experiments with pot roses (Rosa x hybrida) also showed that light with a higher proportion 239 240 of BL than provided by the traditional HPS lamp improved dark-induced stomata closure and tolerance to drought (Terfa et al. 2012). On the contrary, cucumber and tomato grown under high RH 241 242 with HPS alone had a day/night transpiration ratio close to 1.0, which indicates almost no stomatal 243 movement in response to darkness. The reason for the lack of stomatal movement under high RH and 244 HPS alone is not clear but could be due to a higher accumulation of starch in the guard cells. In a study with silver birch, increased starch accumulation and a higher C/N ratio was found in leaves 245 246 developed under high RH compared to ambient RH levels (Lihavainen et al. 2016), and similar results 247 have been obtained in *Hydrangea macrophylla* grown under high RH compared to moderate RH (S. 248 Torre, unpublished data). Starch degradation in guard cells has an important role in plant growth by 249 driving stomatal responses to light. Also, this degradation has been shown to be controlled by the 250 phototropin-dependent blue-light receptor (Horrer et al. 2016). The fact that additional BL increased 251 the ratio between day and night transpiration rates under high RH opens up the possibility that BL 252 triggers stomatal function under high RH through starch degradation as described in Horrer et al. (2016) but further research is needed to confirm this theory. 253

254

255 CONCLUSION

When the proportion of BL increased from 5 to 30% under moderate RH (<60%), a strong growth
inhibition and a significant reduction in dry weight was found in tomato and cucumber. However,
under high RH, plant growth and quality was improved with increased amounts of BL (30 vs. 5%).
We conclude that manipulation of BL can be used as a cultivation strategy to improve plant
production and quality under high RH.

261

262 ACKNOWLEDGEMENTS

- 263 We would like to thank Ida Kristin Hagen for excellent help in taking care of the plants throughout
- the experiments. This research was supported by the Norwegian Research Council, "Bioeconomic
- 265 production of fresh greenhouse vegetables in Norway" project number 255613/E50.
- 266

267 **REFERENCES**

- Arve, L. E., Carvalho, D. R. A., Olsen, J. E. & Torre, S. (2014). ABA induces H2O2 production in guard
 cells, but does not close the stomata on *Vicia faba* leaves developed at high air humidity.
 Plant Signaling & Behavior, 9 (7): e29192.
- Bergstrand, K.-J., Mortensen, L. M., Suthaparan, A. & Gislerød, H. R. (2016). Acclimatisation of
 greenhouse crops to differing light quality. *Scientia Horticulturae*, 204: 1-7.
- Dougher, T. A. O. & Bugbee, B. (2004). Long-term blue light effects on the histology of lettuce and
 soybean leaves and stems. *Journal of the American Society for Horticultural Science*, 129 (4):
 467-472.
- Engels, C., Kirkby, E. & White, P. (2012). Chapter 5 Mineral Nutrition, Yield and Source–Sink
 Relationships A2 Marschner, Petra. In *Marschner's Mineral Nutrition of Higher Plants*(*Third Edition*), pp. 85-133. San Diego: Academic Press.
- Evans, J. (1987). The dependence of quantum yield on wavelength and growth irradiance.
 Functional Plant Biology, 14 (1): 69-79.
- Fanourakis, D., Bouranis, D., Giday, H., Carvalho, D. R. A., Rezaei Nejad, A. & Ottosen, C.-O. (2016).
- Improving stomatal functioning at elevated growth air humidity: a review. *Journal of Plant Physiology*, 207: 51-60.
- Folta, K. M., Lieg, E. J., Durham, T. & Spalding, E. P. (2003). Primary inhibition of hypocotyl growth
 and phototropism depend differently on phototropin-mediated increases in cytoplasmic
 calcium induced by blue light. *Plant Physiology*, 133 (4): 1464.
- Gislerød, H. R., Selmer-Olsen, A. R. & Mortensen, L. M. (1987). The effect of air humidity on nutrient
 uptake of some greenhouse plants. *Plant and Soil*, 102 (2): 193-196.
- Gislerød, H. R. & Mortensen, L. M. (1990). Relative humidity and nutrient concentration affect
 nutrient uptake and growth of *Begonia × hiemalis. HortScience*, 25 (5): 524-526.
- Hogewoning, S. W., Trouwborst, G., Maljaars, H., Poorter, H., van Ieperen, W. & Harbinson, J. (2010).
 Blue light dose-responses of leaf photosynthesis, morphology, and chemical composition of
 Cucumis sativum grown under different combinations of red and blue light. *Journal of*
- 294 *Experimental Botany*, 61 (11): 3107-3117.

295 Horrer, D., Flütsch, S., Pazmino, D., Matthews, Jack S. A., Thalmann, M., Nigro, A., Leonhardt, N., 296 Lawson, T. & Santelia, D. (2016). Blue light induces a distinct starch degradation pathway in 297 guard cells for stomatal opening. *Current Biology*, 26 (3): 362-370. 298 Islam, M. A., Kuwar, G., Clarke, J. L., Blystad, D.-R., Gislerød, H. R., Olsen, J. E. & Torre, S. (2012). 299 Artificial light from light emitting diodes (LEDs) with a high portion of blue light results in 300 shorter poinsettias compared to high pressure sodium (HPS) lamps. Scientia Horticulturae, 147: 136-143. 301 302 Lihavainen, J., Ahonen, V., Keski-Saari, S., Kontunen-Soppela, S., Oksanen, E. & Keinanen, M. (2016). 303 Low vapour pressure deficit affects nitrogen nutrition and foliar metabolites in silver birch. Journal of Experimental Botany, 67 (14): 4353-65. 304 305 Moe, R., Morgan, L. & Grindal, G. (2002). Growth and plant morphology of Cucumis sativus and Fuchsia x hybrid are influenced by light quality during the photoperiod and by diurnal 306 307 temperature alternations: International Society for Horticultural Science (ISHS), Leuven, 308 Belgium. 229-234 pp. 309 Mortensen, L. M. & Gislerød, H. R. (1989). Effect of CO2;, air humidity, and nutrient solution concentration on growth and transpiration of *Begonia X hiemalis* Fotsch. *Die* 310 311 *Gartenbauwissenschaft*, 54 (4): 184-189. 312 Murali, N. S. & Teramura, A. H. (1985). Effects of ultraviolet-B irradiance on soybean. VI. Influence of 313 phosphorus nutrition on growth and flavonoid content. *Physiologia Plantarum*, 63 (4): 413-314 416. Senger, H. & Bauer, B. (1987). The influence of light quality on adaptation and function of the 315 photosynthetic apparatus. *Photochemistry and Photobiology*, 45: 939-946. 316 317 Tanaka, Y., Sano, T., Tamaoki, M., Nakajima, N., Kondo, N. & Hasezawa, S. (2005). Ethylene inhibits abscisic acid-induced stomatal closure in *Arabidopsis*. *Plant Physiology*, 138 (4): 2337-2343. 318 319 Terfa, M. T., Poudel, M. S., Roro, A. G., Gislerød, H. R., Olsen, J. E. & Torre, S. (2012). Light emitting 320 diodes with a high proportion of blue light affects external and internal quality parameters of pot roses differently than the traditional high pressure sodium lamp: International Society for 321 322 Horticultural Science (ISHS), Leuven, Belgium. 635-642 pp. 323 Terfa, M. T., Solhaug, K. A., Gislerød, H. R., Olsen, J. E. & Torre, S. (2013). A high proportion of blue light increases the photosynthesis capacity and leaf formation rate of *Rosa x hybrida* but 324 325 does not affect time to flower opening. *Physiol Plant*, 148 (1): 146-59.

- 326 Torre, S., Fjeld, T., Gislerød, H. R. & Moe, R. (2003). Leaf anatomy and stomatal morphology of
- 327 greenhouse roses grown at moderate or high air humidity. *Journal of the American Society*328 *for Horticultural Science*, 128 (4): 598-602.
- 329 Zhang, D., Zhang, Z., Li, J., Chang, Y., Du, Q. & Pan, T. (2015). Regulation of vapor pressure deficit by
- greenhouse micro-fog systems improved growth and productivity of tomato via enhancing
 photosynthesis during summer season. *PLoS One*, 10 (7): e0133919.