

An explorative study of *Enterococcus faecalis* transcriptional responses to infection-relevant growth environments

Et eksplorativt studium av de transkripsjonelle responsene til *Enterococcus faecalis* under infeksjonsrelevante vekstbetingelser

Philosophiae Doctor (PhD) Thesis

Heidi Cecilie Vebø

Laboratory for Microbial Gene Technology
Dept. of Chemistry, Biotechnology and Food Science
Norwegian University of Life Sciences

Ås 2010



Thesis number 2010:10
ISSN 1503-1667
ISBN 978-82-575-0922-4

TABLE OF CONTENTS

ACKNOWLEDGEMENT	iii
SUMMARY	v
SAMMENDRAG	vi
LIST OF PAPERS	vii
1. INTRODUCTION	1
1.1 The Enterococcus genus	1
1.1.1 <i>Enterococcus faecalis</i>	3
1.2 Antibiotics and antibiotic resistance in enterococci	6
1.2.1 Erythromycin resistance in enterococci	7
1.3 Enterococcal infections	8
1.3.1 Urinary tract infection	10
1.3.2 Bacteremia	10
1.3.3 Other enterococcal infections	11
1.4 Enterococcal virulence factors	11
1.4.1 Secreted virulence factors	12
1.4.2 Cell envelope bound virulence factors	13
1.4.3 Other potential virulence factors	14
1.5 DNA microarray technology	15
1.5.1 Experimental design and data analysis	15
1.5.2 DNA microarray technology in the future	17
2. OBJECTIVES OF THIS STUDY	18
3. MAIN RESULTS AND DISCUSSIONS	19
4. MAIN CONTRIBUTIONS AND FUTURE WORK	24
5. REFERENCES	26

PAPER I-III

ACKNOWLEDGEMENT

The work presented in this thesis was carried out during the period 2002-2010 at the Laboratory for Microbial Gene Technology, Department of Chemistry, Biotechnology and Food Science, Norwegian University of Life Sciences. This work was supported by a grant from the Research Council of Norway.

First, I would like to thank my supervisor Prof. Ingolf F. Nes for having me in his group and for giving me this chance to do a PhD. I also want to thank you for always keeping your door open. You took your time to talk to me about problems regarding the project, new ideas, silly questions or just a social chat, and I'm really grateful for that. I also want to direct a special thanks to my co-supervisors Dr. Ågot Aakra and Dr. Dag. A. Brede. Ågot, thank you for introducing me to the field of microarrays and for the supervision during the first part of my PhD. Dag, you helped me when everything looked impossible, I couldn't have done this without you! You always had time for all my questions and shared your endless knowledge with me. Thank you for everything I have learned from you!

I want to thank Dr. Lars Snipen for all the help with statistics and for doing all the analysis of the microarray data. I also want to thank Margrete Solheim for the good collaboration during the work on paper III, for arranging so many social events for our group, and for being a really good friend. Thank you Daniel Straume for all the ups and downs we have shared, for helping me during the writing process and for being a very good friend. To all the members of the LMG group; I want to thank you all for contributing to a great working environment, for all discussion we've had and for all the laughs we have shared!

Finally, I want to thank my family and friends for their love, support and encouragement during my education. Oliver, I could never have done this without you. You have been standing by my side in all the ups and downs; I can never thank you enough! Thank you so much for taking such good care of Nina while I was working, you have really pulled your load and I'm looking forwards to us being a family again. Nina, finally you can play with your mum again. Oliver and Nina you mean the world to me!

SUMMARY

Enterococcus faecalis is commonly found in the gastrointestinal tract of humans. It is also an opportunistic pathogen and one of the leading causes of nosocomial infections worldwide. The treatment of *E. faecalis* infections is often difficult due to its innate and acquired antibiotic resistance. Several functional studies have been performed to link genetic traits to antibiotic resistance and virulence. However, there are still many unanswered questions regarding the importance of virulence factors, and the global regulation of these and other genes in the pathogenicity of this bacterium. Therefore, a more comprehensive understanding of the global gene regulation of *E. faecalis* in response to different growth environments is needed, as well as an understanding of how this gene regulation affects the establishment of an infection.

This thesis focuses on the importance of *E. faecalis*' gene regulation in response to the infection-relevant growth conditions blood, urine and exposure to erythromycin. The effect of a small percentage (10%) of blood on the transcriptome of *E. faecalis*, as well as the different transcriptional profiles of three diverse *E. faecalis* isolates grown in urine were also examined.

The work in this thesis showed that erythromycin inhibits the growth of the resistant V583 strain and affects the transcription of several genes, but not the constitutively expressed *erm(B)* gene that confers erythromycin resistance. Furthermore, we discovered that growth in blood and urine resulted in several similar transcriptional responses despite the use of four diverse isolates. Important responses to blood and urine include a reduced transcription of genes involved in the production of cell wall polysaccharides, enhanced transcription of several stress genes, including many involved in oxidative stress, and enhanced transcription of genes encoding iron- and manganese-uptake systems. Each environment also activated a unique subset of genes, an effect that was particularly evident for genes related to metabolism. The enhanced transcription of *lrgAB* was unique to growth in blood. The function of these two genes is so far unknown in *E. faecalis* and could be interesting to study more closely. Growth in urine revealed a strain specific enhanced regulation of the virulence related *fsr*-operon, which could give an indication to the isolates pathogenic potential. All three studies revealed that the modulation of the *E. faecalis* cell envelope is important for the adaptation to growth in these three infection-relevant environments.

SAMMENDRAG

Enterococcus faecalis finnes ofte som en del av menneskets tarmflora. Den er også en opportunistisk patogen, og er en av hovedårsakene til sykehusinfeksjoner verden over. Behandlingen av *E. faecalis* infeksjoner er ofte vanskelige på grunn av bakteriens nedarvede og tilegnede antibiotikaresistens. Mange funksjonelle studier har blitt utført for å koble genetiske egenskaper mot antibiotikaresistens og virulens. Imidlertid er det fortsatt ukjent hvor stor betydning antatte virulens faktorer, og den globale reguleringen av disse og andre gener, faktisk har for denne bakteriens patogenisitet. Derfor er det behov for en mer inngående forståelse av hvordan den globale genreguleringen i *E. faecalis* responderer på ulike vekstbetingelser, og hvordan denne genreguleringen influerer etableringen av infeksjoner.

Denne avhandlingen fokuserer på hvordan genreguleringen i *E. faecalis* påvirkes av de infeksjonsrelevante vekstbetingelsene blod, urin og vekstmedium tilsatt antibiotikumet erythromycin. Effekten av en liten prosentandel (10%) blod på transkripsjonen, samt forskjeller i transkripsjonsprofilene til tre ulike *E. faecalis* isolater dyrket i urin ble også undersøkt.

Arbeidet i denne avhandlingen viste at erythromycin hemmer veksten av den resistente V583-stammen og påvirker transkripsjonen av mange gener, men ikke det konstitutivt uttrykte erythromycinresistensgenet *erm(B)*. Videre oppdaget vi at vekst i blod og urin ga mange liknende transkripsjonsprofiler, selv om fire ulike isolater ble undersøkt. Viktige responser i blod og urin omfatter redusert transkripsjon av gener involvert i produksjon av celleveggpolysakkarider, økt transkripsjon av mange stress-gener blant annet flere involvert i oksidativt stress og økt transkripsjon av gener som koder for jern- og mangan-opptakssystemer. Begge vekstbetingelsene aktiverer også et unikt sett med gener, og denne effekten var spesielt tydelig for gener relatert til metabolisme. En unik respons ved vekst i blod var en økt transkripsjon av *lrgAB*, to gener med hittil ukjent funksjon i *E. faecalis* som kan være interessante å studere videre. Ved vekst i urin fant vi et stammespesifikt uttrykk av det virulensrelaterte *fsr*-operonet, noe som dermed kan gi en indikasjon på isolatets patogene potensial. Alle tre studiene viste at forandringer i celleveggen/cellemembranen til *E. faecalis* er viktig for tilpasningen til vekst i disse tre infeksjonsrelevante betingelsene.

LIST OF PAPERS

Paper I

Aakra, Å., Vebø, H., Snipen, L., Hirt, H., Aastveit, A., Kapur, V., Dunny, G., Murray, B. E. & Nes, I. F. (2005). Transcriptional Response of *Enterococcus faecalis* V583 to Erythromycin. *Antimicrobial Agents and Chemotherapy*, 49:2246-2259.

Paper II

Vebø, H. C., Snipen, L., Nes, I. F. & Brede, D. A. (2009). The Transcriptome of the Nosocomial Pathogen *Enterococcus faecalis* V583 Reveals Adaptive Responses to Growth in Blood. *PLoS ONE*, 4(11):e7660.

Paper III

Vebø, H. C., Soheim, M., Snipen, L., Nes, I. F. & Brede, D. A. Comparative genomic analysis of pathogenic and probiotic *Enterococcus faecalis* isolates, and their transcriptional responses to growth in human urine. (Manuscript)

List of related papers by the author, not included in the thesis

Berget, I., Mevik, B.-H., Vebø, H., Næs, T. (2005). A strategy for finding relevant clusters; with an application to microarray data. *Journal of Chemometrics*, 19(9):482-491.

Solheim, M., Aakra, Å., Vebø, H., Snipen, L. & Nes, I. F. (2007). Transcriptional Responses of *Enterococcus faecalis* V583 to Bovine Bile and Sodium Dodecyl Sulfate. *Applied and Environmental Microbiology*, 73(18):5767-74.

Aakra, Å., Vebø, H., Indahl, U., Snipen, L., Gjerstad, Ø. Hjorthaug, H. S., Lunde, M. & Nes, I. F. The response of *Enterococcus faecalis* V583 to chloramphenicol treatment. (Submitted)

1. INTRODUCTION

This thesis describes the use of microarrays as a tool to study the transcriptional responses of *Enterococcus faecalis* when exposed to different infection-relevant growth environments such as blood, urine, and the antibiotic erythromycin. This organism is among the leading causes of hospital acquired infections, which are often difficult to treat due to high level antibiotic resistance [12, 98]. Hence, it is of great importance to understand how this opportunistic pathogen responds and adapts to growth in infectious environments in order to gain further knowledge about its virulence, and to come up with new therapeutic strategies to be able to treat infections caused by *E. faecalis*.

1.1 The *Enterococcus* genus

The term “enterococcal group” was first used by Sherman [137] to describe group D streptococci that grew at 10 and 45°C, and could survive heating at 60°C for 30 minutes. Furthermore, this group grew in environments with broad pH values and could tolerate high salt concentrations (6.5% NaCl) [53]. Although this group of bacteria, described the first time in 1899 [159] was different from the other members of the *Streptococcus* genus, a separate *Enterococcus* genus was not established until almost a century later, in 1984 [130]. The *Enterococcus* genus now comprises of 40 different species [32], and an overview of some of the enterococcal species, and their relatedness to each other is shown in the phylogenetic tree in Figure 1.

Enterococci are Gram-positive, catalase-negative cocci which most often occur in pairs or short chains. They are found in several different environments such as soil, food and in the intestinal flora of most birds and mammals including humans. However, they are also opportunistic pathogens often involved in nosocomial infections. Enterococci are facultative anaerobes with an optimum growth temperature of 35°C and can utilize a wide variety of nutrients such as diverse carbohydrates, arginine, agmatine, many α -keto acids, glycerol, lactate, malate and citrate [56]. Under most conditions enterococci, like other lactic acid bacteria perform fermentative carbohydrate metabolism with lactate as the major end product [75]. It has also been demonstrated that certain enterococci are able to perform respiration utilizing oxygen as terminal electron acceptor in the presence of heme [127]. Their versatile

metabolism combined with their tolerance of extremely harsh growth conditions most likely enhances the enterococcal competitiveness in diverse environments.

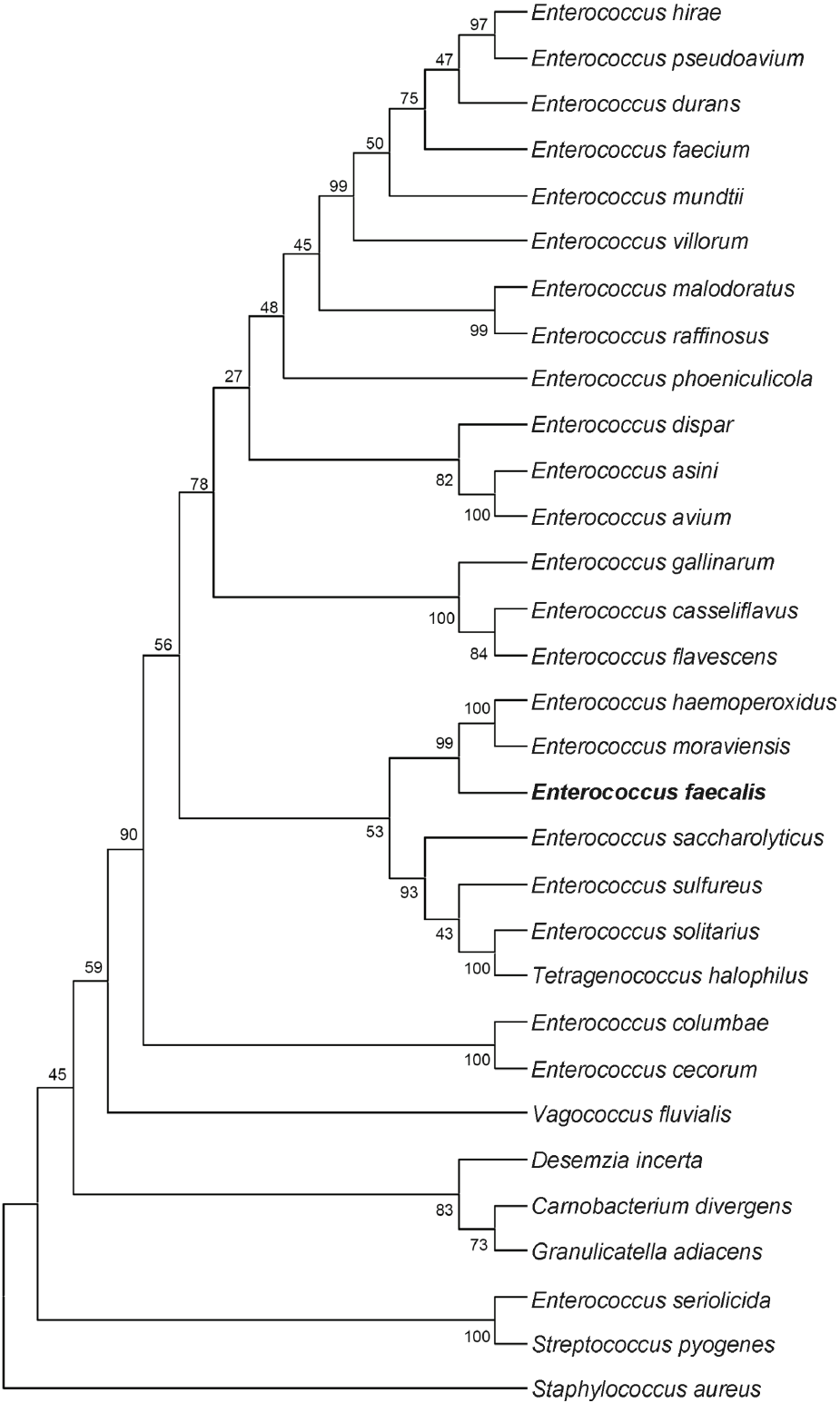


Figure 1: Unrooted phylogenetic tree based on 16S rRNA gene sequences, showing the relationship between *Enterococcus faecalis* and other enterococci and lactic acid bacteria. The sequence of the 16S rRNA gene of *Staphylococcus aureus* was used as an outgroup. (Modified from [74])

1.1.1 *Enterococcus faecalis*

The name *Streptococcus faecalis* was first used by Andrewes and Horder in 1906 to identify an organism of faecal origin [2]. *Enterococcus faecalis* and *Enterococcus faecium* were the first two enterococcal species suggested to comprise the new *Enterococcus* genus [130]. Both species can be found throughout the gastro intestinal (GI) tract in humans, although in low numbers in the oral cavity [85]. *E. faecalis* is normally found in higher numbers (10^5 - 10^7 per gram feces) in the GI tract than *E. faecium* (10^4 - 10^5 per gram feces) [12]. Although often found in the same habitats, there are some phenotypic characteristics that separate these two species (Table 1). Based on these differences there are several rapid test such as API Rapid ID 32 [38] that separates *E. faecalis* from other enterococci by 88.6% accuracy, and *E. faecium* by 67% accuracy [167].

Table 1: Differentiation of *E. faecalis* and *E. faecium* (modified from [130])

Characteristics	<i>E. faecalis</i>	<i>E. faecium</i>
Acid produced from:		
Arabinose	-	+
Tagatose	+	-
Energy from:		
Pyruvate	+	-
Citrate	+	-
Malate	+	-
Serine	+	-
Tolerance to:		
0.04% Tellurite	+	-
0.01% Tetrazolium	+	-

One enterococcal strain intensively studied in this thesis is the V583 strain, since it was the first fully sequenced *E. faecalis* strain [112]. Its chromosome contains 3093 genes, and more than 25% of these genes are within mobile elements such as prophages, conjugative transposons and integrated plasmid remnants [112]. Furthermore, it contains three plasmids, the pAD1-like pTEF1, the pCF10-like pTEF2 and pTEF3 that belongs to the family of pAM^β1 broad host range plasmids [112]. V583 was also the first vancomycin resistant clinical isolate reported in the United States [129], and is resistant to several other antibiotics. This strain was isolated from a patient suffering from a persistent bloodstream infection, and is

included in the clonal complex 2 consisting mainly of infection derived isolates [128]. Another member of the clonal complex 2 studied in this thesis is the MMH594 strain. This was the first *E. faecalis* strain reported to contain a complete *E. faecalis* pathogenicity island (PAI) [134]. It was involved in a hospital outbreak in the middle of the 1980s, causing more than 30 infections and with an elevated mortality rate compared to infections by other nosocomial *E. faecalis* strains [60].

The important laboratory *E. faecalis* strain, OG1RF was also studied in this thesis. It has commonly been used for molecular manipulations and virulence studies, and its sequence was published recently [4]. The sequencing of this strain revealed that it contains only a few mobile elements. This has been suggested to be due to the presence of two CRISPR (comprised of regularly interspaced short palindromic repeats) loci and the *cas*-genes cognate to one of the two CRISPR loci, which prevent establishment of invading phages or plasmids ([4] and references therein). It has 39 unique regions comprising 127 new genes compared to the V583 genome, and OG1RF does not contain a PAI like the two above mentioned strains. Finally, the fourth strain that has been studied more closely in this thesis is Symbioflor 1. This is a probiotic strain isolated from a healthy human adult around 1950 [25]. It has been used as part of a commercially available probiotic product for over 50 years without any reports of it causing infections or other unwanted effects. Symbioflor 1 is susceptible to all clinically relevant antibiotics employed to treat enterococcal infections including ampicillin [25]. About 75% of the Symbioflor 1 genome is identical to the V583 genome [25]. Furthermore, the strain is devoid of any PAI-like structure and is missing the virulence factors cytolysin, enterococcal surface protein (Esp) and gelatinase (coccolysin) [25]. However, the genome sequence of Symbioflor 1 is not publicly available, thus it is not known which traits this strain possess other than what is found in V583. A genome atlas representation [48] of comparative genome hybridization analysis of the latter three strains illustrates the genomic similarities and differences compared to V583 (Figure 2).

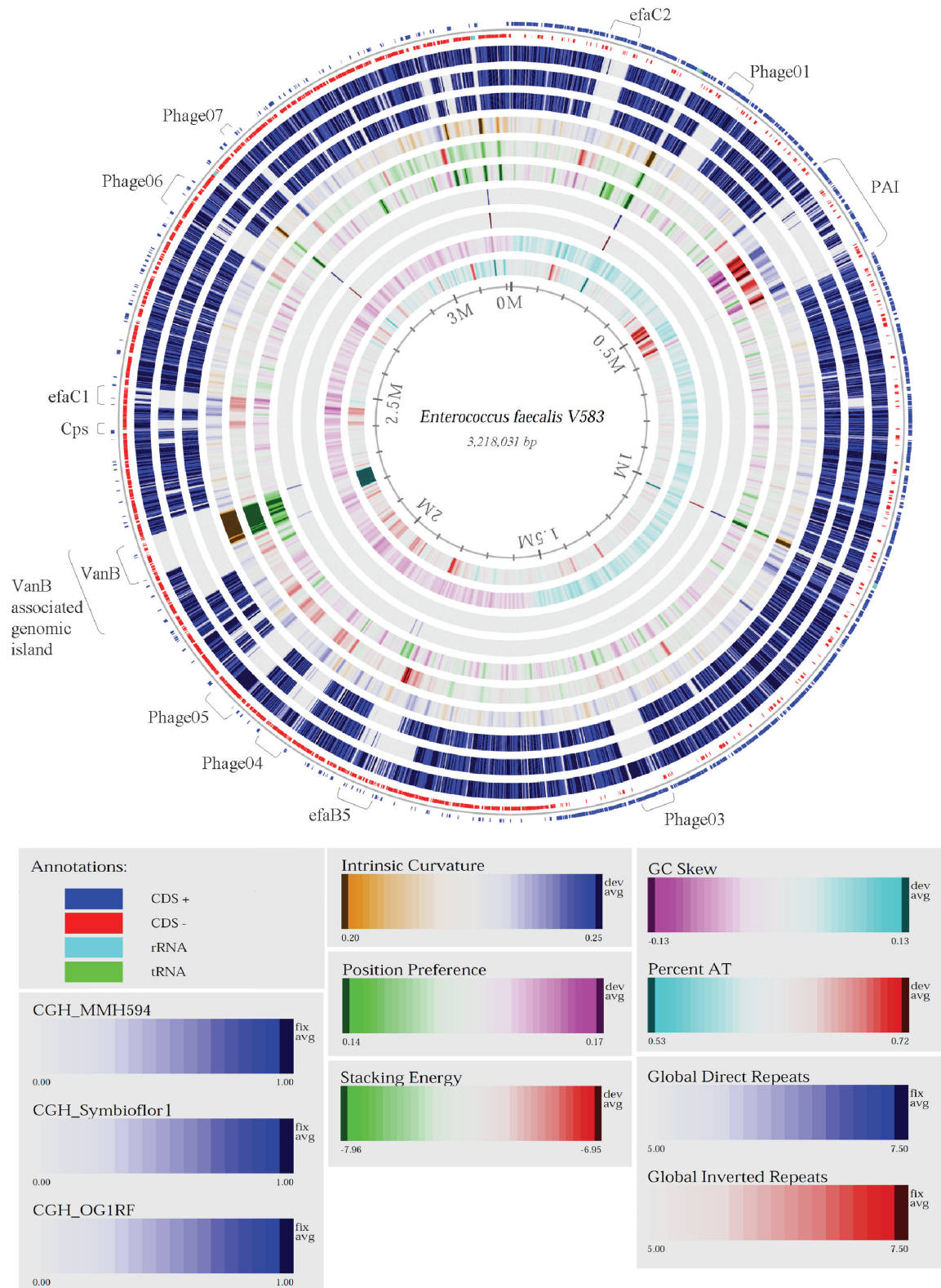


Figure 2: Genome atlas showing comparative genome hybridization of MMH594, Symbioflor 1 and OG1RF compared to the V583 genome. Putative mobile genetic elements predicted in the V583 genome are indicated around the genome atlas.

1.2 Antibiotics and antibiotic resistance in enterococci

Antibiotics were first described in 1929 by Alexander Fleming [35], in the form of penicillin. Since the mid 1940's antibiotics have been extensively used to treat infections, however in the last decades the usage of antibiotics has exploded. Besides the treatment of human infections, antimicrobial agents are used for laboratory purposes, in plant production, for industrial purposes, and in animals, both for treatment of disease and to improve growth. Approximately 35 tons of antibiotics were used to treat human infections in Denmark in 1997 [1]. In comparison, 53 tons were used for animal therapy and 107 tons were used as growth promoters [1]. This extensive usage of antibiotics has led to a massive distribution of antimicrobial agents in the environment, which consequently have promoted a natural selection for bacteria that possess antibiotic resistance genes.

Enterococci are intrinsically resistant to a broad range of antimicrobial agents, such as low concentrations of clindamycin and aminoglycosides, cephalosporins and penicillins [97] and are also able to acquire antibiotic resistance through the exchange of plasmids or transposons [59]. For more than 30 years the glycopeptides vancomycin and teicoplanin were used to treat serious infections caused by resistant Gram-positive organisms, without observing any development of resistance towards these antibiotics. However, in 1986 the first vancomycin resistant *E. faecium* isolates were reported from patients in France and England [78, 166]. The resistance against this widely used antibiotic quickly spread to a large population of enterococci (mainly *E. faecium*) [14], and in 1998, 20% of the enterococcal isolates in the United States (US) were resistant to vancomycin [39]. The rapid spread of vancomycin resistance among enterococci in the US in the 1990s, leading to infections which were extremely difficult to treat raised a newfound interest in enterococci [98]. The number of vancomycin resistant enterococci in Europe was low compared to in the US, but has increased dramatically in the last ten years, apparently following the trend in the US with a ten year delay [178].

Once vancomycin resistant enterococci have colonized a patient, they often persist in the GI tract and may be spread to other patients [98]. Furthermore, *E. faecium* (and to a lesser extent *E. faecalis*) can act as a reservoir for the spread of vancomycin resistance to other more pathogenic species, and the transfer of vancomycin resistance from enterococci to methicillin-resistant *Staphylococcus aureus* (MRSA) has been observed [10, 155]. The increasing spread

of antibiotic resistance other than vancomycin (e.g. ampicillin resistance) primary among *E. faecium* in the US [43, 67], and in the recent years also in Europe [165] rise further concerns about therapeutic options to treat enterococcal infections. Hence, it is a race against time to find new targets for novel antibiotics that the bacteria have not developed resistance against, which can be used to treat infections caused by multi resistant enterococci.

1.2.1 Erythromycin resistance in enterococci

Erythromycin is a macrolide antibiotic that inhibits protein synthesis by binding to the 50S subunit of the bacterial 70S ribosomal RNA complex [151, 169]. This binding interferes with the translocation of the peptidyl tRNA from the A to the P site, blocking the binding of an incoming tRNA and its attached amino acid to the nascent polypeptide chain (Fig. 3), and hence inhibiting production of functional proteins [156].

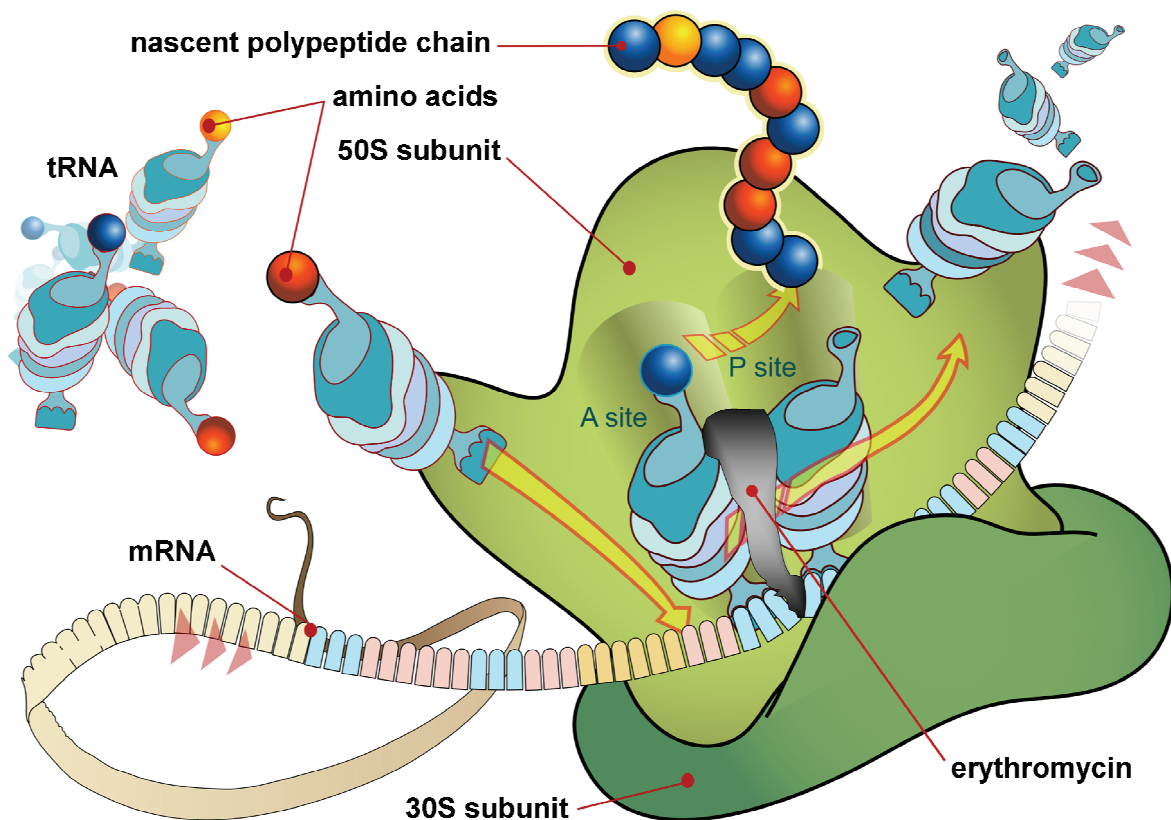


Figure 3. An illustration of prokaryote translation inhibited by binding of erythromycin to the large ribosomal subunit.

(Modified from http://en.wikipedia.org/wiki/File:Ribosome_mRNA_translation_en.svg)

Erythromycin (and other macrolides) are effective against beta-hemolytic streptococci, pneumococci, staphylococci and enterococci [77]. However, resistance to macrolides is very common in enterococci isolated from humans [24, 30, 121]. Three different mechanisms account for the acquired resistance to macrolide antibiotics (and also to lincosamide and streptogramin B antibiotics, hence referred to as MLS-resistance) in Gram-positive bacteria: (i) modification of the drug target, (ii) inactivation of the drug, (iii) and active efflux of the antibiotic [116]. Among enterococci, resistance to erythromycin is most commonly mediated by a series of structurally related *erythromycin-resistant methylase (erm)* genes. These genes encode enzymes that methylate an adenine residue in the 50S ribosomal subunit leading to a reduced binding of erythromycin and most other macrolides to the ribosomal subunit [150].

The expression of *erm* can either be inducible or constitutive; constitutive *erm* expression provides resistance to several macrolide, lincosamide and streptogramin B antibiotics, while inducible expression of *erm* only provides resistance to 14- and 15-membered-ring macrolides [77]. The inducible form of *erm* contains a leader sequence, which in the absence of erythromycin causes translational attenuation due to a stem-loop structure that makes the initiation codon of the *erm*-gene inaccessible for the ribosomes [77, 175]. In the presence of erythromycin, this antibiotic binds the ribosomes causing them to stall, which induces a conformational change of the mRNA that expose the initiation codon, and the translation of the methylase can proceed [77, 175]. The constitutive form does not contain a leader sequence, or has modifications in the leader sequence, such as point mutations, deletions or tandem repeats [92]. The most common *erm* variant in enterococci is *erm(B)* [64, 91, 116], which also is the variant found in V583 (EFA0007). The *erm(B)* gene in V583 does not contain the inducible leader sequence and hence is constitutively expressed. Other mechanisms of erythromycin resistance reported in enterococci include membrane bound efflux proteins encoded by the *mef(A/E)* and *msr(C)* genes [83, 116].

1.3 Enterococcal infections

Although infections due to enterococci were reported already in the mid-20th century [120], it is during the last decades that enterococci have received most attention due to the large increase of nosocomial infections [12, 97], which mainly have been attributed to the emergence of multiple antibiotic resistant strains [12, 98]. The enterococci now rank among the top three leading causes of hospital acquired infections where *E. faecalis* is responsible for

around 80% of these [59, 126]. However, in the last two decades the number of infection caused by *E. faecium* resistant to vancomycin and ampicillin has increased dramatically as described in section 1.2 [39, 165, 178], and with the continuing selective pressure of antibiotics it is possible that *E. faecium* and *E. faecalis* will account for equal numbers of enterococcal infections in the near future.

Before the emergence of vancomycin and aminoglycoside resistant *E. faecium* the number of infections caused by *E. faecium* was relatively low compared to infection caused by *E. faecalis*, and there are different theories around this lower frequency. Several studies have examined the ability of different infection derived isolates of *E. faecalis* and *E. faecium* to produce biofilm. These studies showed that while up to 100% of the *E. faecalis* isolates tested were biofilm producers, less than 50% of the *E. faecium* isolates produced biofilm [94]. A biofilm is formed when planktonic cells colonize a surface, aggregate and/or grow into multicellular colonies, surrounded by an exopolysaccharide matrix [26, 46]. More than 60% of all microbial infections in the human body involve bacteria within biofilms [80]. Bacteria within biofilms represent a therapeutic challenge since they are more protected compared to their planktonic counterparts, and hence more resistant against antibiotics and clearance by effectors of the host immune system [22]. It has been speculated that the larger number of *E. faecalis* involved in enterococcal infections compared to *E. faecium* is due to their differences in biofilm producing ability. However, as mentioned in section 1.1.1 *E. faecalis* is normally more abundant in the GI tract than *E. faecium* and hence the human body has a larger reservoir of *E. faecalis* which might explain the higher ratio of *E. faecalis* infections [34, 36]. There are also a larger number of virulence factors associated with *E. faecalis* than *E. faecium* [34], which will be described in further detail in section 1.4.

Antibiotic resistant *E. faecalis* has been isolated from surfaces in the hospital patient wards, thus the hospital can therefore serve as a reservoir of antibiotic resistant bacteria [12]. *E. faecalis* can be transmitted from patient to patient on the hands of healthcare workers, or on medical instrumentation such as thermometers [12]. Transmission of enterococci from the hands of healthcare workers can involve direct inoculation onto e.g. urinary catheters, but is more likely to result in colonization of the GI tract with the acquired strain, which becomes part of the patient's endogenous flora [85]. When *E. faecalis* has colonized the intestine of a patient, it can translocate through the epithelial cells of the intestine and can cause infection by further spreading to other locations within the body via the lymph nodes or blood stream

[69, 177]. Transmission of enterococcal strains can occur within a unit, between hospitals and even across borders, often due to a persistent colonization of the patients intestine, also after discharge of the patient from the hospital [85].

E. faecalis is associated with a variety of different infections, and some of its most common types of infections will be described in more detail below.

1.3.1 Urinary tract infection

In young healthy women, enterococci accounts for less than 5% of all urinary tract infections (UTI) [12]. However, enterococci are the second most common cause of hospital acquired UTIs world-wide [12] and UTI is the most common infection caused by enterococci [85]. Risk factors for enterococcal UTI are structural abnormalities, urinary tract instrumentation, catheterization, genitourinary tract pathology, as well as prior antibiotic treatment [44, 81, 96]. Enterococcal UTIs acquired in the hospital or long-term care units are likely to be caused by strains resistant to several different antibiotics [157, 158].

The ability to form biofilm has been proposed as an especially important trait for the bacterium's ability to cause UTI. A study of UTI caused by *E. faecalis* over a 12-year period showed that more than 60% of the isolates were medium to strong biofilm producers [133]. It has been shown that *E. faecalis* adhere to urinary tract epithelial cells, and that isolates from UTI adhere better to these cells than endocarditis isolates [45]. Several virulence factors such as Epa, Esp, Ace and Ebp (described in more detail in section 1.4.2) have been shown to be important for *E. faecalis* ability to cause infection in various UTI models [76, 135, 141].

1.3.2 Bacteremia

Enterococci constitute the third most prevalent pathogens isolated from bloodstream infections [24, 126]. Translocation across an intact epithelial barrier is thought to lead to many blood stream infections with no identifiable source [177], whereas identifiable sources include intravenous lines, abscesses and UTIs [65]. Even though an identifiable source of infection is known, the overall mortality rate from enterococcal bacteremia has been reported to be between 12-50% [171, 174]. Risk factors for developing enterococcal bacteremia are

high age, combined with multiple underlying diseases such as diabetes, cancer, heart disease and prior surgery [84, 111, 158]. Infections by cytolysin producing strains, such as MMH594 have been shown to worsen the outcome of the bacteremia [60].

1.3.3 Other enterococcal infections

Enterococci are the third leading cause of infective endocarditis, which is also one of the most therapeutic challenging infections caused by enterococci [89]. Even for antibiotic susceptible enterococci the treatment is difficult, and in situations with endocarditis caused by vancomycin or high-level aminoglycoside resistant strains, surgery is often necessary to remove the infected valve [85]. Enterococci are also frequently found in intra-abdominal, pelvic and soft tissue infections, although they rarely have been shown to cause monomicrobial infections at these sites [85]. The treatment for such infections, besides the use of antibiotics, is drainage of abscesses and debridement of infected tissue [85]. The importance of the intra-abdominal, pelvic and soft tissue infections has been debated, however these infections are sources to enterococcal bacteremia and surgical-site infections [85]. In fact, enterococci represent the most frequent cause of surgical-site infections in intensive care units [126]

1.4 Enterococcal virulence factors

Virulence factors can be defined as the gene products of a pathogen that enhance its ability to cause disease [11]. Virulence factors include secreted proteins, such as bacterial toxins and enzymes, and cell-surface structures, such as cell surface carbohydrates or proteins that mediate bacterial attachment or protect the pathogen from host defense mechanisms [11]. The importance of distinct virulence factors in enterococci has been under debate since they, besides being present in invasive isolates, also often are found in food- and community-derived isolates [23, 28, 37, 86]. However, patients infected with the MMH594 strain, which contains several of the known virulence genes, had an increased risk of death compared to patients infected with other nosocomial *E. faecalis* strains [60]. This suggests that several virulence genes together provide additional challenges for the host immune system. The *E. faecalis* virulence factors can be divided into two groups: secreted factors and cell envelope bound factors (Table 2).

Table 2. List of the major *E. faecalis* virulence factors (modified from [50])

Virulence factor	Description	Reference(s)
Secreted		
Cyl	Cytolysin, Hemolysin/bacteriocin; lyses a broad range of eukaryotic and gram-positive cells	[13, 19, 63, 66]
GelE	Gelatinase, zinc metalloprotease	[31, 118, 138]
SprE	Serine protease	[31, 118, 138]
Toxic metabolites	Reactive oxygen species; extracellular superoxide, hydrogen peroxide	[57]
Cell envelope bound		
Esp	Enterococcal surface protein; enhances biofilm formation and bladder epithelium colonization	[135]
Aggregation substance	Involved in conjugation and adhesion to eukaryotic cells	[13, 54, 71, 131]
Ace	Collagen-binding protein	[102, 125, 164]
Epa	Enterococcal polysaccharide antigen; antiphagocytic cell wall polysaccharide	[141, 152, 154]
Cps	Capsular polysaccharide, antiphagocytic cell wall polysaccharide	[49, 52, 55]
Ebp	Endocarditis and biofilm-associated pili	[104, 142]

1.4.1 Secreted virulence factors

Three important secreted proteins have been identified in *E. faecalis*: cytolysin, gelatinase and a serine protease. Cytolysin displays both hemolytic and bactericidal activity. Its importance for virulence in *E. faecalis* has been extensively studied in several animal models, showing that cytolysin producing strains are up to 10 times more toxic than non-cytolysin producers [13, 63, 66]. The cytolysin operon is either encoded on large pheromone-responsive plasmids [15, 62], or within the *E. faecalis* PAI [134]. The operon consists of six genes; *cylL_L* and *cylL_S* encoding the toxin, and *cylM*, *cylB*, *cylA*, and *cylI* playing a role in peptide modification, secretion, activation, and immunity, respectively [41, 42]. Transcription of the *cyl*-operon is repressed by CylR1 and CylR2, the products of a second operon responding by quorum-sensing to the level of CylL_S [61]. The accumulation of fully processed CylL_S releases the repression on the *cyl*-operon, and hence stimulates the production of more toxin as the cell density increases [61].

Regarding the secreted proteases they first of all provide peptide nutrients to the organism, but might also cause direct or indirect damage to host tissues making them potential virulence factors. The two proteases gelatinase (GelE, a zinc metalloprotease) and serine protease

(SprE) have been shown to be important for virulence in *E. faecalis* in various animal models [31, 118, 138]. It has also been shown that GelE confers immune system evasion by degradation of complement peptide C3, which makes *E. faecalis* less susceptible to phagocytosis [110]. Both proteases are regulated by the *fsr* quorum-sensing locus, which encodes a response regulator FsrA that is phosphorylated by the histidine kinase FsrC [119]. FsrC is autophosphorylated when sensing the accumulation of a peptide lactone molecule encoded by *fsrD* [99, 100]. The *fsr* system also regulates several other genes [5], and provides an additional effect on virulence in *E. faecalis* [31, 95, 138].

Potential virulence factors in *E. faecalis* can also be the production of toxic metabolites such as reactive oxygen species, which can damage cells or organs of the infected host [57]. Although the vast majority of *E. faecalis* strains produce superoxide, Huycke *et al.* showed that invasive *E. faecalis* isolates displayed an increased production of superoxide compared to non-invasive isolates [58], further emphasizing its role for virulence in *E. faecalis*. Due to its superoxide production, *E. faecalis* also has a large battery of genes encoding oxidative stress management [124]. This has been shown to be an advantage for its virulence potential [40, 73, 172, 173], since bacteria are exposed to massive oxidative stress by the hosts immune system during an infection [70, 160].

1.4.2 Cell envelope bound virulence factors

Factors that enhance the bacterium's ability to produce biofilm, adhere to host cells and extracellular matrix proteins such as collagen fibronectin etc. are considered important for the bacterium's ability to cause infection [11]. There are several factors that have been shown to contribute to biofilm formation, *e.g.* the involvement of Fsr and GelE (mentioned in section 1.4.1) in biofilm production [51, 72, 95, 115]. Another important biofilm enhancing factor is the enterococcal surface protein (Esp). Esp is highly associated with *E. faecalis* ability to form biofilm on polystyrene surfaces [163]. Esp has also been shown to be involved in adhesion of *E. faecalis* during colonization of the urinary tract; strains producing Esp were recovered in higher amounts from the bladder and urine compared to the isogenic mutant [135]. The *esp* gene is found within the *E. faecalis* PAI, in close association with the *cyl*-operon (see section 1.4.1) [134]. A locus (*ebpABC*) encoding pilus-like structures has been shown to be important in the initial attachment step of biofilm production [104]. The *ebp*-locus has also been shown to be important for endocarditis and urinary tract infection [104, 142]. Furthermore, the *ebp*-

locus is found in most of the *E. faecalis* strains examined, indicating an essential role for this pili in *E. faecalis* [17, 104, 139].

Two other enterococcal adhesins have also been found to be important for virulence in *E. faecalis*: an aggregation substance (pheromone-responsive plasmid encoded) and an adhesin to collagen in *E. faecalis* (ace). The main role of the aggregation substance is to aid the formation of mating aggregates during bacterial conjugation [16], however this factor also mediates binding to eukaryotic cells [71, 149, 168]. Ace has been shown to bind to collagen and laminin [47, 102, 103], although this binding can only be seen in gelatinase negative strains (or at 46°C), since the heat labile gelatinase can cleave these proteins [164]. Ace was recently shown to be involved in urinary tract infection in a mouse peritonitis model [76] and to be important for experimental endocarditis [143].

Other cell surface bound virulence factors that have received attention the recent years are two serotype-determining cell wall polysaccharides called the enterococcal polysaccharide biosynthesis cluster (*epa*) and the serotype 2 (also known as serotype C) capsular polysaccharide (*cps*). Both exopolysaccharides have been shown to be important for biofilm formation [95, 152]. Furthermore, the two cell wall polysaccharides have been found to be important for *E. faecalis* adherence/colonization and for evasion of the host immune system [49, 52, 55, 95, 141, 152, 154, 182]. The *epa* locus is widespread among *E. faecalis* and has been designated the *E. faecalis* group antigen [152], whereas *cps* is serotype determining and is more abundant among clinical isolates [49]. The cell wall polysaccharides are interesting as they have a potential of being targets in anti-enterococcal vaccines, hence providing an alternative to combat infections caused by antibiotic resistant *E. faecalis*. Furthermore, Cps encapsulated *E. faecalis* have been shown to mask the presence of enterococcal lipoteichoic acids (LTA) [161, 162], which are known to stimulate the immune response [132].

1.4.3 Other potential virulence factors

The composition of the *E. faecalis* PAI is characterized by its plasticity as revealed by inter strain differences, and especially the region surrounding the *cyl*-locus has been shown to be associated with a high frequency deletion [87, 88, 105, 134]. Within the PAI few genes other than the once encoding Esp and cytolysin production and the transcriptional regulator *araC* have been found to contribute to *E. faecalis* virulence [18, 63, 135]. Even so, the other genes

within the PAI might also contribute to the fitness of the strain, although no specific virulence function has so far been assigned, and this can be supported by the high frequency of the PAI in infection derived isolates [88, 134]. Factors that contribute to the robustness and survival of the bacterium during an infection can also be considered as virulence factors. These can be stress survival mechanisms, specialized metabolic features, manganese/iron uptake mechanisms etc. In fact, several other genes have been shown to contribute to the virulence of *E. faecalis*, e.g. the general stress protein Gls24, the superoxide dismutase (SodA) and a manganese scavenging system encoded by *efaCBA* [106, 140, 153, 172].

1.5 DNA microarray technology

In the past decade, a paradigm shift has emerged from the step-by-step studies of expression of individual genes to the analysis of thousands of genes in one experiment. This was made possible by the development of DNA microarrays [181]. Microarrays contain small probe sequences (typically 25- to 70-mer oligonucleotide probes) that are immobilized to specific positions on a glass slide [29]. Target sequences can be labeled in different ways [148], but labeling with two different fluorophores such as Cy3 and Cy5 is widely used. The use of two different fluorophores allows a co-hybridization of two different samples (such as treated-untreated) on one microarray slide. The number of targets bound to their respective probes is quantified by scanning the microarrays and measuring the fluorescence emitted by the hybridized fluorescence-labeled targets when excited by the light from the lasers [148]. Microarrays have many applications, but gene expression profiling, where the transcription of thousands of genes in one state is compared to their transcription in a different state (e.g. treated versus untreated, different growth phases, wild type versus mutant) is the most common application in microbiology, followed by comparative genome hybridizations (CGH) [29].

1.5.1 Experimental design and data analysis

Although microarrays provide a powerful tool, the design of a microarray transcription experiment is of utmost importance in order to gain reproducible and meaningful data. Important steps include knowledge about growth conditions and growth rate, accurate sample preparations; especially time from cell harvest to transcriptional stop/mRNA release due to rapid mRNA decay, and applying an appropriate statistical analysis [20, 122]. There are many critical steps from cell harvest to the data analysis, all which can bias the final results.

Therefore certain control points such as RNA quality control, measurement of amount of cDNA produced, and labeling efficiency are important steps in quality insurance of the final results [20]. A well designed and performed experiment will also reduce the unwanted experimental variance; the variance which is not due to the biological differences in the study, and hence, which only introduce noise to the final data [20, 108].

Due to all the steps that can affect the results, it is crucial to repeat the experiment. In general the more replicates, the more reliable data, although the usual limitation is the cost of each replicate [68, 179]. The number of biological replicates needed depends on the type of experiment performed, where bacterial cultures give a more uniform response from experiment to experiment compared to *e.g.* cancer cell from different individuals [108]. For cell culture studies, a minimum of three biological replicates are normally needed in order to get reliable estimates of the amount of differential gene expression [79]. Before a statistical analysis is carried out, normalization needs to be performed to adjust the microarray data for effects which arise from variation in the technology, rather than from biological differences between the samples or between the printed probes [145].

Microarray results are challenging with regards to statistical analysis due to the complex datasets consisting of several regulated genes compared to relatively few experiments [147]. There are different statistical methods available that are developed to analyze these specific types of data. In this thesis we have analyzed the data using two types of mixed models which accommodate for the two types of effects that arise from microarray experiments, fixed and random effects. Effects modeled as fixed effects are typically those we are interested in finding (*e.g.* the effect of the treatment) or effects that are expected to have a small number of outcomes and that we are interested to learn more about (dye-effect). Random effects are different types of noise (*e.g.* the effect of the array used, variation between replicated spots) [144, 179]. These mixed models, or similar methods are used to test for whether genes are expressed differentially at a certain significance level by providing a p-value. As a measure for reduction of false positive differentially expressed genes, p-values are often adjusted [147]. In this thesis two different methods for corrections of the p-values have been employed; (i) the very strict Bonferroni correction and (ii) the correction to control the false

discovery rate (FDR). The latter method is a less stringent correction and is commonly accepted as a standard in the microarray data analysis community [113].

One of the biggest challenges with microarray transcriptional analysis is to make sense out of the large amount of data produced, and to extract hypotheses and conclusions that are biologically meaningful [82]. Helpful tools for this purpose are online databases (such as KEGG^a and BioCyc^b) which can provide clues about regulatory mechanisms and biochemical pathways the regulated genes are involved in.

1.5.2 DNA microarray technology in the future

The use of microarrays, especially for CGH has in some extent been caught up by the more advanced and rapid genome sequencing, and some have been speculating whether this technique will take over several microarray applications [21, 180]. For example in the last two years the number of sequenced *E. faecalis* strains has increased from one strain to the currently 24^c strains with available complete or annotated draft genome sequences. Nonetheless, CGH has so far the advantage of being a fast and relatively inexpensive way to gain information about differences in the genome of *e.g.* different bacterial strains. The disadvantage with this method is that one can only gain knowledge about which genes are divergent in the strain of interest compared to the reference strain, and not any knowledge about the genes that are unique in the strain examined [27]. Then again, it is now possible to add more genomes to one microarray slide, since development in the production of microarrays allow a much higher density of probes on a slide [93], compared to only a few years back. The use of pan-genome arrays will allow a rapid screen of *e.g.* a strain collection, which can provide a better characterization of new strains, including emerging pathogens, and which also can provide insights into phylogenetic relationships between strains [180].

^a <http://www.genome.jp/kegg/>

^b <http://biocyc.org/>

^c [http://www.ncbi.nlm.nih.gov/sites/entrez?db=genomeprj&cmd=search&term=txid1351\[orgn\]](http://www.ncbi.nlm.nih.gov/sites/entrez?db=genomeprj&cmd=search&term=txid1351[orgn])

2. OBJECTIVES OF THIS STUDY

The main objectives of the work presented in this thesis were to investigate how *E. faecalis* grow and survive in different infection-relevant environments, and how these different environments affected its transcription. To accomplish this, DNA microarrays were used to study the global transcriptome.

This work included the following tasks:

- Investigate how treatment with the clinically relevant antibiotic erythromycin affect the global transcription of an erythromycin resistant nosocomial *E. faecalis* isolate, and identify new traits potentially implicated in intrinsic antibiotic resistance of *E. faecalis*.
- Investigate the transcriptional responses of an *E. faecalis* bacteremia isolate to blood as a biological cue and growth environment. Characterize the transcriptional responses of potential virulence traits, and examine *E. faecalis*' immediate responses as well as its adaptations to a prolonged existence in blood.
- Examine the gene expression of different *E. faecalis* isolates in response to growth in urine, and identify common genes important for the survival in urine as well as strain specific genes, possibly related to its pathogenic potential.

3. MAIN RESULTS AND DISCUSSIONS

E. faecalis is found in several different environments, but it is its ability to cause infections combined with its resistance against antibiotics, that give rise to the concern regarding this bacterium. Several virulence factors have been identified in *E. faecalis* (reviewed in [50]), and although mutants with deletions of the different virulence genes have been impaired in their ability to cause infections compared to the wild type strains, none of these virulence genes have been shown to be absolutely necessary for the ability of *E. faecalis* to cause infection (reviewed in [50]). The importance of the known virulence factors is therefore controversial, and it is likely that the interplay between several (virulence) genes is just as important for the pathogenic potential of a strain as each separate virulence factor. This thesis reveals clues about potential important global regulation in infection-relevant growth environments; when exposed to the commonly used antibiotic erythromycin (paper I), in blood and in the presence of trace amounts of blood (paper II), and finally in human urine (paper III). In the three papers different microarrays, different strains and different statistical analyses have been used; hence a comparison of the results in the three papers is not straightforward. However, some general comments about similarities and/or differences between the three papers will be discussed in the following sections.

Erythromycin is commonly used to treat Gram-positive bacterial infections when a patient is allergic to penicillin [77]. In paper I, gene regulation of *E. faecalis* V583 in response to sub-lethal concentrations of erythromycin was examined. Initial experiments showed that the growth of V583 was impeded by 50 µg/ml erythromycin. This indicated that although bearing an erythromycin resistance genes (*erm(B)*), V583 was affected by erythromycin treatment. We wanted to investigate the effect of erythromycin treatment on the global gene regulation in V583 and to examine the adaptation of the bacterium to this treatment over time. Furthermore, we wanted to uncover genes important for erythromycin treatment besides the *erm(B)* gene.

We discovered a drastic change in the gene expression of around 600 genes, which clearly indicated an effect of the erythromycin treatment on V583 although being erythromycin resistant. Many of the affected genes were related to changes in growth conditions/rate between treated and non-treated cells. Other genes were involved in modifications of the cell envelope, e.g. genes involved in fatty acid and phospholipid metabolism, which possibly can alter the uptake of erythromycin into the bacterial cells due to

changed cell wall/cell membrane properties. Erythromycin also affected genes related to the erythromycin target site, such as an up-regulation of several genes encoding ribosomal proteins and the down-regulation of several tRNA synthase genes. Previous studies have observed similar effects on genes encoding ribosomal proteins and tRNA synthase genes in response to translation inhibitors [33, 107]. This indicated that although the *erm(B)* gene was expressed, the bacterium was affected by the antibiotic treatment. Apparently, V583 compensated for the erythromycin-effect by an enhanced production of ribosomes, while the production of tRNA was decreased to accommodate the protein synthesis activity. Interestingly, six genes involved in (multi)drug resistance/transport, including a gene encoding an MsrC-like macrolide efflux pump were among the genes showing an enhanced expression at most or all time points tested. Hence, several genes other than *erm(B)* seem to be involved in erythromycin tolerance in *E. faecalis*, and one mechanism to overcome erythromycin stress can possibly be transport of erythromycin out of the bacterial cell by specialized transport systems/pumps.

Compared to the enhanced expression of several stress genes seen in *E. faecalis* during growth in blood or urine (paper II and III), erythromycin treatment did not trigger a general stress in the bacterium, as most of the genes related to stress management were either not regulated or down-regulated. Furthermore, the effect of erythromycin on the MsrC-like macrolide efflux pump is very interesting, as the gene encoding this pump was not expressed/not regulated in any strain/timepoint in paper II and III.

In paper II the gene regulation of V583 in response to growth in blood and also in the presence of 10% blood (YTB) was examined. It has previously been shown that *E. faecalis* modulates the expression of virulence genes during growth in serum, and that this regulation also is growth phase dependent [136]. Furthermore, *E. faecalis* strains isolated from urinary tract infections (UTIs) and endocarditis showed enhanced adherence to different human cell cultures after growth in serum [45], and 46 strains from different origins showed an enhanced binding to fibrinogen, fibronectin and collagen after growth in the presence of 40% serum [101]. These previous reports indicated that *E. faecalis* responds to growth in serum by changing its virulence and adherence properties, making it more suited to colonize organs of the infected host. We wanted to investigate what effect growth in blood and YTB had on the global gene expression in *E. faecalis*. Furthermore, we chose to use whole blood, instead of serum since this would better mimic the conditions encountered during bacteraemia.

Growth in blood resulted in a modulation of several genes. Surprisingly we only found an enhanced expression of a few of the virulence genes examined by Shepard and Gilmore [136]. However, several differences in the experimental setup, e.g. choice of *E. faecalis* strains, whole blood versus serum, and sampling at different time points and different growth stages could lead to such discrepancies. Another surprising observation was the down-regulation of both exopolysaccharide genes clusters (*epa* and *cps*) found in V583. Several studies have shown that the cell wall polysaccharides are important for biofilm formation, adherence and virulence of *E. faecalis* in several different studies [49, 52, 55, 95, 141, 152, 182]. However, although capsule formation is crucial for virulence in *Streptococcus pneumoniae* [176], the transcription of its capsular polysaccharide gene-cluster was not enhanced in blood during *in vivo* infection in mice [109]. Furthermore, the expression of a number of virulence traits in *S. pneumoniae* was shown to be body site dependent [109]. Hence, it is possible that the transcription of the *epa* and *cps* loci, and other virulence genes in *E. faecalis* would be enhanced in contact with host cells, and warrants further investigation.

The expression of several genes in V583 was affected immediately (5-15 minutes) after exposure to YTB, including genes related to the cell envelope and stress mechanisms. Some genes such as the fatty acid biosynthesis cluster (FASII) and *lrgAB* showed an enhanced expression throughout the experiment indicating an important role of these genes for the ability of V583 to grow in blood/YTB. The exact function of LrgAB has not been established, but two genes homologous to *lrgAB* were up-regulated in *Streptococcus agalactiae* in a similar study [90]. This previous observation combined with the continuously high level transcription shown in paper II suggest that the role of these genes in blood stream infection of *Enterococcus* and also *Streptococcus* should be investigated further. Interestingly, although growth in blood and urine triggered several common transcriptional changes, no enhanced expression of *lrgAB* was found in either of the strains examined in paper III, implying that this is a specific response to blood.

The work in paper III presents a slightly different perspective, where not only the transcriptional responses caused by growth in urine were examined, but more interestingly an investigation of different transcriptional responses of three *E. faecalis* strains from diverse origins were investigated. UTI is the most common infection caused by enterococci [85], and is also becoming exceedingly difficult to treat due to the antibiotic resistance among enterococci [157, 158]. Hence, to gain further knowledge about genes and processes

important for enterococcal UTI is essential in order to come up with alternative therapies. In paper III we used fresh human urine as a growth medium to partly mimic the conditions of the urinary tract. Growth of *E. faecalis* in urine has previously been shown to modulate several genes related to virulence in *E. faecalis* [9, 136]. Furthermore, we wanted to examine if there were any clear differences between the transcriptional responses in isolates from diverse origins. An initial growth experiment showed that the six strains examined grew equally good in urine, consistent with observations by Carlos and co-workers [8]. We therefore decided to further examine three strains from diverse origins, the pathogenic strains MMH594 and OG1RF, and the probiotic strain Symbioflor 1. Comparative genome hybridization (CGH) was performed to ensure that the oligos presented on the microarrays could reproduce the differences between these three strains and the V583 strain previously established [4, 25, 87, 88].

Similarly to what was found in the growth experiment, the transcriptional responses of the three different strains were also very comparable, hence there were no large differences in the transcriptional profiles of pathogenic and probiotic *E. faecalis* which could indicate crucial traits for causing UTI. Furthermore, the results obtained in paper III, revealed a number of similarities to the transcriptional responses of V583 in blood (paper II). In both paper II and III we found an enhanced expression of the FASII system. It is possible that the modulation of the fatty acid composition is important for virulence in *E. faecalis*. However, a previous study showed that the FASII genes also were modulated in response to the detergent SDS (sodium dodecyl sulphate), bovine bile, and a combination of SDS and bovine bile, suggesting that modulation of the fatty acid composition is a general response in *E. faecalis* to stressful conditions. The regulation of many other general stress genes, such as *gls24* and *gspA1-2* were also enhanced under both conditions, and notably several genes connected to oxidative stress were up-regulated both during growth in urine and blood, suggesting that oxidative stress management is important for growth in these environments. Furthermore, the experiments in both paper II and III suggests that iron- and manganese-uptake is important for growth in both blood and urine, while regulation of genes involved in metabolism was slightly different in urine compared to blood.

Counter-intuitively, similar to what was found in paper II we observed a down-regulation of the cell wall polysaccharide (*epa*) in all three strains. This exopolysaccharide was specifically shown to be important for infection in a UTI model [141]. Interestingly, we also found a reduced transcription of the *cps*-locus in MMH594, which was the only strain

containing these genes. These results may further indicate that the expression of the cell wall polysaccharides could be body-site dependent. The most intriguing results found during growth in urine was however an enhanced regulation of one of the most studied virulence determinants in *E. faecalis*, *fsrABC* in MMH594, but not in OG1RF (these genes are not present in Symbioflor 1). The seemingly higher expression of the *fsr*-regulon in urine by MMH594 compared to by OG1RF could imply a more virulent potential during a UTI of the former strain. However, OG1RF did cause an increased mortality in a mouse peritonitis model compared to V583, and outnumbered V583 in a mouse UTI model showing that OG1RF is equally virulent as/more virulent than V583 [4]. V583 and MMH594 are closely related, although MMH594 contains more virulence genes than V583 [134]. To our knowledge the ability of MMH594 to cause UTI compared to other *E. faecalis* strains has not previously been examined, and hence we can only speculate that the enhanced expression of *fsr* in MMH594, but not in OG1RF, will make it more potent to cause UTI. Most of the other virulence genes, including those involved in adherence were not significantly regulated in either strain during growth in urine.

The implication of a differential expression of genes such as *fsrABC*, the *epa*-cluster etc. is hard to predict since transcript levels detected by microarray do not reflect all regulatory processes in the cell, like post-transcriptional/post-translational processes that can alter the amount of active protein. Proteomics does however suffer from several constraints in terms of resolution and sensitivity, and to date no complete prokaryote or eukaryote proteome has been resolved [170]. Therefore, a combination of transcriptomic- and proteomic-approaches is needed to improve our knowledge on gene expression and cell performance. Even so, it is known that most conditions influence the transcription of several genes, not directly involved in the process studied. One example is the sigma factor B-dependent stress response of *Bacillus subtilis*, which not only is induced by various environmental stressors such as heat or carbon starvation [114, 117], but also by various antibiotics [3, 7]. An increased insight of the transcriptional networks existing in the organism of interest is of great importance in order to be able to extract the biological relevant responses arising from the conditions tested. Besides the work presented in this thesis there are currently only 4 publications devoted to *E. faecalis*' global transcriptional analysis [5, 6, 123, 146]. The results presented in this thesis therefore not only give an insight into the responses of *E. faecalis* to the infection related growth environments examined, but also contribute to the general understanding of *E. faecalis* global transcriptional regulation.

4. MAIN CONTRIBUTIONS AND FUTURE WORK

The work presented in this thesis has provided new insight into the adaptation *E. faecalis* undergoes to be able to grow and survive in infection-relevant growth environments. We have shown the importance of a rapid regulation of genes involved in metabolism, stress mechanisms and cell envelope modification in response to infection-relevant growth environments. Microarray data can point to new potentially interesting genes, not previously connected to virulence, as well as provide clues about global regulation of genes and operons important for virulence and survival. Besides providing much needed information about the global regulation of *E. faecalis* to various growth environments, the most interesting observations in this thesis were:

- The possible involvement of two ABC-transporters (EF1733-2) and a putative MsrC protein in the efflux of erythromycin in V583.
- A potential important role of LrgAB (EF3194-3) for growth of V583 in blood.
- Strain dependent expression of the *fsr*-system might contribute to the pathogenic potential during UTI.
- The involvement of cell envelope modifications, especially the modifications effectuated by the FASII pathway in stress management and survival in various infection-relevant growth environments.

In order to fully exploit the new information provided in this thesis, follow-up studies are needed. The erythromycin study has already been followed up by a study of the transcriptional responses of *E. faecalis* to another translation inhibiting antibiotic, chloramphenicol, and the responses have been compared with those of *E. faecalis* to erythromycin. Further experiments to enhance our understanding of *E. faecalis* resistance and tolerance to antibiotics have not been planned.

To extract more information from the already produced data in paper II and III, a study of the transcriptional responses of *E. faecalis* in a minimal medium performed in a similar manner would provide clues about which of the observed effects are relevant as responses to biological cues/infection-relevant growth environments, and which responses are due to a sudden transfer to a low nutrient growth environment. The extraction of these responses will probably reduce the number of genes important for the specific growth environments, and their relevance for growth and infection can be studied further by *e.g.* knock out studies.

Since we did not observe any large differences between isolates from different sources it would be interesting to examine the virulence potential of these and other strains in infection models, such as *Caenorhabditis elegans*. The importance of the different genes found in this thesis for virulence could also be tested in animal models.

The most intriguing experiments would however be *in vivo* infection studies of *E. faecalis*, either in animal models, but even more relevant in *E. faecalis* found in human infections such as UTIs. The use of pan-genome arrays based on all the sequenced *E. faecalis* strains would be most useful to be able to extract as much information as possible.

5. REFERENCES

1. **Aarestrup, F. M. and H. C. Wegener**, The effects of antibiotic usage in food animals on the development of antimicrobial resistance of importance for humans in *Campylobacter* and *Escherichia coli*. *Microbes and Infection*, 1999. **1**(8): p. 639-644.
2. **Andrewes, F. W. and T. J. Horder**, A STUDY OF THE STREPTOCOCCI PATHOGENIC FOR MAN. *The Lancet*, 1906. **168**(4333): p. 708-713.
3. **Bandow, J. E., H. Brotz, and M. Hecker**, *Bacillus subtilis* tolerance of moderate concentrations of rifampin involves the sigma(B)-dependent general and multiple stress response. *J Bacteriol*, 2002. **184**(2): p. 459-67.
4. **Bourgogne, A., D. A. Garsin, et al.**, Large scale variation in *Enterococcus faecalis* illustrated by the genome analysis of strain OG1RF. *Genome Biol*, 2008. **9**(7): p. R110.
5. **Bourgogne, A., S. G. Hilsenbeck, et al.**, Comparison of OG1RF and an isogenic *fsrB* deletion mutant by transcriptional analysis: the Fsr system of *Enterococcus faecalis* is more than the activator of gelatinase and serine protease. *J Bacteriol*, 2006. **188**(8): p. 2875-84.
6. **Bourgogne, A., L. C. Thomson, and B. E. Murray**, Bicarbonate enhances expression of the endocarditis and biofilm associated pilus locus, *ebpR-ebpABC*, in *Enterococcus faecalis*. *BMC Microbiol*, 2010. **10**(1): p. 17.
7. **Cao, M., T. Wang, et al.**, Antibiotics that inhibit cell wall biosynthesis induce expression of the *Bacillus subtilis* sigma(W) and sigma(M) regulons. *Mol Microbiol*, 2002. **45**(5): p. 1267-76.
8. **Carlos, A. R., J. Santos, et al.**, Enterococci from artisanal dairy products show high levels of adaptability. *Int J Food Microbiol*, 2009. **129**(2): p. 194-9.
9. **Carlos, A. R., T. Semedo-Lemsaddek, et al.**, Transcriptional analysis of virulence-related genes in enterococci from distinct origins. *J Appl Microbiol*, 2009.
10. **Chang, S., D. M. Sievert, et al.**, Infection with vancomycin-resistant *Staphylococcus aureus* containing the *vanA* resistance gene. *N Engl J Med*, 2003. **348**(14): p. 1342-7.
11. **Chen, L., J. Yang, et al.**, VFDB: a reference database for bacterial virulence factors. *Nucleic Acids Res*, 2005. **33**(Database issue): p. D325-8.
12. **Chenoweth, C. E.**, *Enterococcus* species, in *Hospital Epidemiology and Infection Control*, C. G. Mayhall, Editor. 2004, Lippincott Williams & Wilkins: Philadelphia. p. 529-544.
13. **Chow, J. W., L. A. Thal, et al.**, Plasmid-associated hemolysin and aggregation substance production contribute to virulence in experimental enterococcal endocarditis. *Antimicrob Agents Chemother*, 1993. **37**(11): p. 2474-7.
14. **Clark, N. C., R. C. Cooksey, et al.**, Characterization of glycopeptide-resistant enterococci from U.S. hospitals. *Antimicrob Agents Chemother*, 1993. **37**(11): p. 2311-7.
15. **Clewell, D. B.**, Plasmids, drug resistance, and gene transfer in the genus *Streptococcus*. *Microbiol Rev*, 1981. **45**(3): p. 409-36.
16. **Clewell, D. B.**, Bacterial sex pheromone-induced plasmid transfer. *Cell*, 1993. **73**(1): p. 9-12.
17. **Cobo Molinos, A., H. Abriouel, et al.**, Detection of *ebp* (endocarditis- and biofilm-associated pilus) genes in enterococcal isolates from clinical and non-clinical origin. *Int J Food Microbiol*, 2008. **126**(1-2): p. 123-6.

18. **Coburn, P. S., A. S. Baghdayan, et al.**, An AraC-type transcriptional regulator encoded on the *Enterococcus faecalis* pathogenicity island contributes to pathogenesis and intracellular macrophage survival. *Infect Immun*, 2008. **76**(12): p. 5668-76.
19. **Coburn, P. S. and M. S. Gilmore**, The *Enterococcus faecalis* cytolysin: a novel toxin active against eukaryotic and prokaryotic cells. *Cell Microbiol*, 2003. **5**(10): p. 661-9.
20. **Conway, T. and G. K. Schoolnik**, Microarray expression profiling: capturing a genome-wide portrait of the transcriptome. *Mol Microbiol*, 2003. **47**(4): p. 879-89.
21. **Coppee, J. Y.**, Do DNA microarrays have their future behind them? *Microbes Infect*, 2008. **10**(9): p. 1067-71.
22. **Costerton, J. W., P. S. Stewart, and E. P. Greenberg**, Bacterial biofilms: a common cause of persistent infections. *Science*, 1999. **284**(5418): p. 1318-22.
23. **Creti, R., M. Imperi, et al.**, Survey for virulence determinants among *Enterococcus faecalis* isolated from different sources. *J Med Microbiol*, 2004. **53**(Pt 1): p. 13-20.
24. **de Fatima Silva Lopes, M., T. Ribeiro, et al.**, Antimicrobial resistance profiles of dairy and clinical isolates and type strains of enterococci. *Int J Food Microbiol*, 2005. **103**(2): p. 191-8.
25. **Domann, E., T. Hain, et al.**, Comparative genomic analysis for the presence of potential enterococcal virulence factors in the probiotic *Enterococcus faecalis* strain Symbioflor 1. *Int J Med Microbiol*, 2007. **297**(7-8): p. 533-539.
26. **Donlan, R. M. and J. W. Costerton**, Biofilms: survival mechanisms of clinically relevant microorganisms. *Clin Microbiol Rev*, 2002. **15**(2): p. 167-93.
27. **Dorrell, N., S. J. Hinchliffe, and B. W. Wren**, Comparative phylogenomics of pathogenic bacteria by microarray analysis. *Curr Opin Microbiol*, 2005. **8**(5): p. 620-6.
28. **Eaton, T. J. and M. J. Gasson**, Molecular screening of *Enterococcus* virulence determinants and potential for genetic exchange between food and medical isolates. *Appl Environ Microbiol*, 2001. **67**(4): p. 1628-35.
29. **Ehrenreich, A.**, DNA microarray technology for the microbiologist: an overview. *Appl Microbiol Biotechnol*, 2006. **73**(2): p. 255-73.
30. **Emaneini, M., M. Aligholi, and M. Aminshahi**, Characterization of glycopeptides, aminoglycosides and macrolide resistance among *Enterococcus faecalis* and *Enterococcus faecium* isolates from hospitals in Tehran. *Pol J Microbiol*, 2008. **57**(2): p. 173-8.
31. **Engelbert, M., E. Mylonakis, et al.**, Contribution of gelatinase, serine protease, and *fsr* to the pathogenesis of *Enterococcus faecalis* endophthalmitis. *Infect Immun*, 2004. **72**(6): p. 3628-33.
32. **Euzéby, J. P.** List of Prokaryotic names with Standing in Nomenclature - Genus *Enterococcus*. 2010 [cited 2010 Jan]; Available from: <http://www.bacterio.cict.fr/e/enterococcus.html>.
33. **Evers, S., K. Di Padova, et al.**, Mechanism-related changes in the gene transcription and protein synthesis patterns of *Haemophilus influenzae* after treatment with transcriptional and translational inhibitors. *Proteomics*, 2001. **1**(4): p. 522-44.
34. **Fisher, K. and C. Phillips**, The ecology, epidemiology and virulence of *Enterococcus*. *Microbiology*, 2009. **155**(Pt 6): p. 1749-57.
35. **Fleming, A.**, On the antibacterial action of cultures of a penicillium, with special reference to their use in the isolation of *B. influenzae*. *The British Journal of Experimental Pathology*, 1929. **10**: p. 226-236.
36. **Franz, C. M., W. H. Holzappel, and M. E. Stiles**, Enterococci at the crossroads of food safety? *Int J Food Microbiol*, 1999. **47**(1-2): p. 1-24.

37. **Franz, C. M., A. B. Muscholl-Silberhorn, et al.**, Incidence of virulence factors and antibiotic resistance among Enterococci isolated from food. *Appl Environ Microbiol*, 2001. **67**(9): p. 4385-9.
38. **Freney, J., S. Bland, et al.**, Description and evaluation of the semiautomated 4-hour rapid ID 32 Strep method for identification of streptococci and members of related genera. *J Clin Microbiol*, 1992. **30**(10): p. 2657-61.
39. **Fridkin, S. K. and R. P. Gaynes**, Antimicrobial resistance in intensive care units. *Clin Chest Med*, 1999. **20**(2): p. 303-16, viii.
40. **Giard, J. C., E. Riboulet, et al.**, Characterization of Ers, a PrfA-like regulator of *Enterococcus faecalis*. *FEMS Immunol Med Microbiol*, 2006. **46**(3): p. 410-8.
41. **Gilmore, M. S., R. A. Segarra, and M. C. Booth**, An HlyB-type function is required for expression of the *Enterococcus faecalis* hemolysin/bacteriocin. *Infect Immun*, 1990. **58**(12): p. 3914-23.
42. **Gilmore, M. S., R. A. Segarra, et al.**, Genetic structure of the *Enterococcus faecalis* plasmid pAD1-encoded cytolytic toxin system and its relationship to lantibiotic determinants. *J Bacteriol*, 1994. **176**(23): p. 7335-44.
43. **Grayson, M. L., G. M. Eliopoulos, et al.**, Increasing resistance to beta-lactam antibiotics among clinical isolates of *Enterococcus faecium*: a 22-year review at one institution. *Antimicrob Agents Chemother*, 1991. **35**(11): p. 2180-4.
44. **Gross, P. A., L. M. Harkavy, et al.**, The epidemiology of nosocomial enterococcal urinary tract infection. *Am J Med Sci*, 1976. **272**(1): p. 75-81.
45. **Guzman, C. A., C. Pruzzo, et al.**, Role of adherence in pathogenesis of *Enterococcus faecalis* urinary tract infection and endocarditis. *Infect Immun*, 1989. **57**(6): p. 1834-8.
46. **Hall-Stoodley, L., J. W. Costerton, and P. Stoodley**, Bacterial biofilms: from the natural environment to infectious diseases. *Nat Rev Microbiol*, 2004. **2**(2): p. 95-108.
47. **Hall, A. E., E. L. Gorovits, et al.**, Monoclonal antibodies recognizing the *Enterococcus faecalis* collagen-binding MSCRAMM Ace: conditional expression and binding analysis. *Microb Pathog*, 2007. **43**(2-3): p. 55-66.
48. **Hallin, P. F., T. T. Binnewies, and D. W. Ussery**, The genome BLASTatlas-a GeneWiz extension for visualization of whole-genome homology. *Mol Biosyst*, 2008. **4**(5): p. 363-71.
49. **Hancock, L. E. and M. S. Gilmore**, The capsular polysaccharide of *Enterococcus faecalis* and its relationship to other polysaccharides in the cell wall. *Proc Natl Acad Sci U S A*, 2002. **99**(3): p. 1574-9.
50. **Hancock, L. E. and M. S. Gilmore**, Pathogenicity of enterococci, in *Gram-positive pathogens*, V. A. Fischetti, et al., Editors. 2006, ASM Press: Washington DC. p. p. 299-311.
51. **Hancock, L. E. and M. Perego**, The *Enterococcus faecalis* *fsr* two-component system controls biofilm development through production of gelatinase. *J Bacteriol*, 2004. **186**(17): p. 5629-39.
52. **Hancock, L. E., B. D. Shepard, and M. S. Gilmore**, Molecular analysis of the *Enterococcus faecalis* serotype 2 polysaccharide determinant. *J Bacteriol*, 2003. **185**(15): p. 4393-401.
53. **Hardie, J. M. and R. A. Whiley**, Classification and overview of the genera *Streptococcus* and *Enterococcus*. *Soc Appl Bacteriol Symp Ser*, 1997. **26**: p. 1S-11S.
54. **Hirt, H., P. M. Schlievert, and G. M. Dunny**, *In vivo* induction of virulence and antibiotic resistance transfer in *Enterococcus faecalis* mediated by the sex pheromone-sensing system of pCF10. *Infect Immun*, 2002. **70**(2): p. 716-23.

55. **Huebner, J., Y. Wang, et al.**, Isolation and chemical characterization of a capsular polysaccharide antigen shared by clinical isolates of *Enterococcus faecalis* and vancomycin-resistant *Enterococcus faecium*. *Infect Immun*, 1999. **67**(3): p. 1213-9.
56. **Huycke, M. M.**, Physiology of Enterococci in *The Enterococci : Pathogenesis, Molecular biology, and Antibiotic Resistance*, M. S. Gilmore, et al., Editors. 2002, ASM Press: Washington, D.C. p. 133-175.
57. **Huycke, M. M., V. Abrams, and D. R. Moore**, *Enterococcus faecalis* produces extracellular superoxide and hydrogen peroxide that damages colonic epithelial cell DNA. *Carcinogenesis*, 2002. **23**(3): p. 529-36.
58. **Huycke, M. M., W. Joyce, and M. F. Wack**, Augmented production of extracellular superoxide by blood isolates of *Enterococcus faecalis*. *J Infect Dis*, 1996. **173**(3): p. 743-6.
59. **Huycke, M. M., D. F. Sahn, and M. S. Gilmore**, Multiple-drug resistant enterococci: the nature of the problem and an agenda for the future. *Emerg Infect Dis*, 1998. **4**(2): p. 239-49.
60. **Huycke, M. M., C. A. Spiegel, and M. S. Gilmore**, Bacteremia caused by hemolytic, high-level gentamicin-resistant *Enterococcus faecalis*. *Antimicrob Agents Chemother*, 1991. **35**(8): p. 1626-34.
61. **Haas, W., B. D. Shepard, and M. S. Gilmore**, Two-component regulator of *Enterococcus faecalis* cytolysin responds to quorum-sensing autoinduction. *Nature*, 2002. **415**(6867): p. 84-7.
62. **Ike, Y., D. B. Clewell, et al.**, Genetic analysis of the pAD1 hemolysin/bacteriocin determinant in *Enterococcus faecalis*: Tn917 insertional mutagenesis and cloning. *J Bacteriol*, 1990. **172**(1): p. 155-63.
63. **Ike, Y., H. Hashimoto, and D. B. Clewell**, Hemolysin of *Streptococcus faecalis* subspecies zymogenes contributes to virulence in mice. *Infect Immun*, 1984. **45**(2): p. 528-30.
64. **Jensen, L. B., N. Frimodt-Moller, and F. M. Aarestrup**, Presence of erm gene classes in gram-positive bacteria of animal and human origin in Denmark. *FEMS Microbiol Lett*, 1999. **170**(1): p. 151-8.
65. **Jett, B. D., M. M. Huycke, and M. S. Gilmore**, Virulence of enterococci. *Clin Microbiol Rev*, 1994. **7**(4): p. 462-78.
66. **Jett, B. D., H. G. Jensen, et al.**, Contribution of the pAD1-encoded cytolysin to the severity of experimental *Enterococcus faecalis* endophthalmitis. *Infect Immun*, 1992. **60**(6): p. 2445-52.
67. **Jones, R. N., H. S. Sader, et al.**, Emerging multiply resistant enterococci among clinical isolates. I. Prevalence data from 97 medical center surveillance study in the United States. *Enterococcus Study Group. Diagn Microbiol Infect Dis*, 1995. **21**(2): p. 85-93.
68. **Jørstad, T. S., M. Langaas, and A. M. Bones**, Understanding sample size: what determines the required number of microarrays for an experiment? *Trends Plant Sci*, 2007. **12**(2): p. 46-50.
69. **Keely, S., L. E. Glover, et al.**, HIF-dependent Regulation of Platelet Activating Factor Receptor as a Route for Gram Positive Bacterial Translocation Across Epithelia. *Mol Biol Cell*, 2009.
70. **Klebanoff, S. J.**, Oxygen metabolism and the toxic properties of phagocytes. *Ann Intern Med*, 1980. **93**(3): p. 480-9.
71. **Kreft, B., R. Marre, et al.**, Aggregation substance of *Enterococcus faecalis* mediates adhesion to cultured renal tubular cells. *Infect Immun*, 1992. **60**(1): p. 25-30.

72. **Kristich, C. J., Y. H. Li, et al.**, Esp-independent biofilm formation by *Enterococcus faecalis*. J Bacteriol, 2004. **186**(1): p. 154-63.
73. **La Carbona, S., N. Sauvageot, et al.**, Comparative study of the physiological roles of three peroxidases (NADH peroxidase, Alkyl hydroperoxide reductase and Thiol peroxidase) in oxidative stress response, survival inside macrophages and virulence of *Enterococcus faecalis*. Mol Microbiol, 2007. **66**(5): p. 1148-63.
74. **Law-Brown, J. and P. R. Meyers**, *Enterococcus phoeniculicola* sp. nov., a novel member of the enterococci isolated from the uropygial gland of the Red-billed Woodhoopoe, *Phoeniculus purpureus*. Int J Syst Evol Microbiol, 2003. **53**(Pt 3): p. 683-5.
75. **Leblanc, D. J.**, Chapter 1.2.6 *Enterococcus*, in *THE PROKARYOTES. A Handbook on the Biology of Bacteria: Bacteria: Firmicutes, Cyanobacteria*, M. Dworkin, et al., Editors. 2006, Springer: New York. p. 175-204.
76. **Lebreton, F., E. Riboulet-Bisson, et al.**, *ace*, Which encodes an adhesin in *Enterococcus faecalis*, is regulated by Ers and is involved in virulence. Infect Immun, 2009. **77**(7): p. 2832-9.
77. **Leclercq, R. and P. Courvalin**, Bacterial resistance to macrolide, lincosamide, and streptogramin antibiotics by target modification. Antimicrob Agents Chemother, 1991. **35**(7): p. 1267-72.
78. **Leclercq, R., E. Derlot, et al.**, Plasmid-mediated resistance to vancomycin and teicoplanin in *Enterococcus faecium*. N Engl J Med, 1988. **319**(3): p. 157-61.
79. **Lee, M. L., F. C. Kuo, et al.**, Importance of replication in microarray gene expression studies: statistical methods and evidence from repetitive cDNA hybridizations. Proc Natl Acad Sci U S A, 2000. **97**(18): p. 9834-9.
80. **Lewis, K.**, Riddle of biofilm resistance. Antimicrob Agents Chemother, 2001. **45**(4): p. 999-1007.
81. **Lloyd, S., M. Zervos, et al.**, Risk factors for enterococcal urinary tract infection and colonization in a rehabilitation facility. Am J Infect Control, 1998. **26**(1): p. 35-9.
82. **Lockhart, D. J. and E. A. Winzeler**, Genomics, gene expression and DNA arrays. Nature, 2000. **405**(6788): p. 827-36.
83. **Luna, V. A., P. Coates, et al.**, A variety of gram-positive bacteria carry mobile *mef* genes. J Antimicrob Chemother, 1999. **44**(1): p. 19-25.
84. **Maki, D. G. and W. A. Agger**, Enterococcal bacteremia: clinical features, the risk of endocarditis, and management. Medicine (Baltimore), 1988. **67**(4): p. 248-69.
85. **Malani, P. N., C. A. Kauffman, and M. J. Zervos**, Enterococcal Disease, Epidemiology, and Treatment in *The Enterococci: Pathogenesis, Molecular biology, and Antibiotic Resistance*, M. S. Gilmore, et al., Editors. 2002, ASM Press: Washington, D.C. p. p. 385-408.
86. **Martin, B., M. Garriga, et al.**, Genetic diversity and safety aspects of enterococci from slightly fermented sausages. J Appl Microbiol, 2005. **98**(5): p. 1177-90.
87. **McBride, S. M., P. S. Coburn, et al.**, Genetic variation and evolution of the pathogenicity island of *Enterococcus faecalis*. J Bacteriol, 2009. **191**(10): p. 3392-402.
88. **McBride, S. M., V. A. Fischetti, et al.**, Genetic diversity among *Enterococcus faecalis*. PLoS One, 2007. **2**(7): p. e582.
89. **Megran, D. W.**, Enterococcal endocarditis. Clin Infect Dis, 1992. **15**(1): p. 63-71.
90. **Mereghetti, L., I. Sitkiewicz, et al.**, Extensive adaptive changes occur in the transcriptome of *Streptococcus agalactiae* (group B *Streptococcus*) in response to incubation with human blood. PLoS ONE, 2008. **3**(9): p. e3143.

91. **Min, Y. H., J. H. Jeong, et al.**, Heterogeneity of macrolide-lincosamide-streptogramin B resistance phenotypes in enterococci. *Antimicrob Agents Chemother*, 2003. **47**(11): p. 3415-20.
92. **Min, Y. H., A. R. Kwon, et al.**, Molecular analysis of constitutive mutations in *ermB* and *ermA* selected in vitro from inducibly MLSB-resistant enterococci. *Arch Pharm Res*, 2008. **31**(3): p. 377-80.
93. **Mockler, T. C., S. Chan, et al.**, Applications of DNA tiling arrays for whole-genome analysis. *Genomics*, 2005. **85**(1): p. 1-15.
94. **Mohamed, J. A. and D. B. Huang**, Biofilm formation by enterococci. *J Med Microbiol*, 2007. **56**(Pt 12): p. 1581-8.
95. **Mohamed, J. A., W. Huang, et al.**, Influence of origin of isolates, especially endocarditis isolates, and various genes on biofilm formation by *Enterococcus faecalis*. *Infect Immun*, 2004. **72**(6): p. 3658-63.
96. **Morrison, A. J., Jr. and R. P. Wenzel**, Nosocomial urinary tract infections due to *enterococcus*. Ten years' experience at a university hospital. *Arch Intern Med*, 1986. **146**(8): p. 1549-51.
97. **Murray, B. E.**, The life and times of the *Enterococcus*. *Clin Microbiol Rev*, 1990. **3**(1): p. 46-65.
98. **Murray, B. E.**, Vancomycin-resistant enterococcal infections. *N Engl J Med*, 2000. **342**(10): p. 710-21.
99. **Nakayama, J., Y. Cao, et al.**, Gelatinase biosynthesis-activating pheromone: a peptide lactone that mediates a quorum sensing in *Enterococcus faecalis*. *Mol Microbiol*, 2001. **41**(1): p. 145-54.
100. **Nakayama, J., S. Chen, et al.**, Revised model for *Enterococcus faecalis* *fsr* quorum-sensing system: the small open reading frame *fsrD* encodes the gelatinase biosynthesis-activating pheromone propeptide corresponding to staphylococcal AgrD. *J Bacteriol*, 2006. **188**(23): p. 8321-6.
101. **Nallapareddy, S. R. and B. E. Murray**, Role played by serum, a biological cue, in the adherence of *Enterococcus faecalis* to extracellular matrix proteins, collagen, fibrinogen, and fibronectin. *J Infect Dis*, 2008. **197**(12): p. 1728-36.
102. **Nallapareddy, S. R., X. Qin, et al.**, *Enterococcus faecalis* adhesin, *ace*, mediates attachment to extracellular matrix proteins collagen type IV and laminin as well as collagen type I. *Infect Immun*, 2000. **68**(9): p. 5218-24.
103. **Nallapareddy, S. R., K. V. Singh, et al.**, Diversity of *ace*, a gene encoding a microbial surface component recognizing adhesive matrix molecules, from different strains of *Enterococcus faecalis* and evidence for production of *ace* during human infections. *Infect Immun*, 2000. **68**(9): p. 5210-7.
104. **Nallapareddy, S. R., K. V. Singh, et al.**, Endocarditis and biofilm-associated pili of *Enterococcus faecalis*. *J Clin Invest*, 2006. **116**(10): p. 2799-807.
105. **Nallapareddy, S. R., H. Wenxiang, et al.**, Molecular characterization of a widespread, pathogenic, and antibiotic resistance-receptive *Enterococcus faecalis* lineage and dissemination of its putative pathogenicity island. *J Bacteriol*, 2005. **187**(16): p. 5709-18.
106. **Nannini, E. C., F. Teng, et al.**, Decreased Virulence of a *gls24* Mutant of *Enterococcus faecalis* OG1RF in an Experimental Endocarditis Model. *Infect. Immun.*, 2005. **73**(11): p. 7772-7774.
107. **Ng, W. L., K. M. Kazmierczak, et al.**, Transcriptional regulation and signature patterns revealed by microarray analyses of *Streptococcus pneumoniae* R6 challenged with sublethal concentrations of translation inhibitors. *J Bacteriol*, 2003. **185**(1): p. 359-70.

108. **Olson, N. E.**, The microarray data analysis process: from raw data to biological significance. *NeuroRx*, 2006. **3**(3): p. 373-83.
109. **Orihuela, C. J., J. N. Radin, et al.**, Microarray analysis of pneumococcal gene expression during invasive disease. *Infect Immun*, 2004. **72**(10): p. 5582-96.
110. **Park, S. Y., Y. P. Shin, et al.**, Immune evasion of *Enterococcus faecalis* by an extracellular gelatinase that cleaves C3 and iC3b. *J Immunol*, 2008. **181**(9): p. 6328-36.
111. **Patterson, J. E., A. H. Sweeney, et al.**, An analysis of 110 serious enterococcal infections. Epidemiology, antibiotic susceptibility, and outcome. *Medicine (Baltimore)*, 1995. **74**(4): p. 191-200.
112. **Paulsen, I. T., L. Banerjee, et al.**, Role of mobile DNA in the evolution of vancomycin-resistant *Enterococcus faecalis*. *Science*, 2003. **299**(5615): p. 2071-4.
113. **Pawitan, Y., S. Michiels, et al.**, False discovery rate, sensitivity and sample size for microarray studies. *Bioinformatics*, 2005. **21**(13): p. 3017-24.
114. **Petersohn, A., M. Brigulla, et al.**, Global analysis of the general stress response of *Bacillus subtilis*. *J Bacteriol*, 2001. **183**(19): p. 5617-31.
115. **Pillai, S. K., G. Sakoulas, et al.**, Effects of glucose on *fsr*-mediated biofilm formation in *Enterococcus faecalis*. *J Infect Dis*, 2004. **190**(5): p. 967-70.
116. **Portillo, A., F. Ruiz-Larrea, et al.**, Macrolide resistance genes in *Enterococcus* spp. *Antimicrob Agents Chemother*, 2000. **44**(4): p. 967-71.
117. **Price, C. W., P. Fawcett, et al.**, Genome-wide analysis of the general stress response in *Bacillus subtilis*. *Mol Microbiol*, 2001. **41**(4): p. 757-74.
118. **Qin, X., K. V. Singh, et al.**, Effects of *Enterococcus faecalis* *fsr* genes on production of gelatinase and a serine protease and virulence. *Infect Immun*, 2000. **68**(5): p. 2579-86.
119. **Qin, X., K. V. Singh, et al.**, Characterization of *fsr*, a regulator controlling expression of gelatinase and serine protease in *Enterococcus faecalis* OG1RF. *J Bacteriol*, 2001. **183**(11): p. 3372-82.
120. **Rantz, L. A. and W. M. M. Kirby**, Enterococcal infections. An evaluation of the importance of fecal streptococci and related organisms in the causation of human disease. *Arch. Intern. Med.*, 1943. **71**(4): p. 516-528.
121. **Reyes, J., M. Hidalgo, et al.**, Characterization of macrolide resistance in Gram-positive cocci from Colombian hospitals: a countrywide surveillance. *Int J Infect Dis*, 2007. **11**(4): p. 329-36.
122. **Rhodus, V. A. and R. A. LaRossa**, Uses and pitfalls of microarrays for studying transcriptional regulation. *Curr Opin Microbiol*, 2003. **6**(2): p. 114-9.
123. **Riboulet-Bisson, E., M. Sanguinetti, et al.**, Characterization of the *Ers* regulon of *Enterococcus faecalis*. *Infect Immun*, 2008. **76**(7): p. 3064-74.
124. **Riboulet, E., N. Verneuil, et al.**, Relationships between oxidative stress response and virulence in *Enterococcus faecalis*. *J Mol Microbiol Biotechnol*, 2007. **13**(1-3): p. 140-6.
125. **Rich, R. L., B. Kreikemeyer, et al.**, Ace is a collagen-binding MSCRAMM from *Enterococcus faecalis*. *J Biol Chem*, 1999. **274**(38): p. 26939-45.
126. **Richards, M. J., J. R. Edwards, et al.**, Nosocomial infections in combined medical-surgical intensive care units in the United States. *Infect Control Hosp Epidemiol*, 2000. **21**(8): p. 510-5.
127. **Ritchey, T. W. and H. W. Seeley**, Cytochromes in *Streptococcus faecalis* var. *zymogenes* grown in a haematin-containing medium. *J Gen Microbiol*, 1974. **85**(2): p. 220-8.

128. **Ruiz-Garbajosa, P., M. J. Bonten, et al.**, Multilocus sequence typing scheme for *Enterococcus faecalis* reveals hospital-adapted genetic complexes in a background of high rates of recombination. *J Clin Microbiol*, 2006. **44**(6): p. 2220-8.
129. **Sahm, D. F., J. Kissinger, et al.**, In vitro susceptibility studies of vancomycin-resistant *Enterococcus faecalis*. *Antimicrob Agents Chemother*, 1989. **33**(9): p. 1588-91.
130. **Schleifer, K. H. and R. Kilpper-Balz**, Transfer of *Streptococcus faecalis* and *Streptococcus faecium* to the Genus *Enterococcus* nom. rev. as *Enterococcus faecalis* comb. nov. and *Enterococcus faecium* comb. nov. *Int. J. Sys. Bacteriol*, 1984. **34**(1): p. 31-34.
131. **Schlievert, P. M., P. J. Gahr, et al.**, Aggregation and binding substances enhance pathogenicity in rabbit models of *Enterococcus faecalis* endocarditis. *Infect Immun*, 1998. **66**(1): p. 218-23.
132. **Schwandner, R., R. Dziarski, et al.**, Peptidoglycan- and lipoteichoic acid-induced cell activation is mediated by toll-like receptor 2. *J Biol Chem*, 1999. **274**(25): p. 17406-9.
133. **Seno, Y., R. Kariyama, et al.**, Clinical implications of biofilm formation by *Enterococcus faecalis* in the urinary tract. *Acta Med Okayama*, 2005. **59**(3): p. 79-87.
134. **Shankar, N., A. S. Baghdayan, and M. S. Gilmore**, Modulation of virulence within a pathogenicity island in vancomycin-resistant *Enterococcus faecalis*. *Nature*, 2002. **417**(6890): p. 746-50.
135. **Shankar, N., C. V. Lockatell, et al.**, Role of *Enterococcus faecalis* surface protein Esp in the pathogenesis of ascending urinary tract infection. *Infect Immun*, 2001. **69**(7): p. 4366-72.
136. **Shepard, B. D. and M. S. Gilmore**, Differential expression of virulence-related genes in *Enterococcus faecalis* in response to biological cues in serum and urine. *Infect Immun*, 2002. **70**(8): p. 4344-52.
137. **Sherman, J. M.**, The Streptococci. *Bacteriol Rev*, 1937. **1**(1): p. 3-97.
138. **Sifri, C. D., E. Mylonakis, et al.**, Virulence effect of *Enterococcus faecalis* protease genes and the quorum-sensing locus *fsr* in *Caenorhabditis elegans* and mice. *Infect Immun*, 2002. **70**(10): p. 5647-50.
139. **Sillanpaa, J., Y. Xu, et al.**, A family of putative MSCRAMMs from *Enterococcus faecalis*. *Microbiology*, 2004. **150**(Pt 7): p. 2069-78.
140. **Singh, K. V., T. M. Coque, et al.**, In vivo testing of an *Enterococcus faecalis* *efaA* mutant and use of *efaA* homologs for species identification. *FEMS Immunol Med Microbiol*, 1998. **21**(4): p. 323-31.
141. **Singh, K. V., R. J. Lewis, and B. E. Murray**, Importance of the *epa* Locus of *Enterococcus faecalis* OG1RF in a Mouse Model of Ascending Urinary Tract Infection. *J Infect Dis*, 2009. **200**(3): p. 417-20.
142. **Singh, K. V., S. R. Nallapareddy, and B. E. Murray**, Importance of the *ebp* (endocarditis- and biofilm-associated pilus) locus in the pathogenesis of *Enterococcus faecalis* ascending urinary tract infection. *J Infect Dis*, 2007. **195**(11): p. 1671-7.
143. **Singh, K. V., S. R. Nallapareddy, et al.**, Importance of the collagen adhesin *ace* in pathogenesis and protection against *Enterococcus faecalis* experimental endocarditis. *PLoS Pathog*, 2010. **6**(1): p. e1000716.
144. **Smyth, G. K.**, Linear models and empirical bayes methods for assessing differential expression in microarray experiments. *Stat Appl Genet Mol Biol*, 2004. **3**: p. Article3.
145. **Smyth, G. K. and T. Speed**, Normalization of cDNA microarray data. *Methods*, 2003. **31**(4): p. 265-73.

146. **Solheim, M., Å. Aakra, et al.**, Transcriptional responses of *Enterococcus faecalis* V583 to bovine bile and sodium dodecyl sulfate. *Appl Environ Microbiol*, 2007. **73**(18): p. 5767-74.
147. **Stekel, D.**, Analysis of Differentially Expressed Genes, in *Microarrays Bioinformatics*, D. Stekel, Editor. 2003, Cambridge University Press: New York.
148. **Storhoff, J. J., S. S. Marla, et al.**, Labels and Detection Methods in *Microarray technology and its applications*, U. R. Müller and D. V. Nicolau, Editors. 2005, Springer-Verlag Berlin. p. 147-178.
149. **Sussmuth, S. D., A. Muscholl-Silberhorn, et al.**, Aggregation substance promotes adherence, phagocytosis, and intracellular survival of *Enterococcus faecalis* within human macrophages and suppresses respiratory burst. *Infect Immun*, 2000. **68**(9): p. 4900-6.
150. **Tanaka, T. and B. Weisblum**, Systematic difference in the methylation of ribosomal ribonucleic acid from gram-positive and gram-negative bacteria. *J Bacteriol*, 1975. **123**(2): p. 771-4.
151. **Taubman, S. B., N. R. Jones, et al.**, Sensitivity and resistance to erythromycin in *Bacillus subtilis* 168: the ribosomal binding of erythromycin and chloramphenicol. *Biochim Biophys Acta*, 1966. **123**(2): p. 438-40.
152. **Teng, F., K. D. Jacques-Palaz, et al.**, Evidence that the enterococcal polysaccharide antigen gene (*epa*) cluster is widespread in *Enterococcus faecalis* and influences resistance to phagocytic killing of *E. faecalis*. *Infect Immun*, 2002. **70**(4): p. 2010-5.
153. **Teng, F., E. C. Nannini, and B. E. Murray**, Importance of *gls24* in virulence and stress response of *Enterococcus faecalis* and use of the Gls24 protein as a possible immunotherapy target. *J Infect Dis*, 2005. **191**(3): p. 472-80.
154. **Teng, F., K. V. Singh, et al.**, Further characterization of the *epa* gene cluster and *Epa* polysaccharides of *Enterococcus faecalis*. *Infect Immun*, 2009. **77**(9): p. 3759-67.
155. **Tenover, F. C., L. M. Weigel, et al.**, Vancomycin-resistant *Staphylococcus aureus* isolate from a patient in Pennsylvania. *Antimicrob Agents Chemother*, 2004. **48**(1): p. 275-80.
156. **Tenson, T., M. Lovmar, and M. Ehrenberg**, The mechanism of action of macrolides, lincosamides and streptogramin B reveals the nascent peptide exit path in the ribosome. *J Mol Biol*, 2003. **330**(5): p. 1005-14.
157. **Terpenning, M. S., S. F. Bradley, et al.**, Colonization and infection with antibiotic-resistant bacteria in a long-term care facility. *J Am Geriatr Soc*, 1994. **42**(10): p. 1062-9.
158. **Terpenning, M. S., M. J. Zervos, et al.**, Enterococcal infections: an increasing problem in hospitalized patients. *Infect Control Hosp Epidemiol*, 1988. **9**(10): p. 457-61.
159. **Thiercelin, M. E.**, Sur un diplocoque saprophyte de l'intestin susceptible de devenir pathogène. *C.R. Séances Société Biologie*, 1899(5): p. 269-271.
160. **Thomas, E. L., R. I. Lehrer, and R. F. Rest**, Human neutrophil antimicrobial activity. *Rev Infect Dis*, 1988. **10 Suppl 2**: p. S450-6.
161. **Thurlow, L. R., V. C. Thomas, et al.**, *Enterococcus faecalis* capsular polysaccharide serotypes C and D and their contributions to host innate immune evasion. *Infect Immun*, 2009. **77**(12): p. 5551-7.
162. **Thurlow, L. R., V. C. Thomas, and L. E. Hancock**, Capsular polysaccharide production in *Enterococcus faecalis* and contribution of CpsF to capsule serospecificity. *J Bacteriol*, 2009. **191**(20): p. 6203-10.

163. **Toledo-Arana, A., J. Valle, et al.**, The enterococcal surface protein, Esp, is involved in *Enterococcus faecalis* biofilm formation. *Appl Environ Microbiol*, 2001. **67**(10): p. 4538-45.
164. **Tomita, H. and Y. Ike**, Tissue-specific adherent *Enterococcus faecalis* strains that show highly efficient adhesion to human bladder carcinoma T24 cells also adhere to extracellular matrix proteins. *Infect Immun*, 2004. **72**(10): p. 5877-85.
165. **Top, J., R. Willems, et al.**, Ecological replacement of *Enterococcus faecalis* by multiresistant clonal complex 17 *Enterococcus faecium*. *Clin Microbiol Infect*, 2007. **13**(3): p. 316-9.
166. **Uttley, A. H., C. H. Collins, et al.**, Vancomycin-resistant enterococci. *Lancet*, 1988. **1**(8575-6): p. 57-8.
167. **Vandamme, P., E. Vercauteren, et al.**, Survey of enterococcal susceptibility patterns in Belgium. *J Clin Microbiol*, 1996. **34**(10): p. 2572-6.
168. **Vanek, N. N., S. I. Simon, et al.**, *Enterococcus faecalis* aggregation substance promotes opsonin-independent binding to human neutrophils via a complement receptor type 3-mediated mechanism. *FEMS Immunol Med Microbiol*, 1999. **26**(1): p. 49-60.
169. **Vazquez, D.**, The Macrolide Antibiotics. , in *Antibiotics III. Mechanism of action of antimicrobial and antitumor agents.*, J. W. Corcoran and F. E. Hahn, Editors. 1975, Springer-Verlag: New York. p. 459-479.
170. **Vercauteren, F. G., L. Arckens, and R. Quirion**, Applications and current challenges of proteomic approaches, focusing on two-dimensional electrophoresis. *Amino Acids*, 2007. **33**(3): p. 405-14.
171. **Vergis, E. N., N. Shankar, et al.**, Association between the presence of enterococcal virulence factors gelatinase, hemolysin, and enterococcal surface protein and mortality among patients with bacteremia due to *Enterococcus faecalis*. *Clin Infect Dis*, 2002. **35**(5): p. 570-5.
172. **Verneuil, N., A. Maze, et al.**, Implication of (Mn)superoxide dismutase of *Enterococcus faecalis* in oxidative stress responses and survival inside macrophages. *Microbiology*, 2006. **152**(Pt 9): p. 2579-89.
173. **Verneuil, N., A. Rince, et al.**, Contribution of a PerR-like regulator to the oxidative-stress response and virulence of *Enterococcus faecalis*. *Microbiology*, 2005. **151**(Pt 12): p. 3997-4004.
174. **Vigani, A. G., A. M. Oliveira, et al.**, Clinical, epidemiological, and microbiological characteristics of bacteremia caused by high-level gentamicin-resistant *Enterococcus faecalis*. *Braz J Med Biol Res*, 2008. **41**(10): p. 890-5.
175. **Weisblum, B.**, Insights into erythromycin action from studies of its activity as inducer of resistance. *Antimicrob Agents Chemother*, 1995. **39**(4): p. 797-805.
176. **Weiser, J. N., R. Austrian, et al.**, Phase variation in pneumococcal opacity: relationship between colonial morphology and nasopharyngeal colonization. *Infect Immun*, 1994. **62**(6): p. 2582-9.
177. **Wells, C. L., R. P. Jechorek, and S. L. Erlandsen**, Evidence for the translocation of *Enterococcus faecalis* across the mouse intestinal tract. *J Infect Dis*, 1990. **162**(1): p. 82-90.
178. **Werner, G., T. M. Coque, et al.**, Emergence and spread of vancomycin resistance among enterococci in Europe. *Euro Surveill*, 2008. **13**(47).
179. **Wernisch, L., S. L. Kendall, et al.**, Analysis of whole-genome microarray replicates using mixed models. *Bioinformatics*, 2003. **19**(1): p. 53-61.
180. **Willenbrock, H., P. F. Hallin, et al.**, Characterization of probiotic *Escherichia coli* isolates with a novel pan-genome microarray. *Genome Biol*, 2007. **8**(12): p. R267.

181. **Wodicka, L., H. Dong, et al.**, Genome-wide expression monitoring in *Saccharomyces cerevisiae*. Nat Biotechnol, 1997. **15**(13): p. 1359-67.
182. **Xu, Y., K. V. Singh, et al.**, Analysis of a gene cluster of *Enterococcus faecalis* involved in polysaccharide biosynthesis. Infect Immun, 2000. **68**(2): p. 815-23.

PAPER I

Transcriptional Response of *Enterococcus faecalis* V583 to Erythromycin

Ågot Aakra,^{1*} Heidi Vebø,¹ Lars Snipen,² Helmut Hirt,³ Are Aastveit,² Vivek Kapur,⁴ Gary Dunny,³ Barbara Murray,⁵ and Ingolf F. Nes^{1*}

Laboratory of Microbial Gene Technology¹ and Section for Bioinformatics and Data Analysis,² Department of Chemistry, Biotechnology and Food Science, Norwegian University of Life Sciences, Ås, Norway; Department of Microbiology, University of Minnesota Medical School, Minneapolis, Minnesota 55455³; Department of Veterinary Pathobiology, Biomedical Genomics Center, University of Minnesota, St. Paul, Minnesota 55108⁴; and Center for the Study of Emerging and Re-Emerging Pathogens, The University of Texas Medical School, Houston, Texas 77030⁵

Received 1 November 2004/Returned for modification 16 January 2005/Accepted 27 February 2005

A transcriptional profile of *Enterococcus faecalis* V583 (V583) treated with erythromycin is presented. This is the first study describing a complete transcriptional profile of *Enterococcus*. *E. faecalis* is a common and nonvirulent bacterium in many natural environments, but also an important cause of nosocomial infections. We have used a genome-wide microarray based on the genome sequence of V583 to study gene expression in cells exposed to erythromycin. V583 is resistant to relatively high concentrations of erythromycin, but growth is retarded by the treatment. The effect of erythromycin treatment on V583 was studied by a time course experiment; samples were extracted at five time points over a period of 90 min. A drastic change in gene transcription was seen with the erythromycin-treated cells compared to the untreated cells. Altogether, 260 genes were down-regulated at one or more time points, while 340 genes were up-regulated. Genes encoding hypothetical proteins and genes encoding transport and binding proteins were the two most dominating groups of differentially expressed genes. The gene encoding *ermB* (EFA0007) was expressed, but not differentially, which indicated that other genes are important for the survival and growth maintenance of V583 treated with erythromycin. One of these genes is a putative MsrC-like protein, which was up-regulated at all time points studied. Other specific genes that were found to be up-regulated were genes encoding ABC transporters and two-component regulatory systems, and these may be genes that are important for the specific response of V583 to erythromycin.

Enterococci are commonly found in the gastrointestinal tracts of humans and animals. Most enterococci are not virulent, and some are frequently found in artisan fermented foods, in which they contribute to longer shelf life, flavor, and texture. However, enterococci are best known as antibiotic-resistant opportunistic pathogens that are commonly recovered from patients who have received multiple courses of antibiotics and have been hospitalized for prolonged periods. Innate resistance to many antibiotics is a common trait among enterococci (see, e.g., reference 7 and the references therein). They also acquire antibiotic resistance determinants, including resistance to vancomycin, very rapidly from the environment (see, e.g., references 16–18). The two species best known to be involved in infections in human and animal bodies are *Enterococcus faecalis* and *E. faecium* (6).

The complete genome sequence of the vancomycin-resistant *E. faecalis* V583 (V583) is now available (20). Access to the genome sequence opens new possibilities to gain basic information on the molecular biology of the organism, and one of the tools that can be used to exploit the genome sequence experimentally is the DNA microarray technology. Microarrays give us the opportunity to study all transcriptional events

going on in a cell and identify which genes are involved in certain cell processes in one experiment. The main advantage of the microarray technology is the ability to study the transcription of thousands of genes in one experiment. By nature, the microarray technology is explorative and hypothesis generating. The results of microarray experiments are, in principle, snapshots of the transcriptional activities of the cell. Moreover, since transcription and translation are coupled processes in prokaryotes, the transcriptome should reflect the proteome well. Indeed, it has been shown that regulation of the majority of genes parallels the level of proteins produced (2). On a genome-wide scale, it is difficult to speculate about which bacterial genes are regulated or not during certain conditions, and it is also hard to gain such information through the use of traditional low-throughput methods. By the use of microarrays, one can relatively quickly obtain information about gene expression levels and thereby explore the responses of the cells to changing growth conditions.

V583 survives and grows in media containing relatively high levels of the commonly used macrolide antibiotic erythromycin, but the addition of erythromycin to the culture retards cell growth. In sensitive cells, erythromycin inhibits protein synthesis by binding to the large ribosomal subunit close to the peptidyl transfer center. Protein synthesis is thereby aborted during early rounds of translation, since access to the nascent peptide channel is prevented (29). In gram-negative bacteria,

* Corresponding author. Mailing address: Laboratory of Microbial Gene Technology, Norwegian University of Life Sciences, P.O. Box 5003, N-1432 Ås, Norway. Phone: 47 64965894. Fax: 47 64941465. E-mail for Ågot Aakra: agot.aakra@umb.no.

intrinsic resistance to erythromycin is due to the impermeability of the cellular outer membrane to this hydrophobic macrolide. Two main mechanisms of erythromycin resistance that have been identified in gram-positive bacteria are as follows: (i) Target modification by the *erm* (erythromycin resistant methylase) genes that encode enzymes which methylate rRNA has been described. The rRNA methylation causes conformational changes in the P site of the rRNA and prevention of macrolide binding. (*erm*-mediated resistance in enterococci has been described. The pAD1-like plasmid pTEF1 of V583 encodes ErmB, an rRNA adenine dimethylase family protein.) (ii) A macrolide efflux resistance mechanism, which is an energy-dependent pump, has been described for both gram-positive and gram-negative bacteria (22).

This paper describes the use of a genome-wide amplicon-based microarray based on the genome sequence of V583 to obtain a profile of the transcriptional events in V583 cells treated with erythromycin. The aim of the work was to obtain an overview of how erythromycin affects transcriptional events and bacterial growth, aside from described resistance mechanisms, and how this bacterium tolerates stress. Taking into account the observed phenotypic effects of erythromycin on V583, it was expected that genes involved in general stress responses, genes involved in protein synthesis, and genes encoding (multi)drug resistance would be among the genes affected by the antibiotic treatment.

MATERIALS AND METHODS

Bacterial strain and growth conditions. The bacterium studied was *E. faecalis* V583 (V583), which was the first vancomycin-resistant clinical isolate reported in the United States (24) and whose genome sequence has been completed (20). V583 samples were grown overnight in flasks with brain heart infusion (BHI) medium (Difco) at 37°C on a rotary shaker (300 rpm). Cultures were then diluted 50× and grown in BHI, as described above, for 1 h. Cultures were split in two, and erythromycin (Sigma) was added to one of the cultures. The final concentration of erythromycin was 50 µg/ml. The two cultures (BHI and BHI plus erythromycin) were then incubated further, and 3-ml samples of each culture were collected immediately after the addition of erythromycin (*t*0), and after 15 (*t*15), 30 (*t*30), 60 (*t*60), and 90 (*t*90) min. Growth of the cultures was monitored spectrophotometrically (optical density at 600 nm [OD₆₀₀]). Samples were centrifuged for 5 min at 4°C, and pellets were flash frozen in liquid N₂ prior to RNA extraction.

Construction of V583 microarrays. PCR primers for all V583 open reading frames (The Institute for Genomic Research, May 2002 update) were designed using Primer3 (freeware; http://frodo.wi.mit.edu/cgi-bin/primer3/primer3_www.cgi), which has commonly been used for primer design during microarray construction (3, 13). Primers were designed to amplify full-length or ~500-bp amplicons from genomic DNA. PCRs (50-µl reaction volume) were run with 50 mM KCl; 10 mM Tris-HCl (pH 9.0); 0.1% Triton X-100; 2.5 mM MgCl₂; 2 µM of each primer; 0.2 mM of each dATP, dCTP, dGTP, and dTTP; 10 to 20 ng genomic DNA; and 1.25 U *Taq* polymerase (Promega). Amplifications were performed with Perkin-Elmer 9600 thermocyclers with an initial denaturation at 94°C for 3 min and then 35 cycles of denaturation at 94°C for 1 min, annealing at 54 to 56°C for 1 min (the annealing temperature varied depending on the melting temperatures of the primers), and an extension at 74°C for 1 min. The PCR was finalized with incubation at 74°C for 10 min. The quality of the PCR products was assessed by agarose gel electrophoresis. The PCR products were cleaned up using a QIAquick 96 PCR purification kit (QIAGEN). Of a total of 3,337 predicted open reading frames in the V583 genome, 3,160 PCR products were obtained. The purified PCR products were eluted in 3× SSC (1× SSC is 0.15 M NaCl plus 0.015 M sodium citrate)-0.01% sodium dodecyl sulfate (SDS) and printed on Corning UltraGaps slides. All PCR products, representing the predicted V583 open reading frames, were printed in five copies on the slides. In addition, negative controls (three genes from *Arabidopsis thaliana*, buffer [3× SSC, 0.01% SDS], and empty spots) and digested genomic DNA from V583

(positive control) were spotted on the arrays. Altogether, 3,502 samples were spotted on the slides.

RNA isolation. Total RNA extractions from the samples collected at the time points described above were performed with RNeasy Mini columns (QIAGEN), with DNA digestions being done on the columns by the addition of 82 Kunitz units of RNase-free DNase (QIAGEN) and incubation at room temperature for 15 min. The integrity of and concentration of RNA samples were measured using a RNA 600 Nano LabChip kit and a Bioanalyzer 2100 (Agilent Technologies). For both strains, 10 µg of each RNA sample was used in separate hybridization experiments on identical arrays.

cDNA synthesis, fluorescent labeling, hybridization, and microarray data analysis. For reverse transcription and cDNA synthesis, 30 µg of random hexamers (Amersham) and 10 µg of total RNA was initially preheated at 70°C for 10 min and was incubated for 10 min at 4°C and then for 2 h at 42°C in an Eppendorf Mastercycler. The reverse transcription (RT)-PCR mix was 1× first-strand buffer, 10 mM dithiothreitol; 380 units of Superscript II RT; 500 µM concentrations of dATP, dCTP, and dGTP; 300 µM dTTP (all from Invitrogen), and 200 µM amino-allyl-labeled dUTP (Sigma). After hydrolysis with 10 µl of 1 M NaOH and 10 µl of 0.5 M EDTA for 15 min at 65°C, the samples were neutralized with the addition of 25 µl of 1 M Tris-HCl (pH 7.4) and cleanup was performed with Microcon 30 filters (Millipore). The fluorescent monofunctional *N*-hydroxysuccinimide-ester dyes cyanine-3 and cyanine-5 (Amersham) were coupled to the cDNAs originating from cultures grown without erythromycin and with erythromycin, respectively, for 1 h, quenched with 1.5 M hydroxylamine (Sigma), mixed, and finally cleaned with a QIAquick PCR purification kit (QIAGEN). The samples (30 µl) were then dried and used for hybridization within 12 h. Hybridizations to the microarrays were conducted as follows. Slides were prehybridized by incubation at 50°C for 30 min in a solution containing 1% bovine serum albumin (Calbiochem), 3.5× SSC, and 0.1% (wt/vol) SDS. The dried fluorescently labeled cDNA samples were resuspended in the following hybridization solution (40 µl): 5× SSC, 0.1% (wt/vol) SDS, 1.0% (wt/vol) bovine serum albumin, 50% (vol/vol) formamide, and 0.01% (wt/vol) single-stranded salmon sperm DNA. The resuspended probes were added to the arrays and incubated, in darkness, at 42°C for 6 h. After hybridization, excess hybridization solution and unbound probe were washed away by four washing steps and gentle shaking, in darkness, for 2 min in 2× SSC-0.1% SDS, 1 min in 1× SSC, 1 min in 0.2× SSC, and 30 seconds in 0.05× SSC. Immediately after washing, the arrays were dried by centrifugation at 600 rpm for 5 min in an Eppendorf 5810R tabletop centrifuge. Three replicate hybridizations were performed with two separate batches of RNA. The two batches of RNA were obtained in two separate growth experiments.

Hybridized arrays were scanned at wavelengths of 532 nm (cyanine-3) and 635 nm (cyanine-5) at a 10-µm resolution to obtain two TIFF images with a ScanArrayExpress Microarray scanner (Packard Bioscience). Fluorescent intensities and spot morphologies were analyzed using the QuantArray program ver. 3.0 (Packard BioScience), and spots were excluded based on slide or morphology abnormalities.

Raw data from each array was preprocessed independently. A Lowess smoothed background was subtracted from all foreground intensities, and a cross-validated Lowess method was used in an intensity-dependent normalization of every array. The log₂ ratios for each spot were further analyzed using a mixed model (30) to detect differentially expressed genes. A mixed model was fitted to the data for each of the five sample times (0, 15, 30, 60, and 90 min) separately. Data for the three arrays at every sample time were described by $y_{ijk} = \mu_i + u_{ij} + e_{ijk}$ where y_{ijk} is the observed log₂ ratio of gene i (1, ..., 3,502) on array j (1, 2, 3) and in spot k (1, ..., 5) on that array, μ_i is the expected log₂ ratio for gene i , u_{ij} is a random effect of gene i on array j , and e_{ijk} is the remaining noise. The variance components were estimated under the assumption of Gaussian errors using a restricted maximum likelihood approach coping with the unbalanced data due to missing spots. Differentially expressed genes were identified by testing the hypothesis H_0 , defined as $\mu_i = 0$, against H_1 , defined as $\mu_i \neq 0$. A chi-square test for every gene resolves this for the model in the y_{ijk} equation (4), and a Bonferroni-corrected rejection level of a P value of <0.01 was used throughout. If H_0 ($\mu_i = 0$) was rejected, and μ_i is >0, genes were considered to be up-regulated in the erythromycin-treated cells. If H_0 was rejected, and μ_i is <0, genes were considered to be down-regulated. All data analysis algorithms were programmed by using Matlab (MathWorks, Inc.), but a subset of the data was also analyzed by the SAS system (SAS Institute, Inc.) to check the validity of the code.

Confirmation of expression levels of specific genes by real-time RT-PCR. To confirm independently the differential gene expression observed by microarray experiments, four genes were selected for analysis by real-time quantitative RT-PCR (RTQ). Primers and probes for the RTQ were designed using the Assays-

By-Design file builder (ver. 2.0; http://www.appliedbiosystems.com/support/software/assaysbydesign/installs.cfm?prod_id=1541; Applied Biosystems). Primers and probes were synthesized by and purchased from Applied Biosystems. The genes selected for RTQ analyses were EF0633 (*tyrS-1*, encoding tyrosyl-tRNA synthetases), EF2653 (encoding a transcriptional regulator of the Cro/CI family), and EF0105 (*argF-1*, encoding ornithine carbamoyltransferase). EF1964 (*gap-2*, encoding glyceraldehyde-3-phosphate dehydrogenase), which is constitutively expressed, was used to normalize the *TaqMan* data. The RTQ analyses were run on an ABI PRISM 7700 sequence detector (Applied Biosystems). cDNA was synthesized as above, with 15 ng RNA as template. Real-time PCR was performed using the *TaqMan* Universal PCR Master Mix (Applied Biosystems), according to the manufacturer's instructions. To ensure that the cDNA was not contaminated by genomic DNA, reactions without reverse transcriptase were also included. Differential expression was determined by calculating the change in threshold cycle (ΔC_t) for each gene, with RNA isolated from cells grown with and without erythromycin and harvested at the five time points mentioned above.

RESULTS

We have studied the transcriptional responses of *E. faecalis* V583 to treatment with the macrolide antibiotic erythromycin through a time course experiment utilizing a genome-wide microarray based on the V583 genome sequence. By applying a time course experiment in this manner, we were able to investigate both an immediate response to this stress and the cells' adaptation to a more permanent presence of erythromycin. In the following, we focus on genes that were differentially expressed in the untreated and the erythromycin-treated cells.

We applied a stringent confidence level (a *P* value of <0.01 plus Bonferroni correction for multiple comparisons) for the determination of significant differential transcription. Genes were "scored" as significantly up- or down-regulated as described above, with a threshold *P* value of <0.01 and the conservative Bonferroni correction for multiple comparisons, ensuring a very low number of false negatives.

Circa 600 (18%) of the predicted V583 genes were found to be differentially transcribed at one or more of the five time points examined during the 90-min time course experiment. Obviously, the erythromycin exposure seriously affected the transcriptional events in these enterococci: 260 genes were found to be significantly up-regulated (induced; \log_2 ratios of stressed cells/nonstressed cells were different from and higher than 0) and 340 genes down-regulated (repressed; \log_2 ratios of stressed cells/nonstressed cells were different from and lower than 0) at one or more time points. Among the up-regulated genes, four were plasmid encoded, while 27 plasmid-encoded genes were down-regulated. The numbers of differentially expressed genes, sorted by their cellular roles, are shown in Table 1. The total number of V583 genes represented on the array was 3,160 (out of total of 3,337 genes in the V583 genome), which means that ca. 8% of all V583 genes were up-regulated and ca. 10% down-regulated in the erythromycin-exposed cells. This shows that transcriptional events in these enterococcal cells are strongly altered by erythromycin exposure. During the 90-min time course, the number of down-regulated genes is higher than the number of induced genes at all time points except *t*90 (the last time point) (Tables 2 and 3), which indicates a general decrease in transcriptional activity by exposure to erythromycin.

Growth of erythromycin-treated *E. faecalis* V583. Cell samples were collected at five time points after the addition of erythromycin (50 μ g/ml) to one of the cultures. We chose this level of erythromycin in the cultures on the basis of preliminary

TABLE 1. Number of genes that were differentially expressed at one or more time points, sorted by cellular role^a

Cellular role	No. of differentially expressed genes	
	Down-regulated	Up-regulated
Amino acid biosynthesis	5	2
Biosynthesis of cofactors/prosthetic groups/carriers	6	1
Cell envelope	24	12
Cellular processes	28	8
Central intermediary metabolism	5	3
DNA metabolism	5	16
Energy metabolism	30	11
Fatty acid and phospholipid metabolism	1	6
Hypothetical proteins	118	83
Other categories	8	1
Protein fate	9	8
Protein synthesis	14	13
Purine/pyrimidine/nucleoside/nucleotide	11	8
Regulatory functions	17	14
Signal transduction	17	9
Transcription	3	3
Transport and binding protein	40	49
Unknown function	23	17
Viral functions	1	0

^a Differentially expressed plasmid-encoded genes are also included. Genes that were up-regulated at some time points and down-regulated at other time points are counted twice. Likewise, genes that have been assigned more than one cellular role were counted twice.

experiments (results not shown), where we found that growth of V583 was retarded but still significant. (V583 does grow in medium with >400 μ g erythromycin/ml.) Our time course experiment was performed over a period of 90 min, and in this first part of the growth, the erythromycin-treated cells grow more slowly than the untreated cells. Thus, this work describes

TABLE 2. Number of up-regulated genes in each functional category at all time points studied

Cellular role	No. of up-regulated genes at time point indicated				
	<i>t</i> 0	<i>t</i> 15	<i>t</i> 30	<i>t</i> 60	<i>t</i> 90
Amino acid biosynthesis	2	0	0	0	0
Biosynthesis of cofactors/prosthetic groups/carriers	0	1	1	0	0
Cell envelope	1	4	5	5	5
Cellular processes	0	2	6	5	7
Central intermediary metabolism	2	2	3	1	0
DNA metabolism	1	7	6	11	6
Energy metabolism	0	1	3	2	7
Fatty acid and phospholipid metabolism	0	4	5	2	3
Hypothetical protein	4	25	45	36	48
Other categories	0	0	1	0	0
Protein fate	0	2	4	5	5
Protein synthesis	0	8	9	5	3
Purine/pyrimidine/nucleoside/nucleotide	0	3	4	1	4
Regulatory functions	0	4	7	7	9
Signal transduction	0	2	4	2	5
Transcription	0	1	1	1	1
Transport and binding protein	6	18	24	21	28
Unknown function	2	5	9	11	9
Viral functions	0	0	0	0	0
Total	18	89	137	115	140

TABLE 3. Number of down-regulated genes in each functional category at all time points studied

Cellular role	No. of down-regulated genes at time point indicated				
	t0	t15	t30	t60	t90
Amino acid biosynthesis	1	1	2	3	4
Biosynthesis of cofactors/prosthetic groups/carriers	1	4	4	0	1
Cell envelope	5	10	16	10	9
Cellular processes	1	10	10	7	10
Central intermediary metabolism	1	2	2	1	1
DNA metabolism	0	4	4	0	1
Energy metabolism	3	4	12	21	11
Fatty acid and phospholipid metabolism	0	0	0	0	1
Hypothetical protein	29	59	47	35	43
Other categories	0	2	3	2	3
Protein fate	0	2	2	4	4
Protein synthesis	0	6	6	5	6
Purine/pyrimidine/nucleoside/nucleotide	0	4	3	6	5
Regulatory functions	2	7	6	3	4
Signal transduction	1	2	4	11	3
Transcription	0	1	2	0	0
Transport and binding protein	6	11	18	16	12
Unknown function	6	8	9	6	9
Viral functions	0	1	1	0	0
Total	56	138	151	130	127

the response of V583 to erythromycin during the first cycles of growth. We chose a relatively low level of erythromycin for the experiments, according to the observation by Hutter et al. (11) that lower concentrations are better for obtaining good microarray results. During the experiments, growth was measured spectrophotometrically. After 15 min, the OD of the erythromycin-treated culture was approximately 82% of the OD of the untreated (control) culture; after 30 min, the OD of the erythromycin-treated culture was approximately 69% of that of the untreated culture; after 60 min, the OD of the erythromycin-treated culture was approximately 54% of that of the untreated culture; and after 90 min, the OD of the treated

culture was 40% of the OD of the untreated culture. The growth curves of erythromycin-treated and untreated V583 are shown in Fig. 1.

EFA0007, the *ermB* gene. EFA0007, which was referred to as an rRNA adenine dimethylase family protein encoded on pTEF1 (20) is, in fact, ErmB (>99% identity with ErmB from *Bacillus cereus*, *Streptococcus pyogenes*, *E. faecium* [BlastP]). This gene was expressed in cultures both with and without erythromycin, but the gene was not differentially expressed at any time point. This is in accordance with earlier observations that the *ermB* is constitutively expressed in enterococci (B. Murray, unpublished results). The retarded cell growth in the presence of erythromycin confirms that EFA0007 does not fully protect the bacterium against this antimicrobial compound and suggests that additional genes are involved in the cells' protection against this stress factor. Alternatively, the apparent lack of differential expression may be explained by posttranscriptional regulation of this gene; *ermB* may be posttranscriptionally regulated by a stalling on the ribosome during translation.

Up-regulated (induced) genes. The number of up-regulated genes in each functional category at various time points is shown in Table 2. Among the up-regulated genes, the genes encoding hypothetical proteins represented the highest percentage (85 genes; around 32% of all up-regulated genes), while genes related to transport and binding functions represented around 17% (44 genes) of the genes induced by the erythromycin stress. Moreover, genes encoding proteins involved in energy metabolism (10 genes), protein synthesis (13 genes), synthesis of cell envelope components (12 genes), and regulatory functions (13 genes) were up-regulated. The majority of up-regulated genes related to protein synthesis are genes encoding ribosomal proteins. In addition, genes classified in the category of genes of unknown function represent a considerable part of the induced genes (17 genes).

Among the up-regulated genes, three genes encoded drug

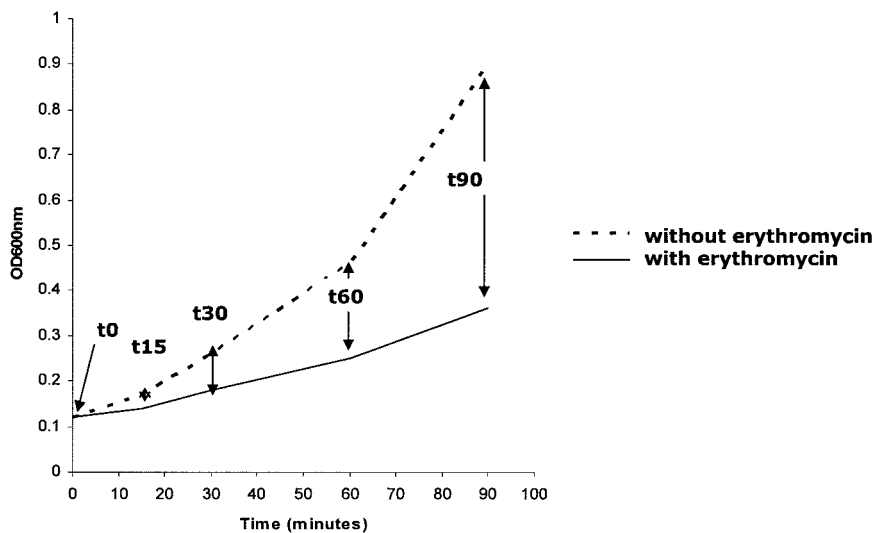


FIG. 1. Growth of *E. faecalis* V583 treated with erythromycin (50 µg/ml; solid line) and of untreated *E. faecalis* V583 (broken line). Growth was measured spectrophotometrically at a λ of 600 nm. Arrows indicate time points at which samples were collected for RNA extraction and microarray experiments.

resistance proteins. These genes were EF0420 (up-regulated at *t*30 and *t*60), EF1370 (up-regulated at *t*30, *t*60, and *t*90), and EF1078 (up-regulated at *t*15, *t*30, *t*60, and *t*90). Four genes only were induced at all time points. These genes were EF1400 (predicted to encode a cadmium-translocating ATPase), EF1413, EF1916 (encoding a GTP-binding protein), and EF2720 (encoding an ABC transporter). The gene encoding a putative MsrC-like protein (EF1413) is of particular interest. MsrC is widely spread among *Enterococcus faecium*, and MsrC is believed to be an efflux pump involved in low-level macrolide resistance in this species (23, 25). However, the EF1413 is only 40% identical (60% similar) to the *E. faecium* MsrC, and it is therefore not clear whether the EF1413 encodes a functional MsrC-like protein. Table 4 gives the full list of genes found to be induced (up-regulated), with \log_2 ratios. Genes predicted to encode hypothetical proteins and plasmid encoded genes are excluded from this list.

Down-regulated (repressed) genes. The numbers of down-regulated genes in each functional category at various time points are shown in Table 3. Among the down-regulated genes, those encoding hypothetical proteins are the dominant category (115 genes; 35%), followed by genes encoding transport and binding proteins (38 genes; 11%), genes with unknown function (23 genes), genes encoding cell envelope-related proteins (24 genes), and genes involved in energy metabolism (29 genes), regulatory function (13 genes), protein synthesis (14 genes), and cellular processes (23 genes). The majority of down-regulated genes related to protein synthesis are eight tRNA synthetases (cysteinyl-tRNA synthetase, tyrosyl-tRNA synthetase, alanyl-tRNA synthetase, aspartyl-tRNA synthetase, tryptophanyl-tRNA synthetase [EF2228], threonyl-tRNA synthetase, valyl-tRNA synthetase, and seryl-tRNA synthetase). The tryptophanyl-tRNA synthetase gene encoded by EF2679 (*trpS*) was significantly up-regulated at all time points except *t*0. Transcription was found to be repressed at all five time points examined for 10 genes. Table 5 lists all of the down-regulated genes, with \log_2 ratios. Genes encoding hypothetical proteins and plasmid genes are not included.

Validation of microarray data by real-time quantitative RT-PCR analyses of selected genes. Verifications of the expression levels of four genes observed by microarrays were performed by real-time quantitative RT-PCR analyses (*TaqMan* assays). The genes selected for these RTQ analyses represented various functional categories and different patterns of expression. The transcription profiles of the genes examined by RTQ correlated well with the transcription profiles observed by the use of microarrays ($r = 0.83$).

DISCUSSION

The present study focuses on functional genomics of *Enterococcus faecalis* V583, applying a genome-wide microarray based on the genome sequence of V583. In general, the knowledge of gene expression in enterococci is scarce. This is the first paper describing a global transcriptional profile of *Enterococcus*, and more studies are definitely needed to obtain a deeper understanding of gene expression in this species. We used a microarray to study the transcriptional responses of V583 to the macrolide antibiotic erythromycin. V583 is resistant to relatively high levels of erythromycin, but exposure to eryth-

romycin retards its growth. The work was performed to (i) gain insight into the mechanisms involved in resistance to erythromycin by V583, (ii) obtain clues about transcriptional responses to general stress in bacteria, and (iii) generate hypotheses for subsequently gaining deeper knowledge on antibiotic resistance and general stress in this bacterium.

In the experiments, V583 was treated with erythromycin (50 $\mu\text{g/ml}$), which decreased the growth of the cells significantly compared to the untreated control during the 90-min time course experiment. The concentration of antibiotic to apply in such experiments is, of course, a matter of discussion. Hutter et al. (11) claimed that compound concentrations should be at concentrations that are just low enough not to affect the growth of the organism. This probably varies with the strains and compounds being studied.

We chose a stringent confidence level plus Bonferroni correction to score for significant differential expression, in the data analysis. Consequently, the number of false positives is, per definition, very low. We preferred this method, although some information might have been lost. Moreover, we decided to score for significant up- and down-regulation instead of using the common fold change cutoff values. Thus, the conclusions are based on experimental data precision (noise levels and number of replicates); this is appropriate since microarrays probably underestimate actual mRNA induction ratios (2). Also, treating each array as a random sample from a population of all possible arrays makes all conclusions more general. Even our simple mixed model gives a quite accurate description of the variance structure of normalized microarray data from repeated experiments and allows for an extended information extraction from every experiment. Our analysis differs from that by Wernisch et al. (30) in that we have excluded the array main effect. Our argument for this is that all arrays have been normalized separately, and thus any array main effects will be ignorable, which is also the conclusion drawn by Wernisch et al. (30) in analyzing similar data. Another variant of our mixed model approach is taken by Wolfinger et al. (31). They use one variance component per gene, which makes the model much more complex. More research must be done in this area of bioinformatics before we can come to a conclusion as to which strategy is appropriate. For now, we have chosen to use a simple model, in line with the principle of parsimony.

The target of erythromycin and other macrolides is protein synthesis, more specifically the 50S (large) ribosomal subunit. These antibiotics block the ribosome exit tunnel, which prevents the movement and release of the nascent peptide chain. The *ermB* gene (EFA0007) was not found to be significantly up- or down-regulated at any time point. Hence, this gene is probably constitutively expressed and contributes to the erythromycin resistance of V583. However, based on the high level of differential transcription and slow growth observed for erythromycin-exposed cells compared to untreated cells, it appears that a battery of other genes respond expressionally to the presence of erythromycin. These responses are not necessarily all due to resistance, but rather an adaptation to changes in growth conditions. Thus, these genes contribute to maintenance of the growth during erythromycin exposure. Three genes encoding drug resistance proteins (EF0420, EF1078, and EF1370), on the other hand, appear to play a role

TABLE 4. Log₂ ratios of genes whose transcription increased at one or more of the five time points (altogether 90 min) at which the effect of erythromycin treatment was studied, sorted by functional category (cellular role)

Protein encoded by gene	Cellular role(s)	ORF	Log ₂ ratio at:				
			t0	t15	t30	t60	t90
Cystathionine beta-lyase (<i>metC</i>) ^a	Amino acid biosynthesis	EF0290	0.71	0	0	0	-1.05
Prephenate dehydrogenase	Amino acid biosynthesis	EF1565	0.71	0	0	0	0
Cobyrinic acid synthase, putative	Biosynthesis of cofactors/prosthetic groups/carriers	EF2586	0	0.87	0.86	0	0
Alanine racemase (<i>alr</i>)	Cell envelope	EF0849	0	0	0.78	0.87	1.11
Membrane protein, putative	Cell envelope	EF0860	0	0	0	0	0.77
Glycosyl transferase, group 2 family protein	Cell envelope	EF0887	0	0.61	0	0.98	0
Extracellular protein, putative	Cell envelope	EF0944	0	0.72	0	0	0
UDP-N-acetylglucosamine-N-acetylmuramyl-(pentapeptide) pyrophosphoryl-undecaprenol N-acetylglucosamine transferase (<i>murG</i>)	Cell envelope	EF0994	0	0.64	0	0	0
Membrane protein, putative	Cell envelope	EF1384	0	0	0	0.79	0.81
Bacterial sugar transferase	Cell envelope	EF2177	0	0.75	0.71	0	0
Glycosyl transferase, group 1 family protein	Cell envelope	EF2890	0	0	0.60	0	0
Glycosyl transferase, group 1 family protein	Cell envelope	EF2891	0	0	0.69	0.83	0
Membrane protein	Cell envelope	EF3176	0.74	0	0	0	0
Adhesion lipoprotein	Cell envelope/cellular processes	EF3206	0	0	0	0.96	0.70
NAD-dependent epimerase/dehydratase family protein	Cell envelope/energy metabolism	EF2165	0	0	0.74	0	0.71
Cell division protein, FtsW/RodA/SpovE family	Cellular processes	EF1300	0	0	0	0.86	1.23
MsrC protein, putative	Cellular processes	EF1413	1.88	2.96	3.24	3.36	3.44
Beta-lactamase, putative	Cellular processes	EF1502	0	0	0.62	0	0
Competence protein (<i>comG3</i>)	Cellular processes	EF2044	0	0	0.65	0	0.76
Competence protein (<i>comG2</i>)	Cellular processes	EF2045	0	0	0	0.73	0.85
Competence protein (<i>comG1</i>)	Cellular processes	EF2046	0	0	0.73	0.90	0
Chromosome partitioning protein ParB	Cellular processes	EF3298	0	0	0	0	0.69
ATPase, ParA family	Cellular processes	EF3299	0	0	0	0	0.89
Drug resistance transporter, EmrB/QacA family protein	Cellular processes/transport and binding protein	EF0420	0	0	0.69	0.98	0
Multidrug resistance protein, putative	Cellular processes/transport and binding protein	EF1078	0	0.78	1.24	1.18	0.93
Drug resistance transporter, EmrB/QacA family protein	Cellular processes/transport and binding protein	EF1370	0	0	0.83	0.97	0.68
Cell division ABC transporter, permease protein FtsX, putative	Cellular processes/transport and binding protein	EF1760	0	0.80	1.13	0.72	0.69
Glycerol dehydrogenase, putative	Central intermediary metabolism	EF1358	0	0	0.66	0	0
Alkaline phosphatase (<i>phoZ</i>)	Central intermediary metabolism	EF2973	0.79	0.85	0.62	0.70	0
Heptaprenyl diphosphate synthase, component II, putative	Central intermediary metabolism	EF3260	0.83	0.84	0.84	0	0
Chromosomal replication initiator protein DnaA	DNA metabolism	EF0001	0	0	0	0	0.65
MutT/nudix family protein	DNA metabolism	EF0780	0	0.70	0	0.95	0
DNA repair exonuclease family protein	DNA metabolism	EF0972	0	0	0	1.01	0
DNA polymerase III, alpha subunit (<i>dnaE</i>)	DNA metabolism	EF1044	0	0.65	0	0.71	0
Endonuclease III (<i>nth</i>)	DNA metabolism	EF1155	0	0	0.66	0	0
ATP-dependent DNA helicase RecQ	DNA metabolism	EF1545	0	0.76	0.88	1.00	1.08
MutT/nudix family protein	DNA metabolism	EF1587	0	0	0	0	0.78
Site-specific recombinase, phage integrase family	DNA metabolism	EF1648	0	0	0	0.73	0.81
Endonuclease IV (<i>nfo</i>)	DNA metabolism	EF1736	0	0.61	0.75	0.67	0
Site-specific recombinase, phage integrase family	DNA metabolism	EF2043	0	0	0	0.66	0
Toprim domain protein	DNA metabolism	EF2305	0	0	0	0.76	0.76
A/G-specific adenine glycosylase (<i>mutY</i>)	DNA metabolism	EF2704	0	0.65	0.68	0	0
Regulatory protein RecX, putative	DNA metabolism	EF2705	0	0.65	0	0.94	0
Exonuclease	DNA metabolism	EF2736	0.91	0	0.97	0	0
ATP-dependent DNA helicase RecG	DNA metabolism	EF3113	0	0.60	0	0.74	0.69
Beta-phosphoglucomutase (<i>pgmB</i>)	Energy metabolism	EF0956	0	0	0	-0.90	1.27
Glycosyl hydrolase, family 65	Energy metabolism	EF0957	0	0	0	-1.12	1.36
Galactose-1-phosphate uridylyltransferase (<i>galT</i>)	Energy metabolism	EF1071	0	0	0.60	0	0
Glycosyl hydrolase, family 13	Energy metabolism	EF1347	0	0	0	-0.99	0.80
Glucan 1,6-alpha-glucosidase, putative	Energy metabolism	EF1348	0	0	0	-1.01	0.77
Glycosyl hydrolase, family 13	Energy metabolism	EF1349	0	0	0	-0.99	0.93
Glycosyl hydrolase, family 4	Energy metabolism	EF1411	0	0	0	0	0.87
Ferredoxin (<i>fer</i>)	Energy metabolism	EF1543	0	0	0.60	0.79	0
Glycosyl hydrolase, family 1	Energy metabolism	EF1606	0	0.69	0.70	0.70	0
Glycerate kinase, putative	Energy metabolism	EF2646	0	0	0	-1.65	0.70
3-Oxoacyl-(acyl-carrier-protein) synthase II (<i>fabF-1</i>)	Fatty acid and phospholipid metabolism	EF0283	0	0.83	0	0	0
Holo-(acyl-carrier-protein) synthase (<i>acpS</i>)	Fatty acid and phospholipid metabolism	EF0848	0	0	0.69	0.67	0.91
Cardiolipin synthetase, putative	Fatty acid and phospholipid metabolism	EF1608	0	0.71	0.63	0	0
1-Acyl-sn-glycerol-3-phosphate acyltransferase, putative	Fatty acid and phospholipid metabolism	EF2691	0	1.06	0.65	0.74	1.09
Enoyl-(acyl-carrier-protein) reductase II (<i>fabK</i>)	Fatty acid and phospholipid metabolism	EF2883	0	0.67	0.72	0	0.89
Fatty acid/phospholipid synthesis protein PlsX	Fatty acid and phospholipid metabolism	EF3112	0	0	0.64	0	0
TraE protein, putative	Other categories	EF2320	0	0	0.61	0.69	0

Continued on following page

TABLE 4—Continued

Protein encoded by gene	Cellular role(s)	ORF	Log ₂ ratio at:				
			t0	t15	t30	t60	t90
ATP-dependent Clp protease, ATP-binding subunit ClpE	Protein fate	EF0706	0	0	0.79	0	0
DnaJ protein	Protein fate	EF1310	0	0	0	0	0.79
Heat shock protein HslV	Protein fate	EF1647	0	0	0	0.74	0
Signal recognition particle protein (<i>flh</i>)	Protein fate	EF1700	0	0.74	0.80	0.81	1.05
Preprotein translocase, SecE subunit	Protein fate	EF2730	0	0	0.75	1.17	0.71
Signal peptidase I	Protein fate	EF3073	0	0.64	0	0	0.72
Ribosomal protein L18 (<i>rplR</i>)	Protein synthesis	EF0223	0	0.66	0.75	0	0
Ribosomal protein S5 (<i>rpsE</i>)	Protein synthesis	EF0224	0	0.58	0.64	0	0
Ribosomal protein L30 (<i>rplM</i>)	Protein synthesis	EF0225	0	0.60	0.61	0	0
Ribosomal protein L15 (<i>rplO</i>)	Protein synthesis	EF0226	0	0.66	0.92	0	0
Ribosomal protein L27 (<i>rplA</i>)	Protein synthesis	EF0970	0	0.73	0	0	0
Ribosomal protein S16 (<i>rpsP</i>)	Protein synthesis	EF1694	0	0	0.73	0	0
Ribosome recycling factor (<i>frr</i>)	Protein synthesis	EF2395	0	0	0.81	0.77	0
Translation elongation factor Ts (<i>tsf</i>)	Protein synthesis	EF2397	0	0.72	0.82	0	0
Ribosomal-protein-alanine acetyltransferase, putative	Protein synthesis	EF2473	0	0.63	0	0.75	0
Ribosomal-protein-alanine acetyltransferase, putative	Protein synthesis	EF2474	0	0	0	0	0.66
Tryptophanyl-tRNA synthetase (<i>trpS</i>)	Protein synthesis	EF2679	0	0.86	1.23	1.01	0.82
Ribosomal protein L33 (<i>rplM</i>)	Protein synthesis	EF2731	0	0	0.62	0.99	1.17
Ribosomal protein S4 (<i>rpsD</i>)	Protein synthesis	EF3070	0	0	0	0.66	0
Dihydroorotate dehydrogenase (<i>pyrD-1</i>)	Purine/pyrimidine/nucleoside/nucleotide	EF0285	0	0.82	0.67	0	0
Dihydroorotate dehydrogenase (<i>pyrD-2</i>)	Purine/pyrimidine/nucleoside/nucleotide	EF1714	0	0.71	0	0	0
Dihydroorotate (<i>pyrC</i>)	Purine/pyrimidine/nucleoside/nucleotide	EF1718	0	0	0	0	0.66
Phosphoribosylaminoimidazole carboxylase, ATPase subunit (<i>purK-2</i>)	Purine/pyrimidine/nucleoside/nucleotide	EF2362	0	0	0	0	0.68
Xanthine phosphoribosyltransferase (<i>xpt</i>)	Purine/pyrimidine/nucleoside/nucleotide	EF2365	0	0	0.79	1.04	0
Uridylate kinase (<i>pyrH</i>)	Purine/pyrimidine/nucleoside/nucleotide	EF2396	0	0.76	0.68	0	0
Inosine-uridine preferring nucleoside hydrolase	Purine/pyrimidine/nucleoside/nucleotide	EF2587	0	0	0.79	0	0.90
2',3'-Cyclic-nucleotide 2'-phosphodiesterase, putative	Purine/pyrimidine/nucleoside/nucleotide	EF2902	0	0	0	0	0.80
Transcriptional regulator, DeoR family	Regulatory functions	EF0719	0	0	0	1.78	0
Transcriptional regulator, PemK family	Regulatory functions	EF0850	0	0	0.67	0.84	0.95
DNA-binding response regulator	Regulatory functions	EF1050	0	0	0.76	0	0
Transcriptional regulator, Cro/CI family	Regulatory functions	EF1369	0	0.68	1.18	1.40	1.13
Transcription antiterminator BglG family protein	Regulatory functions	EF1515	0	0.69	0.62	0	0.89
Protease synthase and sporulation negative regulatory protein pai 1	Regulatory functions	EF1590	0	0.70	0.65	0.66	0.87
Sucrose operon repressor ScrR (<i>scrR-1</i>)	Regulatory functions	EF1604	0	0	0.65	1.05	0.75
Transcriptional regulator (<i>codY</i>)	Regulatory functions	EF1645	0	0	0.69	0.73	0.78
Transcriptional regulator, TetR family	Regulatory functions	EF2066	0	0	0	0.68	0
Phosphosugar-binding transcriptional regulator, putative	Regulatory functions	EF2259	0	0	0	0	0.70
Transcriptional regulator, Cro/CI family	Regulatory functions	EF2653	0	0	0	0	0.94
Transcriptional regulator, Cro/CI family	Regulatory functions	EF2852	0	0	0	0	0.93
Zinc-binding transcriptional regulator, Cro/CI family	Regulatory functions	EF3272	0	0.84	0	0	0
PTS system component, mannose-specific IIC component	Signal transduction	EF0021	0	0	0	-1.22	1.04
PTS system, mannose-specific IID component	Signal transduction	EF0022	0	0	0	-1.34	1.04
PTS system, fructose-specific family, IIABC component	Signal transduction	EF0717	0	0	0	0.90	0
PTS system, IIABC component	Signal transduction	EF0958	0	0	0	-1.27	0
Sensor histidine kinase	Signal transduction	EF1051	0	0	0.69	0	0
Sensor histidine kinase	Signal transduction	EF1261	0	0	0.65	0	0
PTS system, IIABC component	Signal transduction	EF1516	0	1.13	1.44	1.06	1.06
PTS system, IIC component	Signal transduction	EF2978	0	0	0	0	0.66
PTS system, IIB component	Signal transduction	EF2979	0	0.69	0.72	0	0.94
ATP-dependent RNA helicase, DEAD/DEAH box family	Transcription	EF0846	0	0	0	0.68	0
Transcription termination factor Rho	Transcription	EF1170	0	0	0.68	0	0.84
ATP-dependent helicase, DEAH-box family, putative	Transcription	EF3214	0	0.62	0	0	0
Iron compound ABC transporter, substrate-binding protein Na ⁺ /H ⁺ antiporter (<i>nhaC-2</i>)	Transport and binding protein	EF0188	0	0	0	0	0.82
Sodium/dicarboxylate symporter family protein	Transport and binding protein	EF0636	0	0.64	0	1.01	0
Amino acid ABC transporter, ATP-binding protein	Transport and binding protein	EF0744	0	1.44	1.20	0.87	0.71
ABC transporter, ATP-binding/permease protein	Transport and binding protein	EF0760	0	0	0.63	0	0
ABC transporter, ATP-binding/permease protein	Transport and binding protein	EF0789	0	0.70	1.08	1.09	0
ABC transporter, ATP-binding/permease protein	Transport and binding protein	EF0790	0	0	0.75	0.96	0
Potassium uptake protein	Transport and binding protein	EF0872	0	0.65	0	0	0
ABC transporter, ATP-binding/permease protein	Transport and binding protein	EF0941	0	0	0	0.72	0
Amino acid permease family protein	Transport and binding protein	EF1103	0	0	0.84	0.93	0.66
ABC transporter, ATP-binding protein	Transport and binding protein	EF1255	0.94	0.75	0	0	0
ABC transporter, ATP-binding/permease protein	Transport and binding protein	EF1341	0	0.69	0	0	0
Sugar ABC transporter, permease protein	Transport and binding protein	EF1344	0	0.62	0	-0.93	1.25
Cadmium-translocating P-type ATPase	Transport and binding protein	EF1400	0.65	0.71	0.86	0.78	1.09
V-type ATPase, subunit B	Transport and binding protein	EF1499	0	0	0	0	0.79
Iron compound ABC transporter, ATP-binding protein	Transport and binding protein	EF1639	0	0	0.62	0	0.95
Iron compound ABC transporter, permease protein	Transport and binding protein	EF1640	0	0.60	0.74	0.67	1.04

Continued on facing page

TABLE 4—Continued

Protein encoded by gene	Cellular role(s)	ORF	Log ₂ ratio at:				
			t0	t15	t30	t60	t90
ABC transporter, ATP-binding/permease protein, MDR family	Transport and binding protein	EF1732	0	1.18	0.86	0.78	1.69
ABC transporter, ATP-binding/permease protein, MDR family	Transport and binding protein	EF1733	0	1.13	0.95	0.81	1.54
Amino acid permease family protein	Transport and binding protein	EF2047	0	0	0.90	1.21	0
Transport ATP-binding protein CydD, putative	Transport and binding protein	EF2058	0	0	0.68	0	0
Transport ATP-binding protein CydC, putative	Transport and binding protein	EF2059	0	0	0.61	0	0.83
ABC transporter, ATP-binding/permease protein	Transport and binding protein	EF2226	0	0	0	1.00	0.70
Xanthine permease	Transport and binding protein	EF2364	0.73	0	0.97	1.07	0
Amino acid permease	Transport and binding protein	EF2377	0	0.63	0.74	0.92	0
ABC transporter, permease protein	Transport and binding protein	EF2485	0	0	0.62	0	0
AzIC protein	Transport and binding protein	EF2509	0	0	0	0.75	0
Glycine betaine/L-proline transport ATP binding subunit	Transport and binding protein	EF2641	0	0	0	1.30	0
Glycine betaine/L-proline ABC transporter, glycine betaine/L-proline-binding/permease protein	Transport and binding protein	EF2642	0	0	0.82	1.75	0.86
Permease, GntP family	Transport and binding protein	EF2647	0	0	0	-2.31	1.32
Spermidine/putrescine ABC transporter	Transport and binding protein	EF2649	0	0	0	0	1.05
Spermidine/putrescine ABC transporter	Transport and binding protein	EF2650	0	0	0	0	0.92
Spermidine/putrescine ABC transporter	Transport and binding protein	EF2651	0	0	0	0	0.94
Spermidine/putrescine ABC transporter	Transport and binding protein	EF2652	0	0	0	0	0.67
ABC transporter, ATP-binding protein	Transport and binding protein	EF2720	0.93	0.69	1.12	0.86	0.80
ABC transporter, ATP-binding protein	Transport and binding protein	EF2769	0	0	-0.60	0	0.72
Ion transporter, putative	Transport and binding protein	EF2854	0	0	0	0	0.72
Potassium uptake protein TrkA	Transport and binding protein	EF2910	0	0.65	0.84	0.86	1.15
ABC transporter, ATP-binding protein	Transport and binding protein	EF2986	0	0	0.62	0	0
Cytosine/purines, uracil, thiamine, allantoin permease family protein	Transport and binding protein	EF3000	0	0	0	0	0.82
Formate/nitrite transporter family protein	Transport and binding protein	EF3069	0.82	0.87	0	0.73	0.66
Iron compound ABC transporter, permease protein	Transport and binding protein	EF3085	0	0	0	0	0.69
ABC transporter, permease protein	Transport and binding protein	EF3199	0	0	0.98	0	1.26
ABC transporter, ATP-binding protein	Transport and binding protein	EF3200	0	0.70	0.86	0	0.94
ABC transporter, permease protein	Transport and binding protein	EF3208	0	0.71	0.98	0	0
Oxidoreductase, DadA family	Unknown function	EF0414	0	0	0	0.72	0
LysM domain protein	Unknown function	EF0443	0.85	1.21	1.30	1.03	0
Glyoxalase family protein	Unknown function	EF0666	0	0.71	0	0	0
Methyltransferase, putative	Unknown function	EF0691	0	0	0	0.75	0
Type 2 phosphatidic acid phosphatase family protein	Unknown function	EF0796	0	0	0	0.69	0
DegV family protein	Unknown function	EF1191	0	0	0	0	0.77
Metallo-beta-lactamase YycJ	Unknown function	EF1197	0	0	0.90	0.93	0.92
GTP-binding protein	Unknown function	EF1527	0	0	0.77	0.68	0.74
GTPase, putative	Unknown function	EF1549	0	0	0.71	0	0
Acetyltransferase, GNAT family	Unknown function	EF1589	0	0	0	0	1.00
LacX protein, putative	Unknown function	EF1644	0	0.64	0.70	0.81	0
Lipase/acylhydrolase, putative	Unknown function	EF1683	0	0	0.69	0	0.67
GTP-binding protein	Unknown function	EF1916	0.99	1.02	1.39	1.09	1.18
GTP-binding protein LepA	Unknown function	EF2352	0	0	0.63	1.04	0
Diacylglycerol kinase catalytic domain protein	Unknown function	EF2661	0	0	0	0.86	0.92
Oxidoreductase, pyridine nucleotide-disulfide family	Unknown function	EF2899	0	0.75	0.73	1.06	1.02
Glucose-inhibited division protein B (<i>gidB</i>)	Unknown function	EF3300	0	0	0	0	0.67

^a Gene names in parentheses.

for the ability of V583 to survive erythromycin treatment. Their influence must, however, be limited, since the growth of V583 is so retarded by erythromycin.

It was expected that the exposure of V583 to erythromycin would affect genes involved in protein synthesis, despite the expression of *ermB*. The induction of eight genes encoding ribosomal proteins (r-proteins), along with two r-protein-Ala-acetyltransferases, might be an indication that the overproduction of ribosomal proteins is one way to evade the effect of erythromycin. To balance the number and function of the ribosomes, the cells may have to compensate by up-regulation of these genes. Five of the induced r-protein genes and one of the r-protein-Ala-acetyltransferase genes are induced after 15 min of growth, i.e., their induction is a quick response. Closely related to the known target of erythromycin in sensitive cells is

the fact that eight tRNA synthetase genes are repressed. The repression of tRNA synthetase genes may be seen as a logical consequence of the reduced ability of a cell's capacity to synthesize proteins, when an antibiotic binds a ribosomal subunit. The up-regulation of genes encoding r-proteins and two translation elongation factors, as well as down-regulation of tRNA synthetase genes, in response to translation inhibitors, has been noticed also by other authors (5, 19, 28).

Although the V583 cells are defined as resistant to erythromycin and will survive in the presence of relatively high levels of erythromycin, the cell growth is impaired, and, as shown above, the transcriptional activities are greatly altered. The differential transcription reflects the effect of erythromycin as well as the adaptation of V583 to the general stress. The retarded growth follows as a consequence of a slow-down of

TABLE 5. Log₂ ratios of genes whose transcription decreased at one or more of the five time points (altogether 90 min) at which the effect of erythromycin treatment was studied, sorted by functional category

Protein encoded by gene	Cellular role(s)	ORF	Log ₂ ratio at:				
			t0	t15	t30	t60	t90
Ornithine carbamoyltransferase (<i>argF-1</i>) ^a	Amino acid biosynthesis	EF0105	0	0	-1.37	-2.81	-2.19
Cysteine synthase B, putative	Amino acid biosynthesis	EF0289	0	0	0	0	-0.75
Cystathionine beta-lyase (<i>metC</i>)	Amino acid biosynthesis	EF0290	0.71	0	0	0	-1.05
Arginine repressor (<i>argR</i>)	Amino acid biosynthesis	EF0676	0	0	0	-0.94	0
Aspartate aminotransferase, putative	Amino acid biosynthesis	EF0891	-1.00	-1.08	-0.96	-0.74	-1.03
Phosphomevalonate kinase	Biosynthesis of cofactors/prosthetic groups/carriers	EF0902	-0.75	-0.73	0	0	0
Thiamin biosynthesis ApbE, putative	Biosynthesis of cofactors/prosthetic groups/carriers	EF1225	0	0	-0.92	0	0
NH ₃ -dependent NAD ⁺ synthetase (<i>nadE</i>)	Biosynthesis of cofactors/prosthetic groups/carriers	EF2625	0	0	0	0	-0.88
Phosphomethylpyrimidine kinase (<i>thiD</i>)	Biosynthesis of cofactors/prosthetic groups/carriers	EF2775	0	-0.83	-0.95	0	0
Thiamine-phosphate pyrophosphorylase (<i>thiE</i>)	Biosynthesis of cofactors/prosthetic groups/carriers	EF2776	0	-0.75	-0.76	0	0
Hydroxyethylthiazole kinase, putative	Biosynthesis of cofactors/prosthetic groups/carriers	EF2777	0	-0.63	-0.72	0	0
Adhesion lipoprotein	Cell envelope	EF0055	0	0	-0.84	0	0
Glycosyl hydrolase, family 20	Cell envelope	EF0114	0	0	0	-0.87	0
Surface exclusion protein, putative	Cell envelope	EF0146	-0.84	-0.85	-0.72	0	0
Chitinase, family 2	Cell envelope	EF0361	0	0	0	-0.86	-0.86
Chitin binding protein, putative	Cell envelope	EF0362	0	0	0	-1.09	-0.96
LemA protein	Cell envelope	EF0468	0	0	-1.11	0	0
Penicillin-binding protein, putative	Cell envelope	EF0746	0	-0.73	-1.39	0	-0.87
Gram-positive anchor protein, putative	Cell envelope	EF0775	-0.95	-1.10	-0.81	-0.67	-0.91
Membrane protein, putative	Cell envelope	EF1027	0	0	-0.60	0	0
Pheromone cAM373 precursor lipoprotein	Cell envelope	EF1340	0	-0.79	0	0	0
Coccolysin	Cell envelope	EF1818	0	0	0	-0.85	0
Endocarditis specific antigen	Cell envelope	EF2076	0	-0.76	-0.77	-1.12	-1.51
DltD protein	Cell envelope	EF2746	0	0	-0.65	0	0
D-Alanyl carrier protein (<i>dltC</i>)	Cell envelope	EF2747	-0.86	-0.79	-1.22	0	0
Basic membrane protein DtlB	Cell envelope	EF2748	0	0	-0.65	0	0
D-Alanine-activating enzyme, putative	Cell envelope	EF2749	0	0	-0.74	0	0
Sortase family protein	Cell envelope	EF3056	0	0	-0.77	-1.56	-1.32
Rod shape-determining protein MreD	Cell envelope	EF3061	0	0	0	-0.93	-1.28
Rod shape-determining protein MreC	Cell envelope	EF3062	0	0	0	-1.01	-1.21
Pheromone cAD1 precursor lipoprotein	Cell envelope	EF3256	0	-0.68	-0.73	0	0
Gls24 protein	Cellular processes	EF0079	0	-1.04	-0.97	-1.16	-1.39
Gls24 protein	Cellular processes	EF0080	0	-0.98	-1.20	-1.28	-1.87
Regulatory protein, putative (<i>pfoR</i>)	Cellular processes	EF0097	0	0	-0.78	0	0
Superoxide dismutase, Mn (<i>sodA</i>)	Cellular processes	EF0463	0	-0.65	0	0	0
Low-temperature requirement C protein, putative	Cellular processes	EF0639	0	0	0	0.96	-1.21
Polysaccharide lyase, family 8	Cellular processes	EF0818	0	-0.70	-0.89	0	-1.24
Universal stress protein family	Cellular processes	EF1058	0	-1.20	-1.28	-1.44	-2.27
Streptomycin 3-adenylyltransferase, putative	Cellular processes	EF1076	-0.71	-0.97	-0.64	-0.88	-0.77
Autoinducer-2 production protein LuxS	Cellular processes	EF1182	0	-0.68	-0.66	0	-0.99
Cold shock protein CspC	Cellular processes	EF1991	0	-0.98	0	0	0
Negative regulator of genetic competence MecA, putative	Cellular processes	EF2677	0	0	0	0	-0.66
Alkyl hydroperoxide reductase, C subunit (<i>ahpC</i>)	Cellular processes	EF2739	0	-0.70	-0.68	-0.68	-0.68
Dps family protein	Cellular processes	EF3233	0	0	-0.70	0	0
Nucleotidyltransferase family protein	Central intermediary metabolism	EF0137	0	-0.69	-0.87	0	-0.77
Hydrolase, alpha/beta hydrolase fold family	Central intermediary metabolism	EF1028	0	0	-0.70	0	0
6-Aminohexanoate-cyclic-dimer hydrolase, putative	Central intermediary metabolism	EF1033	0	0	0	-0.95	0
Microcompartment protein	Central intermediary metabolism	EF1623	-0.66	-0.76	0	0	0
Chromosomal replication initiator protein DnaA	DNA metabolism	EF0001	0	0	0	0	0.65
Thermonuclease precursor (<i>nuc-1</i>)	DNA metabolism	EF0511	0	0	0	0	-0.70
Exonuclease RexB	DNA metabolism	EF1112	0	-0.67	-0.64	0	0
MutT/nudix family protein	DNA metabolism	EF1141	0	-0.58	-0.59	0	0
L-Serine dehydratase, iron-sulfur-dependent, beta subunit (<i>sdhB-1</i>)	Energy metabolism	EF0098	0	0	-1.08	0	0
L-Serine dehydratase, iron-sulfur-dependent, alphasubunit (<i>sdhA-1</i>)	Energy metabolism	EF0099	0	0	-0.67	0	0
Arginine deiminase (<i>arcA</i>)	Energy metabolism	EF0104	0	-0.65	-1.73	-3.16	-2.55
Carbamate kinase (<i>arcC-1</i>)	Energy metabolism	EF0106	0	0	-0.92	-1.99	-1.26
Phosphoglycerate mutase 1 (<i>gpm</i>)	Energy metabolism	EF0195	0	0	-0.62	0	-0.65
Aldehyde dehydrogenase	Energy metabolism	EF0253	-0.74	-0.82	-0.90	-0.72	-0.91
L-Lactate dehydrogenase (<i>ldh-2</i>)	Energy metabolism	EF0641	0	0	0	-0.78	-0.75
Phosphoglucomutase/phosphomannomutase family protein	Energy metabolism	EF0677	0	0	0	-1.16	0

Continued on facing page

TABLE 5—Continued

Protein encoded by gene	Cellular role(s)	ORF	Log ₂ ratio at:				
			t0	t15	t30	t60	t90
Aldehyde-alcohol dehydrogenase (<i>adhE</i>)	Energy metabolism	EF0900	0	0	-0.77	0	0
Methylglyoxal synthase (<i>mgsA</i>)	Energy metabolism	EF0939	-0.88	-1.10	-0.96	-0.85	-0.96
Beta-phosphoglucomutase (<i>pgmB</i>)	Energy metabolism	EF0956	0	0	0	-0.90	1.27
Glycosyl hydrolase, family 65	Energy metabolism	EF0957	0	0	0	-1.12	1.36
Glycosyl hydrolase, family 1	Energy metabolism	EF1020	0	0	0	-1.93	0
Aldose 1-epimerase (<i>galM</i>)	Energy metabolism	EF1068	-0.84	-1.07	-0.72	0	-0.75
Iron-sulfur cluster binding protein	Energy metabolism	EF1109	0	0	-0.78	0	0
NADH peroxidase (<i>npr</i>)	Energy metabolism	EF1211	0	0	-0.59	0	-0.73
Glycosyl hydrolase, family 13	Energy metabolism	EF1347	0	0	0	-0.99	0.80
Glucan 1,6-alpha-glucosidase, putative	Energy metabolism	EF1348	0	0	0	-1.01	0.77
Glycosyl hydrolase, family 13	Energy metabolism	EF1349	0	0	0	-0.99	0.93
Glyceraldehyde 3-phosphate dehydrogenase (<i>gap-1</i>)	Energy metabolism	EF1526	0	0	0	0	-0.98
Pyruvate formate-lyase activating enzyme (<i>pflA</i>)	Energy metabolism	EF1612	0	0	0	-0.73	0
Formate acetyltransferase (<i>pflB</i>)	Energy metabolism	EF1613	0	0	0	-0.86	0
Alcohol dehydrogenase, zinc containing	Energy metabolism	EF1826	0	0	-0.84	-0.70	-1.01
Mannose-6-phosphate isomerase, class I (<i>manA</i>)	Energy metabolism	EF2589	0	0	0	-0.69	0
Glycerate kinase, putative	Energy metabolism	EF2646	0	0	0	-1.65	0.70
Endo-beta-N-acetylglucosaminidase	Energy metabolism	EF2863	0	0	0	-1.39	0
Ribokinase (<i>rhsK</i>)	Energy metabolism	EF2961	0	0	0	-1.32	-1.16
Glycosyl hydrolase, family 65	Energy metabolism	EF3157	0	0	0	-1.34	0
Lipase/acylhydrolase	Fatty acid and phospholipid metabolism	EF0169	0	0	0	0	-1.00
IS1216, transposase	Other categories	EF0514	0	0	-0.63	-0.81	-0.81
Transposase, putative	Other categories	EF0913	0	-0.69	0	0	0
Aminopeptidase C (<i>pepC</i>)	Protein fate	EF0302	0	0	0	0	-0.82
Rotamase family protein	Protein fate	EF0685	0	-0.60	-0.75	-0.80	-1.01
Preprotein translocase, YajC subunit, putative	Protein fate	EF0897	0	0	0	0	-0.83
Signal peptidase I	Protein fate	EF1111	0	-0.66	0	0	0
Serine proteinase, V8 family	Protein fate	EF1817	0	0	0	-0.86	0
Serine protease DO (<i>htrA</i>)	Protein fate	EF3027	0	0	0	-0.67	0
Peptidase, M20/M25/M40 family	Protein fate	EF3178	0	0	-0.62	-0.80	-1.32
CysteinyI-tRNA synthetase (<i>cysS</i>)	Protein synthesis	EF0045	0	0	-0.59	0	0
RNA methyltransferase, TrmH family	Protein synthesis	EF0047	0	-0.60	-0.61	0	0
Tyrosyl-tRNA synthetase (<i>tryS-1</i>)	Protein synthesis	EF0633	0	0	0	-0.67	-1.61
Glutamyl-tRNA (Gln) amidotransferase, C subunit (<i>gatC</i>)	Protein synthesis	EF0724	0	-0.65	-0.68	0	0
Ribosomal protein L25 (<i>rplY</i>)	Protein synthesis	EF0820	0	0	0.68	-1.01	-2.07
Alanyl-tRNA synthetase (<i>alaS</i>)	Protein synthesis	EF1379	0	0	0	0	-0.69
Ribosomal subunit interface protein (<i>yfiA</i>)	Protein synthesis	EF1764	0	-0.78	0	0	0
Aspartyl-tRNA synthetase (<i>aspS</i>)	Protein synthesis	EF1970	0	0	0	0	-0.77
Tryptophanyl-tRNA synthetase	Protein synthesis	EF2228	0	0	-0.62	0	0
Ribosomal protein L33 (<i>rpmG-3</i>)	Protein synthesis	EF2856	0	-0.98	0	0	0
Threonyl-tRNA synthetase (<i>thrS</i>)	Protein synthesis	EF2858	0	-0.72	-0.64	-0.90	-1.20
Valyl-tRNA synthetase (<i>valS</i>)	Protein synthesis	EF2931	0	0	0	-0.67	0
Seryl-tRNA synthetase (<i>serS-2</i>)	Protein synthesis	EF3292	0	0	0	-0.67	-0.83
Ribosomal protein L34 (<i>rpmH</i>)	Protein synthesis	EF3333	0	-0.61	0	0	0
Adenylosuccinate synthetase (<i>purA</i>)	Purine/pyrimidine/nucleoside/nucleotide	EF0014	0	-0.73	-0.73	0	-0.70
Phosphopentomutase (<i>deoB</i>)	Purine/pyrimidine/nucleoside/nucleotide	EF0185	0	0	-0.69	0	0
Ribonucleoside-diphosphate reductase 2, beta subunit (<i>nrdF</i>)	Purine/pyrimidine/nucleoside/nucleotide	EF0470	0	0	0	-0.80	0
Ribonucleoside-diphosphate reductase 2, alpha subunit (<i>nrdE</i>)	Purine/pyrimidine/nucleoside/nucleotide	EF0471	0	0	0	-0.69	-0.79
Ribonucleoside-diphosphate reductase 2, NrdH-redoxin (<i>nrdH</i>)	Purine/pyrimidine/nucleoside/nucleotide	EF0473	0	-0.59	0	-0.80	-0.83
Aspartate carbamoyltransferase (<i>pyrB</i>)	Purine/pyrimidine/nucleoside/nucleotide	EF1719	0	-0.63	0	0	0
Pyrimidine operon regulatory protein PyrR (<i>pyrR</i>)	Purine/pyrimidine/nucleoside/nucleotide	EF1721	0	-0.82	0	0	0
Inosine-uridine-preferring nucleoside hydrolase	Purine/pyrimidine/nucleoside/nucleotide	EF1921	0	0	0	-0.92	0
Anaerobic ribonucleoside-triphosphate reductase (<i>hrdD</i>)	Purine/pyrimidine/nucleoside/nucleotide	EF2754	0	0	0	-0.96	-0.83
Anaerobic ribonucleoside-triphosphate reductase activating protein (<i>nrdG</i>)	Purine/pyrimidine/nucleoside/nucleotide	EF2755	0	0	0	-0.96	-1.14
Transcriptional regulator, Cro/CI family	Regulatory functions	EF0129	0	-0.69	-0.73	0	0
Transcriptional regulator, Cro/CI family	Regulatory functions	EF0869	0	-0.64	0	0	0
Transcriptional regulator, DeoR family	Regulatory functions	EF1124	-0.79	-0.84	-0.68	0	-0.71
Transcriptional regulator, LysR family	Regulatory functions	EF1303	0	0	-0.87	0	0
Transcriptional regulator, Cro/CI family	Regulatory functions	EF2142	-0.67	0	-0.67	0	0
Transcriptional regulator, Cro/CI family	Regulatory functions	EF2338	0	-0.79	-0.60	0	-0.88
Transcriptional regulator, Cro/CI family	Regulatory functions	EF2544	0	-0.75	0	0	0
Transcriptional regulator	Regulatory functions	EF2703	0	-0.65	-0.84	0	0
Transcriptional regulator, AraC family	Regulatory functions	EF2711	0	0	0	-1.00	0
Transcriptional antiterminator, bglG family	Regulatory functions	EF2966	0	0	0	-1.08	-1.28
DNA-binding response regulator	Regulatory functions	EF3289	0	0	0	0	-0.84

Continued on following page

TABLE 5—Continued

Protein encoded by gene	Cellular role(s)	ORF	Log ₂ ratio at:				
			t0	t15	t30	t60	t90
PTS system, mannose-specific IIB component	Signal transduction	EF0020	0	0	0	-0.88	0
PTS system, mannose-specific IIC component	Signal transduction	EF0021	0	0	0	-1.22	1.04
PTS system, mannose-specific IID component	Signal transduction	EF0022	0	0	0	-1.34	1.04
PTS system, mannitol-specific IIBC component	Signal transduction	EF0411	0	0	0	0	-0.73
PTS system, IID component	Signal transduction	EF0553	0	0	0	-0.73	0
PTS system, IABC component	Signal transduction	EF0958	0	0	0	-1.27	0
PTS system, IIB component	Signal transduction	EF1012	0	0	0	-2.91	0
PTS system, IIB component	Signal transduction	EF1017	0	0	0	-2.75	0
PTS system, IIA component	Signal transduction	EF1018	0	0	0	-2.68	0
PTS system, IIC component	Signal transduction	EF1019	0	0	0	-3.28	0
Response regulator	Signal transduction	EF1336	0	-0.79	-0.60	0	0
Response regulator	Signal transduction	EF1633	-0.80	-1.06	-0.92	-0.80	-0.86
PTS system, IIA component	Signal transduction	EF1801	0	0	-0.69	0	0
PTS system, IIBC component	Signal transduction	EF2213	0	0	0	-1.83	0
PTS system, IID component	Signal transduction	EF3029	0	0	-0.66	0	0
Sensor histidine kinase	Signal transduction	EF3290	0	0	0	0	-0.97
RNA polymerase sigma-54 factor (<i>rpoN</i>)	Transcription	EF0782	0	-0.76	-0.64	0	0
RNA polymerase sigma-70 factor, ECF subfamily	Transcription	EF3180	0	0	-1.12	0	0
ABC transporter, ATP-binding protein	Transport and binding protein	EF0056	0	0	-0.75	0	0
ABC transporter, permease protein	Transport and binding protein	EF0179	0	-0.59	0	0	0
Amino acid ABC transporter, amino acid-binding/permease protein	Transport and binding protein	EF0247	0	0	-0.68	0	0
Na ⁺ /H ⁺ antiporter (<i>nhaC-1</i>)	Transport and binding protein	EF0402	0	0	0	-1.10	-0.98
Cation ABC transporter, permease protein	Transport and binding protein	EF0576	0	0	-0.62	0	0
ABC transporter, ATP-binding protein	Transport and binding protein	EF0793	-0.85	-0.92	-0.86	0	-0.75
Amino acid ABC transporter, ATP-binding protein	Transport and binding protein	EF0892	0	-0.61	0	0	0
Peptide ABC transporter, peptide-binding protein	Transport and binding protein	EF0907	0	0	0	-0.67	-1.39
ABC transporter, ATP-binding/TOBE domain protein	Transport and binding protein	EF0938	0	0	0	-1.39	0
Mn ²⁺ /Fe ²⁺ transporter, NRAMP family	Transport and binding protein	EF1057	0	-1.13	-1.12	-1.23	-1.78
ABC transporter, ATP-binding/permease protein	Transport and binding protein	EF1100	-0.76	-0.86	-0.70	0	0
Phosphotransferase enzyme II, B component SgaB (<i>sgaB</i>)	Transport and binding protein	EF1128	-0.86	-0.74	0	0	0
ABC transporter, permease protein	Transport and binding protein	EF1254	0	0	0	0	-0.88
Cation-transporting ATPase, E1-E2 family	Transport and binding protein	EF1268	0	-0.64	-0.72	0	0
Magnesium-translocating P-type ATPase	Transport and binding protein	EF1304	-0.84	-1.01	-1.31	0	0
ABC transporter, ATP-binding protein	Transport and binding protein	EF1331	0	-0.79	-0.69	0	0
Sugar ABC transporter, permease protein	Transport and binding protein	EF1344	0	0.62	0	-0.93	1.25
Sugar ABC transporter, sugar-binding protein	Transport and binding protein	EF1345	0	0	0	-1.39	0
Magnesium-translocating P-type ATPase	Transport and binding protein	EF1352	0	0	-0.62	0	0
V-type ATPase, subunit F	Transport and binding protein	EF1492	0	0	-0.86	0	0
ABC transporter, ATP-binding protein	Transport and binding protein	EF1673	-0.65	0	-0.82	0	0
Uracil permease	Transport and binding protein	EF1720	0	-0.60	0	0	0
Phosphate ABC transporter, phosphate-binding protein	Transport and binding protein	EF1759	0	0	-0.61	0	0
ABC transporter, permease protein, putative	Transport and binding protein	EF2049	0	0	-1.18	0	0
ABC transporter, ATP-binding protein	Transport and binding protein	EF2050	0	0	-1.16	0	0
ABC transporter, ATP-binding protein	Transport and binding protein	EF2074	0	0	0	-0.70	-0.76
ABC transporter, permease protein	Transport and binding protein	EF2081	0	0	0	-0.68	0
ABC transporter, ATP-binding protein	Transport and binding protein	EF2394	0	0	-0.61	0	0
Permease, GntP family	Transport and binding protein	EF2647	0	0	0	-2.31	1.32
ABC transporter, ATP-binding protein	Transport and binding protein	EF2769	0	0	-0.60	0	0.72
Ribose uptake protein, putative	Transport and binding protein	EF2959	0	0	0	-1.19	-0.77
Ribose transporter protein RbsD	Transport and binding protein	EF2960	0	0	-0.70	-1.48	-1.30
Peptide ABC transporter, peptide-binding protein	Transport and binding protein	EF3106	0	0	0	-1.88	-1.13
Peptide ABC transporter, permease protein	Transport and binding protein	EF3107	0	0	0	-1.75	-0.82
Peptide ABC transporter, permease protein	Transport and binding protein	EF3108	0	0	0	-2.19	-0.87
Peptide ABC transporter, ATP-binding protein	Transport and binding protein	EF3109	0	0	0	-1.33	0
Peptide ABC transporter, ATP-binding protein	Transport and binding protein	EF3110	0	0	0	-1.65	-0.86
Cytosine permease, putative	Transport and binding protein	EF3277	-0.80	-0.74	0	0	0
AMP-binding family protein	Unknown function	EF0452	0	0	-0.61	0	0
OsmC/Ohr family protein	Unknown function	EF0453	0	0.95	-1.05	-1.16	-1.67
Acetyltransferase, GNAT family	Unknown function	EF0678	0	0	0	-1.32	0
Oxidoreductase, aldo/keto reductase 2 family	Unknown function	EF0877	0	0	0	-0.67	0
Pentapeptide repeat family protein	Unknown function	EF0905	-0.81	-1.04	-0.92	0	-0.75
N-acyl-D-amino-acid deacylase family protein	Unknown function	EF1062	-0.92	-1.02	0	0	0
Acetyltransferase, GNAT family	Unknown function	EF1075	-0.87	-0.97	-0.79	0	-0.78
Acetyltransferase, GNAT family	Unknown function	EF1077	-0.64	-0.92	-0.64	0	0
YkgG family protein	Unknown function	EF1110	-0.67	-0.71	0	0	0
Oxidoreductase, aldo/keto reductase family	Unknown function	EF1138	0	0	0	0	-0.77
HesA/MoeB/ThiF family protein	Unknown function	EF1329	0	-0.58	-0.64	0	0
CelC-related protein	Unknown function	EF2440	0	0	0	-1.31	0
PDZ domain protein	Unknown function	EF2450	0	0	0	0	-0.68
HD domain protein	Unknown function	EF2470	0	0	0	0	-0.80

Continued on facing page

TABLE 5—Continued

Protein encoded by gene	Cellular role(s)	ORF	Log ₂ ratio at:				
			t0	t15	t30	t60	t90
Aldehyde oxidoreductase, putative	Unknown function	EF2570	0	0	-0.80	0	0
Hydrolase, haloacid dehalogenase-like family	Unknown function	EF2681	0	-0.86	0	0	0
Oxidoreductase, Gfo/Idh/MocA family	Unknown function	EF2734	0	0	0	0	-0.76
Thioredoxin reductase/glutathione-related protein	Unknown function	EF2738	0	0	-0.62	0	0
DNA-binding protein, putative	Unknown function	EF2933	0	0	-0.69	0	0
Glyoxalase family protein	Unknown function	EF3092	0	0	0	-0.71	0
Hydrolase, haloacid dehalogenase-like family	Unknown function	EF3158	0	0	0	-1.16	-0.72
Cell-envelope-associated acid phosphatase	Unknown function	EF3245	0	0	-0.78	0	0
Gp35	Viral functions	EF2143	0	-0.59	-0.59	0	0

^aGene names in parentheses.

many cellular activities when important processes, such as protein synthesis, apparently are repressed. In our experiments, we found that a lower number of genes was induced than repressed at all time points except for t90 (see Tables 1 to 5). At t90, it is tempting to look upon the altered transcriptional activity in the stressed cells as a consequence of an adaptation to the presence of erythromycin. At this point, the cells may have adapted to the stressing conditions and may have overcome the critical effects of the drug.

Ng et al. (19) report the induction of several genes related to purine biosynthesis in *Streptococcus pneumoniae* R6 exposed to various translation inhibitors, including erythromycin. This effect on purine synthesis is not clear for V583; only *purK* is up-regulated, and *purA* is down-regulated. Three genes related to pyrimidine biosynthesis (*pyrD-1*, *pyrD-2*, and *pyrC*) are induced, however. Similarly to what was found with *S. pneumoniae* R6 (19), the purine salvage gene *xpt* had a relative increase in transcription in V583 in response to the erythromycin treatment. Therefore, it looks as if the responses of the two relatively closely related bacteria *E. faecalis* and *S. pneumoniae* to erythromycin have common themes, but most of the transcriptional responses appear distinct.

Transport and binding proteins represent the second most dominant group of differentially expressed genes (Tables 1 to 5). This group of genes appears to be heavily influenced by various stressors in many bacteria (see, e.g., references 19, 21, and 26). The altered transcription of transport and binding proteins indicates that altered transport is an important part of general stress response mechanisms. The large number of differentially expressed genes encoding ABC transporters and permeases emphasizes the importance of such genes in stress responses.

Another interesting group of induced genes are those related to fatty acid and phospholipid metabolism (six genes). As mentioned briefly above, erythromycin resistance in gram-negative bacteria is usually mediated by a low permeability of the outer membrane to the hydrophobic macrolide. One may speculate as to whether a slightly altered cell membrane may have contributed also to the erythromycin adaptation in the gram-positive V583 cells.

In time course experiments like ours, one should draw attention to genes that are differentially expressed at all time points examined. Our results showed that 14 genes were regulated, 4 up and 10 down, at all five time points. Among these, the most interesting gene is EF1413, which encodes a putative MsrC protein, although the sequence identity with the de-

scribed MsrC is not more than ca. 40%. EF1413 was up-regulated at all time points and showed the strongest differential expression of all genes on the microarray (Table 4). EF1413 was not differentially expressed in experiments with chloramphenicol treatment of V583 (Å. Aakra, unpublished results). The *msrC* is distributed in many isolates of *E. faecium* and has even been suggested to be specific for this species (23, 25). Comparative genomic hybridizations with five *E. faecalis* strains indicate the presence of the putative *msrC* in these genomes, as well (Aakra, unpublished). Singh et al. (25) found that the *msrC* conferred an increased resistance to macrolides among *E. faecium* isolates. Our observation of strong induction of EF1413 (log₂ ratio of 1.88 at t0; log₂ ratio of 3.44 at t90) in erythromycin exposed *E. faecalis* V583 support the involvement of an MsrC homologue in macrolide resistance in this species as well.

EF1732 and EF1733 encode two ABC transporters belonging to the MDR family. These two genes were up-regulated at all time points except t0. (In a similar study, the same two genes were also strongly up-regulated in response to chloramphenicol treatment [Aakra, unpublished].) Taken together, these results make it tempting to speculate that the efflux of erythromycin by proteins encoded by EF1413, EF1732, and EF1733 is an important part of the survival mechanism for V583 when exposed to this antibiotic.

At t0, i.e., immediately after the addition of erythromycin, 18 genes were up-regulated and 56 genes were down-regulated, which were considerably fewer genes than those regulated at the other time points under study. It must be assumed that the genes that show an immediate response also are crucial for the growth and survival of erythromycin-exposed V583 cells.

Several studies on stress responses (and genes involved therein) of *E. faecalis* have been published (e.g., see references 1, 8, 9, 14, and 15). Some papers focus on two-component signal transduction pathways, which commonly are related to bacterial stress (10, 15). A few of the genes found to have a significantly differential expression in our work have been discussed in these papers, e.g., genes encoding the Gls24 proteins (EF0079 and EF0080), two histidine kinases and their cognate response regulators (EF3290 and EF3289, EF1051 and EF1050), L-lactate dehydrogenase (EF0641), L-serine dehydratase (EF0098, EF0099), carbamate kinase (EF0106), and superoxide dismutase (EF0463). In this work, we found that the *gls24* genes (EF0079, EF0080) were repressed at all time points except t0, which is in contrast with the hypotheses of Giard et al. (9) that these genes are induced under stress. The

repression of the *gls24* genes was also found in our study of the transcriptional responses of V583 to chloramphenicol treatment (Aakra, unpublished). Similarly to Giard et al. (9), the EF0604 which encodes another Gls24 protein was not found to be differentially expressed at any time point of our study. Therefore, the role of Gls24 in stress responses in *E. faecalis* is not obvious. Regarding the two-component signal transduction pathways, the histidine kinase (HK) EF3290 and the cognate response regulator (RR) EF3289 were both down-regulated at *t*90. The function of this HK-RR pair is not known (10), and it remains to be seen whether the repression of EF3290/EF3289 at *t*90 in this study is stress related. The other pair of differentially expressed HK-RR (EF1051/EF1050) is similar to known HK-RR systems in *Listeria monocytogenes* and *S. pneumoniae* (10). EF1051 and EF1050 were both induced at *t*30. Recently, the EF1051/EF1050 pair was described by Teng et al. (27), who named this pair of genes *etaRS* (enterococcal two-component system a). Teng et al. (27) showed the involvement of *EtaRS* in both stress responses and virulence, and our results support their hypothesis on the function of *etaRS*. EF0641 encoding L-lactate dehydrogenase (*ldh*) was down-regulated at *t*60 and *t*90. Previously, it was shown that *ldh* was induced under stress (8), and it was speculated that also this gene could be involved in stress responses (8), but the function is unclear. The repression of *ldh* in this study indicates that the possible involvement of this gene in stress is not general. Likewise, *sodA* (encoding superoxide dismutase), *arcC* (encoding carbamate kinase), and *sdhA* and *sdhB* (encoding L-serine dehydratase) were down-regulated in our work, while they were induced in the study by Giard et al. (8). Based on these comparisons, it appears that a considerable number of the genes affected by environmental stress in *E. faecalis* are not general stress genes but, rather, are specified by certain conditions.

Interestingly, EF1078, which was induced at all time points except *t*0, is identical to *emeA* (enterococcal multidrug resistance efflux), which was characterized by Jonas et al. (12). *emeA* is probably a homolog of the *norA* found in *Staphylococcus aureus* (12). Jonas et al. (12), showed that *emeA* is involved in resistance to many toxic compounds, among them erythromycin (12). Thus, our study also supports the results of Jonas et al. on the function of *emeA*.

We have presented the transcriptional profile of erythromycin-exposed *E. faecalis* V583. The addition of erythromycin to this bacterium causes numerous events of differential transcription. Efflux of the macrolide molecules from the cells may be an important part of the survival mechanism, in addition to the resistance conferred by *ErmB*. This work adds information to the growing archive of condition-specific bacterial transcription signatures, which, in the future, will aid the elucidation of microbial physiology, metabolism, ecology, pathogenesis, etc. With a comprehensive archive of such transcription signatures, it will also be possible to pay more attention to all genes encoding hypothetical proteins or putative genes. This group represents a significant part of prokaryotic genomes and a dominant group of regulated genes in many microarray studies published so far. Profiling of transcriptional events in V583 under other stress conditions is in progress. Hopefully, this will lead to a deeper understanding of the biology of this bacterium.

ACKNOWLEDGMENTS

This work was financially supported by grant 142656/140 from the Research Council of Norway to I.F.N.

We thank Linda Godager and Merete Lunde for assistance with the RTQ analyses. Bjørn E. Kristiansen, of the Norwegian Microarray Consortium, Oslo, is acknowledged for printing of the microarray slides.

REFERENCES

- Breton, Y. L., A. Maze, A. Hartke, S. Lemariner, Y. Auffray, and A. Rince. 2002. Isolation and characterization of bile salts-sensitive mutants of *Enterococcus faecalis*. *Curr. Microbiol.* **45**:434–439.
- Conway, T., and G. K. Schoolnik. 2003. Microarray expression profiling: capturing a genome-wide portrait of the transcriptome. *Mol. Microbiol.* **47**:879–889.
- Cummings, C. A., M. M. Brinig, P. W. Lepp, S. van de Pas, and D. A. Relman. 2004. *Bordetella* species are distinguished by patterns of substantial gene loss and host adaptation. *J. Bacteriol.* **186**:1484–1492.
- Diggle, P. J., K. Liang, and S. L. Zeger. 1994. Analysis of longitudinal data. Clarendon Press, Oxford, United Kingdom.
- Evers, S., K. Di Padova, M. Meyer, H. Langen, M. Fountoulakis, W. Keck, and C. P. Gray. 2001. Mechanism-related changes in the gene transcription and protein synthesis patterns of *Haemophilus influenzae* after treatment with transcriptional and translational inhibitors. *Proteomics* **1**:522–544.
- Facklam, R. R., M. G. S. Carvalho, and L. M. Teixeira. 2002. History, taxonomy, biochemical characteristics and antibiotic susceptibility testing of enterococci, p. 1–54. In M. S. Gilmore, D. B. Clewell, P. Courvalin, G. M. Dunny, B. E. Murray, and L. B. Rice (ed.), *The enterococci: pathogenesis, molecular biology, and antibiotic resistance*. ASM Press, Washington, D.C.
- Franz, C. M., W. H. Holzappel, and M. E. Stiles. 1999. Enterococci at the crossroads of food safety? *Int. J. Food Microbiol.* **47**:1–24.
- Giard, J. C., J. M. Laplace, A. Rince, V. Pichereau, A. Benachour, C. Lebouef, S. Flahaut, Y. Auffray, and A. Hartke. 2001. The stress proteome of *Enterococcus faecalis*. *Electrophoresis* **22**:2947–2954.
- Giard, J. C., N. Verneuil, Y. Auffray, and A. Hartke. 2002. Characterization of genes homologous to the general stress-inducible gene *gls24* in *Enterococcus faecalis* and *Lactococcus lactis*. *FEMS Microbiol. Lett.* **206**:235–239.
- Hancock, L., and M. Perego. 2002. Two-component signal transduction in *Enterococcus faecalis*. *J. Bacteriol.* **184**:5819–5825.
- Hutter, B., C. Schaab, S. Albrecht, M. Borgmann, N. A. Brunner, C. Freiberg, K. Ziegelbauer, C. O. Rock, I. Ivanov, and H. Loferer. 2004. Prediction of mechanisms of action of antibacterial compounds by gene expression profiling. *Antimicrob. Agents Chemother.* **48**:2838–2844.
- Jonas, B. M., B. E. Murray, and G. M. Weinstock. 2001. Characterization of *emeA*, a *norA* homolog and multidrug resistance efflux pump, in *Enterococcus faecalis*. *Antimicrob. Agents Chemother.* **45**:3574–3579.
- Koide, T., P. A. Zaini, L. M. Moreira, R. Z. Vencio, A. Y. Matsukuma, A. M. Durham, D. C. Teixeira, H. El-Dorry, P. B. Monteiro, A. C. da Silva, S. Verjovski-Almeida, A. M. da Silva, and S. L. Gomes. 2004. DNA microarray-based genome comparison of a pathogenic and a nonpathogenic strain of *Xylella fastidiosa* delineates genes important for bacterial virulence. *J. Bacteriol.* **186**:5442–5449.
- Laport, M. S., J. A. C. Lemos, M. D. C. F. Bastos, R. A. Burne, and M. Giambiagi-de Marval. 2004. Transcriptional analysis of the *groE* and *dnaK* heat-shock operons of *Enterococcus faecalis*. *Res. Microbiol.* **155**:252–258.
- Le Breton, Y., G. Boel, A. Benachour, H. Prevost, Y. Auffray, and A. Rince. 2003. Molecular characterization of *Enterococcus faecalis* two-component signal transduction pathways related to environmental stresses. *Environ. Microbiol.* **5**:329–337.
- Murray, B. E. 1990. The life and times of the enterococcus. *Clin. Microbiol. Rev.* **3**:46–65.
- Murray, B. E. 2000. Vancomycin-resistant enterococcal infections. *N. Engl. J. Med.* **342**:710–721.
- Murray, B. E. 1997. Vancomycin-resistant enterococci. *Am. J. Med.* **102**:284–293.
- Ng, W. L., K. M. Kazmierczak, G. T. Robertson, R. Gilmour, and M. E. Winkler. 2003. Transcriptional regulation and signature patterns revealed by microarray analyses of *Streptococcus pneumoniae* R6 challenged with sublethal concentrations of translation inhibitors. *J. Bacteriol.* **185**:359–370.
- Paulsen, I. T., L. Banerjee, G. S. Myers, K. E. Nelson, R. Seshadri, T. D. Read, D. E. Fouts, J. A. Eisen, S. R. Gill, J. F. Heidelberg, H. Tettelin, R. J. Dodson, L. Umayam, L. Brinkac, M. Beanan, S. Daugherty, R. T. DeBoy, S. Durkin, J. Kolonay, R. Madupu, W. Nelson, J. Vamathevan, B. Tran, J. Upton, T. Hansen, J. Shetty, H. Khouri, T. Utterback, D. Radune, K. A. Ketchum, B. A. Dougherty, and C. M. Fraser. 2003. Role of mobile DNA in the evolution of vancomycin-resistant *Enterococcus faecalis*. *Science* **299**:2071–2074.
- Paustian, M. L., B. J. May, and V. Kapur. 2001. *Pasteurella multocida* gene expression in response to iron limitation. *Infect. Immun.* **69**:4109–4115.

22. **Pechere, J. C.** 2001. Macrolide resistance mechanisms in Gram-positive cocci. *Int. J. Antimicrob. Agents* **18**(Suppl. 1):S25–S28.
23. **Portillo, A., F. Ruiz-Larrea, M. Zarazaga, A. Alonso, J. L. Martinez, and C. Torres.** 2000. Macrolide resistance genes in *Enterococcus* spp. *Antimicrob. Agents Chemother.* **44**:967–971.
24. **Sahm, D. F., J. Kissinger, M. S. Gilmore, P. R. Murray, R. Mulder, J. Solliday, and B. Clarke.** 1989. In vitro susceptibility studies of vancomycin-resistant *Enterococcus faecalis*. *Antimicrob. Agents Chemother.* **33**:1588–1591.
25. **Singh, K. V., K. Malathum, and B. E. Murray.** 2001. Disruption of an *Enterococcus faecium* species-specific gene, a homologue of acquired macrolide resistance genes of staphylococci, is associated with an increase in macrolide susceptibility. *Antimicrob. Agents Chemother.* **45**:263–266.
26. **Smoot, L. M., J. C. Smoot, M. R. Graham, G. A. Somerville, D. E. Sturdevant, C. A. Migliaccio, G. L. Sylva, and J. M. Musser.** 2001. Global differential gene expression in response to growth temperature alteration in group A *Streptococcus*. *Proc. Natl. Acad. Sci. USA* **98**:10416–10421.
27. **Teng, F., L. Wang, K. V. Singh, B. E. Murray, and G. M. Weinstock.** 2002. Involvement of PhoP-PhoS homologs in *Enterococcus faecalis* virulence. *Infect. Immun.* **70**:1991–1996.
28. **VanBogelen, R. A., and F. C. Neidhardt.** 1990. Ribosomes as sensors of heat and cold shock in *Escherichia coli*. *Proc. Natl. Acad. Sci. USA* **87**:5589–5593.
29. **Weisblum, B.** 1995. Erythromycin resistance by ribosome modification. *Antimicrob. Agents Chemother.* **39**:577–585.
30. **Wernisch, L., S. L. Kendall, S. Soneji, A. Wietzorrek, T. Parish, J. Hinds, P. D. Butcher, and N. G. Stoker.** 2003. Analysis of whole-genome microarray replicates using mixed models. *Bioinformatics* **19**:53–61.
31. **Wolfinger, R. D., G. Gibson, E. D. Wolfinger, L. Bennett, H. Hamadeh, P. Bushel, C. Afshari, and R. S. Paules.** 2001. Assessing gene significance from cDNA microarray expression data via mixed models. *J. Comput. Biol.* **8**:625–637.

PAPER II

The Transcriptome of the Nosocomial Pathogen *Enterococcus faecalis* V583 Reveals Adaptive Responses to Growth in Blood

Heidi C. Vebø^{1*}, Lars Snipen², Ingolf F. Nes¹, Dag A. Brede¹

1 Laboratory of Microbial Gene Technology and Food Microbiology, Biotechnology and Food Science, The Norwegian University of Life Sciences, Ås, Norway, **2** Section for Biostatistics, Department of Chemistry, Biotechnology and Food Science, The Norwegian University of Life Sciences, Ås, Norway

Abstract

Background: *Enterococcus faecalis* plays a dual role in human ecology, predominantly existing as a commensal in the alimentary canal, but also as an opportunistic pathogen that frequently causes nosocomial infections like bacteremia. A number of virulence factors that contribute to the pathogenic potential of *E. faecalis* have been established. However, the process in which *E. faecalis* gains access to the bloodstream and establishes a persistent infection is not well understood.

Methodology/Principal Findings: To enhance our understanding of how this commensal bacterium adapts during a bloodstream infection and to examine the interplay between genes we designed an *in vitro* experiment using genome-wide microarrays to investigate what effects the presence of and growth in blood have on the transcriptome of *E. faecalis* strain V583. We showed that growth in both 2xYT supplemented with 10% blood and in 100% blood had a great impact on the transcription of many genes in the V583 genome. We identified several immediate changes signifying cellular processes that might contribute to adaptation and growth in blood. These include modulation of membrane fatty acid composition, oxidative and lytic stress protection, acquisition of new available substrates, transport functions including heme/iron transporters and genes associated with virulence in *E. faecalis*.

Conclusions/Significance: The results presented here reveal that cultivation of *E. faecalis* in blood *in vitro* has a profound impact on its transcriptome, which includes a number of virulence traits. Observed regulation of genes and pathways revealed new insight into physiological features and metabolic capacities which enable *E. faecalis* to adapt and grow in blood. A number of the regulated genes might potentially be useful candidates for development of new therapeutic approaches for treatment of *E. faecalis* infections.

Citation: Vebø HC, Snipen L, Nes IF, Brede DA (2009) The Transcriptome of the Nosocomial Pathogen *Enterococcus faecalis* V583 Reveals Adaptive Responses to Growth in Blood. PLoS ONE 4(11): e7660. doi:10.1371/journal.pone.0007660

Editor: Stefan Bereswill, Charité-Universitätsmedizin Berlin, Germany

Received: October 2, 2009; **Accepted:** October 10, 2009; **Published:** November 4, 2009

Copyright: © 2009 Vebø et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work was financially supported by grant 142656 from The Research Council of Norway. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: heidi.vebo@umb.no

Introduction

Enterococcus faecalis is a common resident of the gastrointestinal tract of humans [1]. This bacterium displays a rough physiology that enables it to withstand oxidative stress [2] and harsh conditions such as high pH and salt concentrations [3]. *E. faecalis* is also an opportunistic pathogen, ranked among the leading causes of nosocomial infections worldwide [4]. Enterococci constitute the third most prevalent pathogens isolated from bloodstream infections, and represent the most frequent cause of surgical-site infections in intensive care units [4]. In the United States, *E. faecalis* accounts for approximately 80% of all enterococcal nosocomial infections [5].

E. faecalis V583 (referred to as V583 hereafter) originates from a patient suffering from a persistent bloodstream infection, and it was the first vancomycin-resistant clinical isolate reported in the United States [6]. V583 is part of the high risk clonal complex 2 [7,8], which comprises mostly of isolates derived from hospital

infections world wide. The genome of V583 contains several virulence related genes [8], including several antigens such as *E. faecalis* antigen A (EfaA) [9], and two well characterized antigenic exopolysaccharides; the serotype 2 capsular polysaccharide (*cps*) [10,11], and the enterococcal polysaccharide antigen biosynthesis cluster (*epa*) [12,13]. It has been acknowledged that *E. faecalis* acquires genetic traits by horizontal gene transfer, which includes virulence and antibiotic resistance determinants, to survive and persist in complex environments such as different infection sites (reviewed in [14]). However, *E. faecalis* pathogenesis most likely involves an orchestrated interplay between the regulation of virulence factors and multiple genetic traits that govern adaptation of the bacterial cell physiology during the process of infection. Several functional studies have been performed to link genetic traits to virulence [10,15–21], but few studies have examined such genome wide transcriptional interplay in *E. faecalis*.

Even though *E. faecalis* is a clinically significant pathogen implicated in different types of infections, little is known regarding

the molecular mechanisms involved in the adaptive process this bacterium undertakes to permit survival and growth in e.g. the bloodstream of an infected patient. Several studies have demonstrated that *E. faecalis* has evolved opportunistic strategies to sense and respond to entrance into the bloodstream of a host [22–26]. To improve the current understanding of *E. faecalis*' ability to cause bloodstream infections, we performed a genome wide transcriptional analysis of V583 during growth in 2xYT supplemented with 10% blood (YTB). We have employed a biphasic approach, performing a time-course experiment to examine the immediate responses of *E. faecalis* to blood as a biological cue, but also to explore the adaptation of *E. faecalis* in the presence of blood. Secondly, to increase our knowledge regarding the initial phase of an *E. faecalis* bloodstream infection, growth capacity and transcriptome responses in 100% blood were assessed. These experiments revealed that both growth in the presence of a small percentage (10%) of blood, and pure blood alters the transcription of the bacterium extensively. The results presented here provide new insights into processes essential for the survival and growth of *E. faecalis* in the complex blood environment.

Results

Growth of V583 in Blood and YTB Compared to 2xYT-Culture Medium

E. faecalis is able to establish a persistent infection in the bloodstream and internal organs of an infected host [27,28], and it is of utmost importance to understand the mechanisms that enable *E. faecalis* to survive in this complex growth environment. A rich laboratory medium (2xYT) was selected as the reference culture medium since it is considered to contain minuscule amounts of

infection relevant biological cues [24]. Initial experiments were performed to assess growth and behavior of V583 in 2xYT, in 2xYT supplemented with different concentrations of blood, and in pure blood. Based on these experiments it was decided to use 10% blood in 2xYT (YTB) and 100% blood in the subsequent transcriptome profiling experiments. Since *E. faecalis* is known to sense and respond to target cells such as erythrocytes, e.g. by expressing virulence factors like the toxin cytolysin [23], we decided to use whole blood rather than serum or plasma to mimic the *in vivo* environment and to examine other responses possibly modulated by erythrocytes.

The morphology of V583 was examined by light microscopy, revealing that it grew in chains consisting of up to 8 cells in 2xYT. It was also evident that in the presence of blood, bacterial cells aggregated, probably due to agglutination. In order to obtain reliable colony forming units (CFU) counts, gentle sonication was applied to break up aggregates and long chains of V583 cells. Several tests were performed to ensure that the aggregates and chains were properly dissolved without affecting the viability of the cells. V583 was cultivated in 2xYT until mid-exponential growth phase (cell density $\sim 1 \times 10^8$ CFU/ml), prior to exposing the cells to the test conditions; pre-warmed 2xYT, YTB or blood, as described in the Materials and Methods section. The average growth curves of V583 grown in 2xYT, YTB and blood measured by CFU counts are presented in Fig. 1.

The doubling time (Td) of V583 was similar for growth in 2xYT and YTB, with a Td of 39.7 and 36.8 minutes respectively. However, growth of V583 in blood was constrained compared to in 2xYT with a Td of 80.5 minutes. When grown in 2xYT or YTB, V583 reached a maximum cell density of 1×10^9 CFU/ml, whereas in blood V583 reached approximately 2×10^8 CFU/ml.

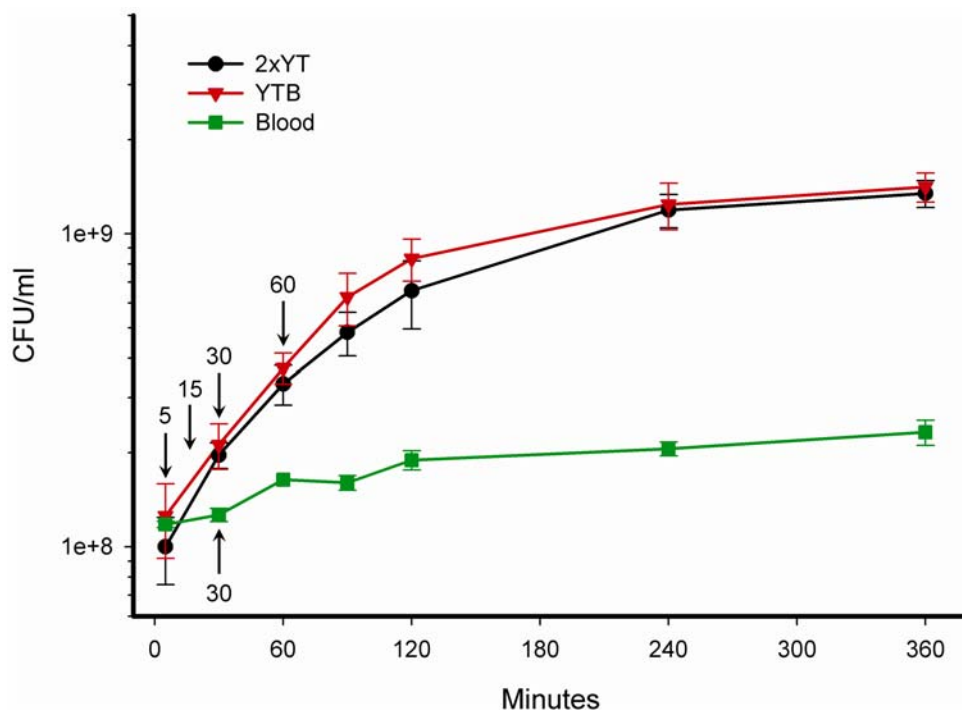


Figure 1. Characterization of *E. faecalis* V583 growth in 2xYT, YTB and blood. Growth of *E. faecalis* V583 was determined from cultures pre-cultivated in 2xYT, and transferred to a fresh medium (either 2xYT, YTB or blood). The growth curves are represented by colony forming units per millilitre (CFU/ml) on the Y-axis, and minutes as indicated on the X-axis. The growth curves correspond to the mean \pm STD of three individual experiments. Arrows indicate the time points when samples were harvested prior to RNA extraction. Samples from 2xYT were harvested either only after 30 minutes (when compared to blood), or after 5, 15, 30 and 60 minutes (when compared to YTB). doi:10.1371/journal.pone.0007660.g001

Based on these results, it was decided to investigate transcriptional responses after 5, 15, 30 and 60 minutes growth in YTB, and after 30 minutes growth in blood as indicated in Fig. 1.

Global Adaptation of the V583 Transcriptome Reveals Changes Comprising Most Functional Gene Categories

To examine *E. faecalis*' immediate response to blood as a biological cue and adaptation to a prolonged existence in blood, a time-course experiment was carried out. The time points for cell harvesting followed by RNA isolation (within 60 minutes after addition of blood), were carefully chosen to reflect the actively growing V583 cells in both 2xYT and in YTB (Fig. 1), as well as to portray the different stages of adaptation that V583 undergo upon the first encounter of blood. To further examine the *E. faecalis* adaptation towards persistence in blood, the transcriptional response of V583 grown for 30 minutes in blood was assessed. The obtained \log_2 -ratios and p-values for all the V583 genes found during exposure to YTB and blood compared to 2xYT are listed in Table S1. Statistical analysis using a mixed model [29] combined with a stringent Bonferroni corrected confidence level of $p < 0.05$, identified 148 significantly regulated genes during growth of *E. faecalis* in YTB. Of these, 72 genes were up-regulated, 73 where down-regulated and 3 genes where both up- and down-

regulated at one or more time points. The most pronounced transcriptional responses to YTB occurred after 15 and 30 minutes. When V583 was grown in blood for 30 minutes, a total of 549 genes were differentially expressed (225 genes were up-regulated and 324 were down-regulated). The heat map in Fig. 2 presents an overview of the regulated genes within each functional category in the YTB and blood experiments. This revealed similarities in the expression patterns of a number of genes between the YTB and the blood experiments, e.g. genes involved in fatty acid and phospholipid metabolism, energy metabolism, transcription, genes with a regulatory function and genes encoding proteins with binding and/or transport functions (Fig. 2). Growth in pure blood caused additional transcriptional responses within several gene categories such as the cell envelope, protein synthesis and amino acid biosynthesis, which can be seen as a distinct pattern in the heat map (Fig. 2). The down-regulation of several genes within the latter two functional groups most likely reflects the reduced growth rate in blood compared to in 2xYT (Fig. 1).

Stress Response of V583 Caused by Blood Exposure

Several of the V583 genes associated with stress protection mechanisms in enterococci were found to respond to blood, while only a few stress-related genes were regulated in response to

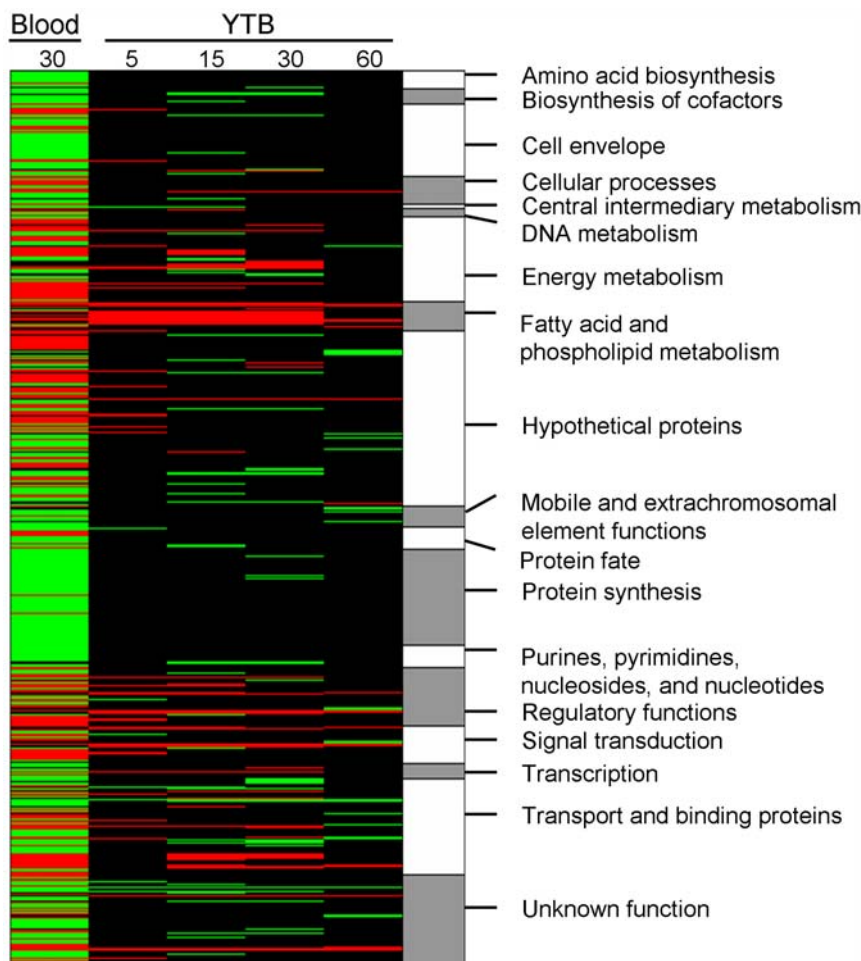


Figure 2. Heat map visualizing regulated genes in V583 grown in blood and YTB compared to growth in 2xYT. Genes found to be significantly regulated are indicated by either red (up-regulated), or green (down-regulated). Genes regulated after growth for 30 minutes in blood, compared to in 2xYT are listed in column 1. Genes regulated during growth in YTB compared to 2xYT are listed in columns 2–4, time-points in minutes indicated on the top of each column. The genes are sorted alphabetically by functional categories (column 5). doi:10.1371/journal.pone.0007660.g002

growth in YTB as expected. For growth in blood an operon (EF0076-81) which includes the *gls24* and *glsB* genes [30,31], was up-regulated (Table 1 and S1). The Gls24 protein appears to be implicated in both stress protection and virulence of *E. faecalis* [30,32]. Exposure to blood also mounted universal stress protection genes, including EF1084 and the general stress operon *gspA1-2* (EF1810-11). A gene encoding a cold shock protein *cspC* (EF1991) was found to be up-regulated in both blood and in YTB after 15, 30 and 60 minutes, which indicates that this gene might be important for V583 to overcome the stress triggered by growth in blood. Interestingly, in response to growth in blood we also observed an up-regulation of EF1560, a hypothetical gene that has been reported to show an enhanced transcription under six different stress conditions in previous studies [33,34].

During the course of infection, bacteria are exposed to massive oxidative stress [35,36]. The microarray results revealed that the expression of several genes associated with oxidative stress response in *E. faecalis* was affected by blood exposure. Interestingly, previous studies have established a relationship between the oxidative stress response and virulence in *E. faecalis* [37–40]. A total number of nine oxidative stress protection genes [41] were regulated during growth in YTB or blood (Table 1 and S1). Five of these genes were up-regulated in response to growth in blood, including an organic hydroperoxide resistance gene *ohr* (EF0453) [42], the *dps* gene (EF0606) predicted to protect DNA from oxidative damage [8], the NADH peroxidase *npr* (EF1211), the peptide methionine reductase *msrB* (EF3164), and the superoxide dismutase gene *sodA* (EF0463). The *sodA* gene is important for *E. faecalis* to survive ingestion by macrophages [38] and was also up-regulated in *Streptococcus agalactiae* during growth in blood [43]. Four genes related to oxidative stress were found to be down-regulated in response to growth in blood (Table 1 and S1). Two of these genes, an oxidoreductase (EF3257) and a thioredoxin reductase *ahpF* (EF2738), were down-regulated both during growth in blood and YTB. An NADH oxidase *nox* (EF1586) was also found to be down-regulated during growth in blood, and an alkyl hydroperoxide reductase *ahpC* (EF2739) showed a reduced expression after 15 minutes growth in YTB.

Exposure to Blood Induces Modifications to the Cell Envelope

The integrity and composition of the cell envelope of an invading bacterium is important to evade the challenges evoked by the host defense systems [44,45]. A number of changes in the transcriptome of *E. faecalis* imply extensive adaptations in the cell membrane composition and surface related structures (Table S1). A particularly pronounced change was detected in two gene clusters (EF0282-84 and EF2886-75), responsible for type II fatty acid biosynthesis (FASII) and isomerization of membrane phospholipids. These loci were up-regulated throughout the time-course of the YTB experiment and also in response to growth in blood. Interestingly, these gene clusters have previously been shown to be regulated in response to exposure to the cell membrane detergents SDS and bovine bile [46], indicating that remodeling of the fatty acid composition in the cell membrane might be an important response to a broad range of external stressors. Furthermore, the up-regulation of the cardiolipin synthase gene EF1608 (Table 1 and S1), which modulates the phosphatidylglycerol content in the cell membrane, also suggests that exposure to blood affects the membrane composition of *E. faecalis*. Evidence of lipolytic activity in response to growth in blood was substantiated by the up-regulation of two lipases (EF0169 and EF3191) (Table 1 and S1). The latter gene did also show an

enhanced expression after 60 minutes growth in YTB and it has been proposed by Paulsen *et al* [8] as a potential virulence factor.

The *lrgAB* operon (EF3194-93) that encodes a putative lysis inhibitory system was highly up-regulated in response to blood (Table S1) with a log₂-value of 7.9 for *lrgB* and 5.6 for *lrgA*. The two genes were also found to be highly up-regulated throughout the time-course experiment in YTB. The rapid induction and consistently high level of transcription of the *lrgAB* operon, in addition to its putative function suggests a role in modification of the cell wall structure, which might be propitious for growth in blood. Noticeably, the synonymous genes (*gbs0182-83*) in *S. agalactiae* showed an increased expression during growth in blood [43]. The expression of *lrgAB* in *Staphylococcus aureus* is regulated by the closely located LytSR two-component system [47]. In V583, a two-component system (EF3197-96) homologous to the LytSR system in *S. aureus*, resides directly upstream from *lrgAB*. It is possible that this system is involved in the regulation of *lrgAB*, but only a modest enhanced expression (not statistically significant) of this two-component system was observed.

E. faecalis contains two gene clusters responsible for the production of two serotype-determining exopolysaccharides: the serotype 2 capsular polysaccharide (*cps*) [10], and the enterococcal polysaccharide antigen biosynthesis cluster (*epa*) [13]. It has previously been shown that the *cps* and *epa* clusters affect virulence in mice [48–50] and also contribute to resistance against phagocytic killing [10,20,51]. In our experiments most of the genes within the *epa* cluster (EF2200-2189 and EF2184-77) and the *cps* cluster (EF2492-84) were down-regulated during growth in blood (Table 1 and S1). In addition, some of the *cps* genes were also down-regulated after 15 and 30 minutes growth in YTB. These results are consistent with previous work on *E. faecalis* FA2-2, which showed that genes in the *cps* locus were down-regulated during growth in serum [11].

The V583 genome contains an operon, *dltABCD* (EF2749-46), responsible for incorporating D-alanine into cell-wall associated teichoic acids and lipoteichoic acids [52]. The first two genes of this operon, *dltA* and *dltB*, were found to be down-regulated in response to growth in blood (Table 1 and S1). Reduced content of D-alanine esters in the teichoic acid results in an increased net negative charge on the bacterial cell surface, which in turn can affect several bacterial properties such as susceptibility to cationic antimicrobial peptides and biofilm formation [52]. In addition, the expression of a genetic locus known to be involved in biofilm formation and maltose metabolism [53,54] showed an enhanced expression in response to growth in blood (Table 1 and S1). This locus includes one operon, the *bopABCD/malPBMR* operon (EF0957-54), and a phosphoenolpyruvate phosphotransferase system (PTS) *malT* (EF0958) of which the genes *malT* and *bopAB/malPB* were up-regulated in blood. The genes *bopCD/malMR* showed the same trend, although not statistically significant. Furthermore, the secreted antigen *salB* (EF0394) showed reduced expression during growth in blood, and after 5 minutes growth in YTB (Table 1 and S1). This gene has also been demonstrated to be important for biofilm formation [55].

It has been proposed that lipoproteins are implicated in virulence in *E. faecalis* [56]. Interestingly, we found that the transcription of nine genes encoding lipoproteins were regulated (five up-regulated and four down-regulated). Among these, the cAD1 conjugation pheromone precursor (EF3256) showed decreased expression in blood and after 15 minutes in YTB (Table 1 and S1), while transcription of an operon encoding an ABC-transporter and an YaeC family lipoprotein (EF3200-EF3198) was elevated in blood and after 15 and 30 minutes in YTB (Table 1 and S1). The cAD1 conjugation pheromone

Table 1. Genes proven or predicted to be important for virulence in V583 found to be regulated during growth in blood.

ORF #	Function (Gene name)	Characteristic	log ₂	Reference
EF0079	<i>gls24</i>	Stress response	3.9	[30,32]
EF0169	lipase/acylhydrolase	Fatty acid and phospholipid degradation	2.9	[97]
EF0361	chitinase, family 2	Cell envelope	4.2	[8]
EF0362	chitin binding protein, putative	Cell envelope	4.4	[8]
EF0373	sensor histidine kinase	Signal transduction	3.0	[12]
EF0394	secreted antigen, putative (<i>salB</i>)	Protein fate	-1.6	[8]
EF0453	OsmC/Ohr family protein (<i>ohr</i>)	Oxidative stress response	3.2	[41,42]
EF0463	superoxide dismutase, Mn (<i>sodA</i>)	Oxidative stress response	1.7	[38]
EF0606	Dps family protein (<i>dps</i>)	Oxidative stress response	2.2	[41]
EF0956	beta-phosphoglucomutase (<i>malB/bopB</i>)	Energy metabolism	2.5	[54]
EF0957	glycosyl hydrolase, family 65 (<i>malA/bopA</i>)	Energy metabolism	2.5	[54]
EF0958	PTS system, IIABC components (<i>malT</i>)	Signal transduction	2.2	[54]
EF1051	sensor histidine kinase (<i>etaS</i>)	Signal transduction	-2.6	[70]
EF1211	NADH peroxidase (<i>npr</i>)	Oxidative stress response	2.8	[41]
EF1586	NADH oxidase (<i>nox</i>)	Oxidative stress response	-2.4	[41]
EF1608	cardiolipin synthetase, putative	Fatty acid and phospholipid biosynthesis	2.5	[98]
EF2074	ABC transporter, ATP-binding protein (<i>efaC</i>)	Transport and binding proteins	3.6	[9,12]
EF2075	ABC transporter, permease protein (<i>efaB</i>)	Transport and binding proteins	2.4	[9,12]
EF2076	endocarditis specific antigen (<i>efaA</i>)	Cellular processes	1.4*	[9,12]
EF2167	glycosyl transferase, group 2 family protein	Cell envelope	-2.1	[8]
EF2170	glycosyl transferase, group 2 family protein	Cell envelope	-2.2	[8]
EF2174	conserved domain protein	Hypothetical proteins	-1.7	[8]
EF2177	bacterial sugar transferase (<i>epaR</i>)	Cell envelope	-2.7	[8,20]
EF2178	membrane protein, putative (<i>epaQ</i>)	Cell envelope	-2.8	[8,20]
EF2179	conserved hypothetical protein (<i>epaP</i>)	Hypothetical proteins	-3.1	[8,20]
EF2180	glycosyl transferase, group 2 family protein (<i>epaO</i>)	Cell envelope	-3.0	[8,20]
EF2181	glycosyl transferase, group 2 family protein (<i>epaN</i>)	Cell envelope	-2.9	[8,20]
EF2182	ABC transporter, ATP-binding protein (<i>epaM</i>)	Transport and binding proteins	-2.5	[8,20]
EF2183	ABC transporter, permease protein (<i>epaL</i>)	Transport and binding proteins	-1.9	[8,20]
EF2189	conserved hypothetical protein (<i>epaJ</i>)	Hypothetical proteins	-2.0	[8,20]
EF2190	glycosyl transferase, group 2 family protein (<i>epaI</i>)	Cell envelope	-1.9	[8,20]
EF2192	dTDP-glucose 4,6-dehydratase (<i>epaG</i>)	Cell envelope	-1.7	[8,20]
EF2194	glucose-1-phosphate thymidyltransferase (<i>epaE</i>)	Cell envelope	-2.2	[8,20]
EF2195	glycosyl transferase, group 2 family protein (<i>epaD</i>)	Cell envelope	-1.9	[8,20]
EF2197	glycosyl transferase, group 2 family protein (<i>epaB</i>)	Cell envelope	-1.8	[8,20]
EF2198	glycosyl transferase, group 4 family protein (<i>epaA</i>)	Cell envelope	-2.1	[8,20]
EF2439	undecaprenol kinase, putative	Toxin production and resistance	-3.2	[8]
EF2485	ABC transporter, permease protein (<i>cpsK</i>)	Transport and binding proteins	-2.9	[10]
EF2486	ABC transporter, ATP-binding protein (<i>cpsJ</i>)	Transport and binding proteins	-3.5	[10]
EF2487	UDP-galactopyranose mutase (<i>cpsI</i>)	Cell envelope	-2.4	[10]
EF2488	lipoprotein, putative (<i>cpsH</i>)	Cell envelope	-2.5	[10]
EF2489	MurB family protein (<i>cpsG</i>)	Cell envelope	-2.9	[10]
EF2490	conserved hypothetical protein (<i>cpsF</i>)	Hypothetical proteins	-3.2	[10]
EF2491	glycosyl transferase, group 2 family protein (<i>cpsE</i>)	Cell envelope	-1.9	[8,10]
EF2492	glycosyl transferase, group 2 family protein (<i>cpsD</i>)	Cell envelope	-2.1	[8,10]
EF2658	FemAB family protein	Toxin production and resistance	-1.7	[8]
EF2713	cell wall surface anchor family protein	Cell envelope	2.8	[8]
EF2738	thioredoxin reductase (<i>ahpF</i>)	Oxidative stress response	-1.8	[41]
EF2748	basic membrane protein DtlB (<i>dltB</i>)	Cell envelope	-2.2	[52]
EF2749	D-alanine-activating enzyme, putative (<i>dltA</i>)	Cell envelope	-2.9	[52]

Table 1. Cont.

ORF #	Function (Gene name)	Characteristic	log ₂	Reference
EF2795	LysM domain lipoprotein	Cell envelope	-2.0	[8]
EF3082	iron compound ABC transporter (<i>fatB</i>)	Transport and binding proteins	3.5	[8]
EF3106	peptide ABC transporter, peptide-binding protein	Transport and binding proteins	6.1	[8]
EF3164	PilB family protein (<i>msrB</i>)	Oxidative stress response	2.1	[41]
EF3191	lipase, putative	Fatty acid and phospholipid degradation	2.3	[8]
EF3198	lipoprotein, YaeC family	Cell envelope	3.2	[8]
EF3245	cell-envelope associated acid phosphatase	Enzymes of unknown specificity	-1.8	[8]
EF3256	pheromone cAD1 precursor lipoprotein	Cell envelope	-2.9	[8]
EF3257	oxidoreductase	Oxidative stress response	-4.4	[41]
EFC0001	pheromone binding protein (<i>prgZ</i>)	Transport and binding proteins	1.6	[8]

*The corresponding p-value was above the chosen level of statistical significance. However the up-regulation of this gene was confirmed by real-time QPCR (Fig. 3). doi:10.1371/journal.pone.0007660.t001

precursor and the YaeC family lipoprotein have both been predicted by Paulsen *et al* to contribute to virulence in V583 [8]. Furthermore, twelve putative membrane-protein encoding genes were affected by growth in blood (six up-regulated and six down-regulated), and three glycosyl transferase genes were down-regulated. We also found a down-regulation of eight genes involved in peptidoglycan biosynthesis. Noticeably, we observed an up-regulation of two genes encoding chitin binding proteins (EF0361 and EF0362) and a gene encoding a cell wall surface anchor family protein, EF2713 (Table 1 and S1), all three proposed as potential virulence factors by Paulsen *et al* [8].

Adaptive Metabolic Shift during Whole Blood Exposure

A massive transcriptional response was observed for genes involved in metabolism when V583 was grown in blood or YTB compared to 2xYT, which could be expected from the results of the growth experiments (Fig. 1). Accordingly, the transcriptome analysis portrayed rapid adjustments of gene expression to accommodate the changed nutritional conditions. The results revealed changed expression of genes involved in pathways in the central metabolism of V583 indicating that a wide range of alternative energy sources were utilized (Table S1).

After 30 minutes of growth in YTB or blood we found a reduced transcription of the main uptake system of glucose in *E. faecalis*, the mannose PTS *mptBACD* (EF0019-22) [57], signifying exhaustion of the glucose reservoir at this stage of the experiment. This notion was supported by the down-regulation of the *ptsHI* operon (EF0709-10), which constitutes the signal transduction components that mediate carbon catabolite repression (CCR) [58]. Furthermore, the observed down-regulation of *pfk-pyk* (EF1045-46) and *fba* (EF1167) involved in the first steps of the glycolysis is also consistent with depletion of intracellular glucose catabolic intermediates. Simultaneously, glycolysis genes *gap-1* (EF1526), glycerate kinase (EF2646) and *pgm* (EF2982) were up-regulated, indicating increased carbon flux from sources other than hexose sugars. Interestingly, the glycerol catabolic pathway (EF1929-27), was highly up-regulated in response to growth in blood, and was also found to be up-regulated after 5, 15 and 30 minutes growth in YTB, suggesting that glycerol and other C3-glycerides from blood serves as a source of energy.

Several metabolic systems subject to catabolite control protein A (CcpA) mediated CCR [58] were also regulated during growth in blood, indicating the use of certain amino acids and available sugars as alternative energy sources. Of these, the gene cluster responsible for citrate catabolism (EF3327-15) and three genes involved in arginine

catabolism, *argF-1* (EF0105), *arcC-1* (EF0106) and EF0108 were up-regulated, while the gene-cluster involved in serine degradation (EF0097-100) showed a reduced expression. After 30 minutes growth in YTB we also found an enhanced expression of catabolism of branched chain amino acids (*ptb*, *buk*, *bdkDAB*, EF1663-59), and the same trend was found during growth in blood. Furthermore, derepression of four PTS systems regulated by CCR mediated binding of CcpA to cis-acting catabolite-responsive elements (*cre*) [58] was observed. These include the predicted cellobiose (EF0292-91), N-acetyl galactosamine (EF0456), lactose/galactose (EF1801) and gluconate (EF3139-36) PTS systems. The latter is part of a predicted metabolic pathway consisting of two operons (EF3142-37 and EF3136-34) that facilitates gluconate uptake and catabolism via the mannonate route [58]. The expression of both operons was enhanced during growth in blood, whereas only some of these genes showed a significantly enhanced expression during growth in YTB.

Significantly altered expression was observed for six PTS systems that probably are regulated by a sigma 54 dependent PTS regulation domain (PRD) and/or Bgl antiterminator mechanisms. Induced transcription of such PTS systems requires both release of CCR and availability of the specific sugars [58]. Two sigma 54 dependent PRD controlled PTS systems, *mphAD* (EF1953-50) [57], and a putative N-acetylglucosamine PTS (EF1516), were down-regulated in YTB and blood respectively. Also, the *mpoAD* PTS system (EF2980-76) [57] was found to be up-regulated and appeared to be co-regulated with EF2982-81. Similarly, the inferred sorbitol metabolism operon (EF3310-04) was up-regulated during growth in blood. Moreover, an indication of co-metabolism of glucose and other sugars in blood was seen by several up-regulated PTS systems for which no CCR mechanism has been identified. This includes the ascorbate PTS *sgaB* (EF1128), during growth in blood and mannitol (EF0411-12) during growth in YTB. The fructose PTS system (EF0717), was up-regulated in blood, but down-regulated in YTB. The PTS mannose/fructose/sorbose (EF3029) showed a reduced expression during growth in both blood and YTB.

The observed changes in substrate utilization and metabolism influenced the pyruvate metabolic pathways. An increased expression of L-lactate dehydrogenase (*ldh-1*; EF0255) was observed in YTB, whereas the *plfAB* genes (EF1612 and EF1613) involved in formate formation were reduced during growth in blood, and after 15 minutes growth in YTB. Furthermore, a reduced expression of *adhE* (EF0900) signifies low contribution of ethanol fermentation in blood. The pyruvate dehydrogenase complex gene-cluster *pdhAB*, *aceF* and *lpdA*

(EF1353-56) involved in acetyl-CoA biosynthesis was up-regulated during growth in blood and also after 15 minutes growth in YTB. The cell can harvest an additional ATP from acetyl-CoA by oxidation to acetate, but no change was detected in transcription of the involved genes *eutD* (EF0949) or *ackA* (EF1983). We previously mentioned up-regulation of the fatty acid biosynthesis pathway FASII, and since acetyl-CoA also serves as the precursor of the FASII pathway, it is possible that acetyl-CoA is funneled to fatty acid biosynthesis.

Blood Specific Components Influence Transport and Biosynthesis Pathways

Iron uptake is a crucial factor in bacterial virulence and in gut colonization of commensal bacteria [59,60]. The mechanisms that enable bacteria to acquire iron from the surroundings have thus received a lot of attention [61]. Our data reveal a potentially important role of iron acquisition and homeostasis during growth in blood. Six genes involved in iron transport, including *feuA* (EF0188), *feoB* (EF0476), *ceuBCD* and *fatB* (EF3085-82) were up-regulated in YTB. The latter four genes comprise an operon that also was up-regulated during growth in blood (Table 1 and S1). The *hrtB* (EF0793) ABC-transporter gene, which homologue in *S. aureus* facilitates expulsion of toxic surplus of heme [60] was up-regulated after 15 minutes growth in YTB. Another scarcely available co-factor is manganese, which also is essential for growth of *E. faecalis in vitro* [62,63]. Notably, a gene cluster, *efaCBA* (EF2074-76) originally identified to encode the endocarditis associated antigen is responsible for Mn²⁺ acquisition in manganese depleted environments [64]. The notion that the ability to acquire manganese is important for growth of *E. faecalis* was sustained by the observed up-regulation of *efaCB* in blood (Table 1 and S1).

The presence of blood also changed the transcription of a number of genes encoding other transport systems in the cell. For example the up-regulation of genes encoding two sugar ABC-transporters (EF1345 and EF1344-43), further supports that V583 utilizes alternative sugars from the blood. Certain transport system encoding genes were down-regulated during growth in blood including a major facilitator ABC transporter (EF0082), amino acid ABC-transporters (EF0761-60, EF2642 & EF2649), a cell division ABC-transporter (EF1760) and a phosphate ABC-transporter (EF1756). Thirteen additional genes encoding ABC-transporters with unknown substrates showed altered transcription in response to growth in blood (3 up-regulated and 10 down-regulated), indicating the requirement for balancing numerous solutes to maintain the cell homeostasis.

Previous studies on the biosynthetic capacities and nutritional requirements of *E. faecalis* have shown that all strains investigated, including the sequenced OG1RF [65], require histidine, isoleucine, methionine, and tryptophan for growth, and that arginine, glutamate, glycine, leucine, or valine was essential for growth of some strains [66]. By comparing the genome sequences of V583 and OG1RF they appear to have similar requirements for amino acids. Since our data show that the transcription of several genes encoding oligo-peptide ABC-transporters (EF0907 and EF3110-06) was up-regulated, and the transcription of two amino acid importer genes (EF0440 and EF0635) was down-regulated (Table S1), it is possible that V583 meets its demand for amino acids by acquiring oligo-peptides when growing in the host bloodstream. This is analogous to what was observed in *Streptococcus pyogenes* in a similar study [67]. Furthermore, the increased expression of *cysK* (EF1584) implies that cysteine is not abundant in blood. Glutamine and glutamate on the other hand seems to be readily available in blood, since we observed a reduced expression of the glutamine synthase operon *glnRA* (EF2160-59), glutamate synthase

gltA (EF2560) and transamination of aspartate to glutamate by *aspB* (EF2372) during growth in blood and/or in YTB.

During growth in blood we noticed up-regulation of an isochorismatase gene (EF3192). This coincided with a down-regulation of the EF1561-68 operon, responsible for biosynthesis of chorismate, which is a precursor of aromatic amino acids, folate and quinones. In agreement with this observation we also noticed a reduced transcription of several genes responsible for biosynthesis of cofactors, prosthetic groups, and carriers including menaquinone and ubiquinone (EF0446-50, EF3255-54 and EF3260). Noticeably, modulation of chorismate acquisition was also observed in similar experiments in *S. pyogenes* and *S. agalactiae* [43,67], suggesting that chorismate might be involved in virulence development of these bacteria.

E. faecalis is prototrophic for purines and pyrimidines [66]. The down-regulation of several genes involved in biosynthesis of these compounds (e.g. EF0014, EF0058, EF1547, EF2362-61 and EF3293) might imply that the requirement for nucleotides was covered by scavenging (Table S1). However, no evidence of modulation of ribose/deoxyribose metabolism was observed, which suggests a lowered demand for nucleotides, most likely as a result of the reduced growth rate (Fig. 1).

Virulence Traits and Regulatory Genes

Several virulence traits have been identified in *E. faecalis* [10,15–21]. The origin of V583 as a nosocomial isolate, causing a persistent bloodstream infection made this strain suitable for investigating responses of virulence traits by growth in blood. As previously described, the expression of several of the loci that contribute to *E. faecalis* virulence, such as oxidative and nutritional stress management genes, capsule formation genes and genes involved in acquisition of biometals, were affected by blood. In addition, some genes predicted to be involved in virulence in *E. faecalis* were found to be regulated, and a summary of these genes can be found in Table 1.

A number of regulatory genes showed altered expression in blood-containing growth environment, in particular regulators of carbohydrate metabolism. In addition, several genes encoding TetR-repressors and other unassigned regulators were found to be differentially expressed (Table S1). V583 inherits 4 sigma factors, of which the *sigA* (EF1522) and *sigV* (EF3180) showed a changed expression in blood. The sigma factor *sigV* was up-regulated, but only one (EF0315) of its five potential target genes [68] showed an altered transcription. The fact that *sigA*, the primary sigma factor of the cell was down-regulated after 30 minutes in blood, is consistent with the observed down-regulation of the entire transcription and protein synthesis apparatus, which in turn can be ascribed the lowered growth rate in blood compared to 2xYT (Fig. 1).

Of the 18 two-component systems present in V583 [8,69], only two of the sensor histidine kinase genes were found to be significantly regulated in blood. Transcription of *etaS* (EF1051) a sensor histidine kinase involved in stress and virulence [70] was reduced, and the expression of a sensor histidine kinase gene (EF0373) previously identified as antigenic (antigen yx84) during infection in humans [12], was highly elevated (Table 1 and S1). However, the exact functions of these two-component systems are not well characterized. The *fsr* locus is an important virulence determinant in *E. faecalis* shown to contribute to virulence in mouse peritonitis models [17]. Due to microarray spot abnormalities, regulation of selected genes including the *fsr* locus was assessed by real-time quantitative RT-PCR (described in further detail below). The RT-PCR revealed that both *fsrB* and *gelE* were down-regulated both in blood and YTB.

Table 2. List of genes and primers (5'→ 3') used for real-time quantitative RT-PCR.

ORF	Gene	Forward primer	Reverse primer	Reference
EF0282	<i>fabI</i>	TGATGGTTTCCTATTAGCACAAG	GTTAGGAATCGCACGTTCCGG	This work
EF0957	<i>bopA</i>	CAGCGACATGGACAGCCTAC	TTGCAGGACCGTCGAGTAAA	This work
EF1099	<i>ace</i>	CGGCGACTCAACGTTTGAC	TCCAGCCAAATCGCCTACTT	[24]
EF1818	<i>gelE</i>	CGGAACATACTGCCGGTTTAGA	TGGATTAGATGCACCCGAAAT	[24]
EF1821	<i>fsrB</i>	TGCTCAAAAAGCAAAGCCTTATAA	GATGACGAGACCGTAGAGTATTACTGAA	[24]
EF2076	<i>efaA</i>	TGGGACAGACCCTCACGAATA	CGCCTGTTTCTAAGTTCAAGCC	[24]
EF2493	<i>cpsC</i>	GGTTGATGCCAAGAGCTCAG	GTCCCATGCCACGCTGTAT	This work
EF2749	<i>dltA</i>	ACGCGTTTGCACAAATTAAC	GCGCAGTGCTGGTAGATGTT	This work
EF2883	<i>fabK</i>	GCTGGATTGCCATGACCTGTGCG	GGTAGCCGATGCTTCATTAGCAAGTGC	This work
EF2886	<i>fabT</i>	ACTACACGTCGATCATCTTCACTACGC	CATTACGGAGATGCACACAATCGAAGC	This work
EF3192	-	GAACGACGGGCGTAATCTG	GTCCAAATCCGTGCCACTAA	This work
EF3193	<i>lrgB</i>	CGACAGTAGCGTTTGGCGATT	ACAGCCACTAGCGAACCCAAA	This work
EF3198	-	GCTGATTTAGTGGCTGTGCAA	AGCACGACCTTCATTGGTTG	This work
235		CCTATCGGCCCTCGGCTTAG	AGCGAAAGACAGGTGAGAATCC	[24]

doi:10.1371/journal.pone.0007660.t002

Gene Regulation Examined by Real-time Quantitative RT-PCR

In order to confirm the results from the microarray experiments and to investigate the transcription of a few genes of special interest (excluded from the microarray results due to spot abnormalities), real-time quantitative RT-PCR (QPCR) was performed on 13 genes listed in Table 2. We only examined the 30 minutes time point from the YTB experiment, which is most comparable to the blood experiment. The QPCR results were

consistent with the results obtained by the microarray experiments (Fig. 3). For most of the genes the QPCR produced similar or greater \log_2 ratios than the corresponding microarray results. One gene, *efaA*, showed a \log_2 -ratio of 1.4 during growth in blood, but with a p-value indicating non-significance. However, the QPCR showed a \log_2 -ratio identical to the microarray results supporting that this gene is in fact up-regulated. Most of the genes tested by QPCR were not found to be significantly regulated in YTB using microarrays. However, the QPCR-analysis showed equal to larger

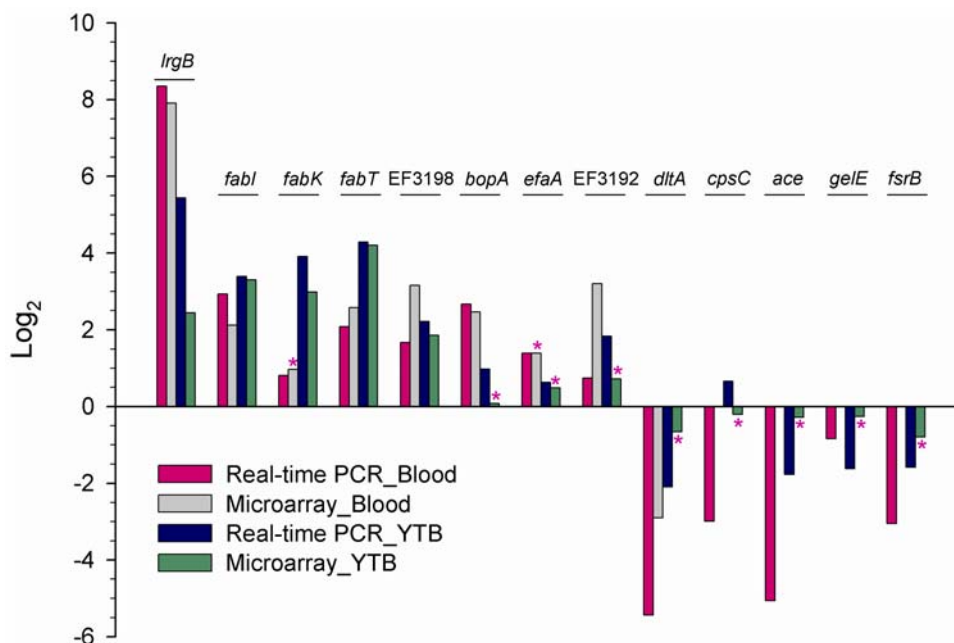


Figure 3. Gene regulation (\log_2) after 30 minutes growth in either blood or YTB compared to 2xYT analyzed by microarray or real-time quantitative RT-PCR. The asterisks indicate values from the microarray experiments that were found to be outside the rejection level. The corresponding orf-numbers for the genes tested are: *lrgB*; EF3193, *fabI*; EF0285, *fabK*; EF2883, *fabT*; EF2886, *bopA*; EF0957, *efaA*; EF2076, *dltA*; EF2749, *cpsC*; EF2493 *ace*; EF1099, *gelE*; EF1818, *fsrB*; EF1821.

doi:10.1371/journal.pone.0007660.g003

responses in terms of fold change of these genes. An exception was the gene *cpsC* which showed a slightly reduced expression when examined by microarrays, while a slightly enhanced expression was seen using QPCR. Thus, the QPCR analysis confirmed the reliability of the transcriptional data obtained from the microarray experiments, indicating that the rejection level applied on the microarray data was adequate. Furthermore, the four genes which were excluded from the microarray results due to spot abnormalities (*cpsC*, *ace*, *gelE* and *fsrB*) were all found to be down-regulated in blood when examined by QPCR.

Discussion

In the present study we describe the first microarray analysis of the global transcriptional response of *E. faecalis* to blood. Although certain aspects of *E. faecalis* virulence development in blood have been studied [23–25], the global gene regulation and interplay during adaptation to the blood environment remained largely unaddressed. However, *in vivo* mouse infection models in which *E. faecalis* was intravenously injected, showed that the bacterial cell number initially dropped before stabilizing at about 10^3 CFU/ml blood for at least 7 days [27]. This indicates that *E. faecalis* has to adapt to overcome serious challenges upon entering the bloodstream of a host. With the present study we provide novel information about the gene regulation relevant to such an adaptation process.

Transcriptomics has been successfully employed for studying adaptation to blood *ex vivo* of the closely related pathogens *S. pyogenes* [67] and *S. agalactiae* [43]. As emphasized in these studies [43,67], it is important to note that *in vitro* experiments like the one we present here, can not entirely reproduce the environment encountered by the bacterium during *in vivo* infections with respect to oxygen tension, interaction with the host immune system, the potential depletion of nutrients etc. Consequently, our results have been cautiously interpreted with respect to inherent constraints. Hence, we have used this *in vitro* model to investigate the initial adaptation phase needed for dissemination of an infection, and to explore molecular mechanisms that might be involved in the complex host-bacterium interaction in blood.

The most significant transcriptional changes found in this study include genes relevant for cell envelope structures. In the cell membrane both the fatty acid (FASII) and phosphatidylglycerol biosynthesis genes were up-regulated. This suggests that *E. faecalis* adjusts its fatty acid composition and membrane fluidity to accommodate stress imposed by blood constituents, and possibly also by involving phospholipids from the blood. A recent study demonstrated the ability of *S. agalactiae*, *S. pyogenes*, *S. pneumoniae* and *E. faecalis* to utilize free phospholipids in serum and thus overcome the FASII pathway inhibiting antibiotics [71]. Brinster *et al* [71] also showed by QPCR that growth of *S. agalactiae* in serum results in a down-regulation of the eight FASII genes tested. Moreover, other studies have shown by microarray that the gene clusters involved in FASII were down-regulated in *S. pyogenes*, but not significantly regulated in *S. agalactiae* or *S. pneumoniae* during growth in blood [43,67,72]. Although *E. faecalis* and the three streptococcal species mentioned above show a similar phenotype when grown in serum containing FASII inhibiting drugs, it appears that the regulation of the FASII pathway is different during growth in blood. This might be due to the differences in the genetic organization of the FASII pathway between enterococci and streptococci. Another possible explanation is that *E. faecalis* processes the unsaturated fatty acids from blood in order to accommodate a lipid composition compatible with its membrane. The observed increased transcription of two lipase genes during

growth in blood might imply a connection between the modulation of fatty acid and phospholipid metabolism and lipolysis/tissue tropism. A recent study showed that 71% of the invasive *E. faecalis* isolates examined produced lipase, whereas only 35% of the noninvasive isolates produced lipase indicating that lipase activity might be important for the pathogenicity of *E. faecalis* [73].

It has been shown that orally administered or intravenously injected *E. faecalis* in mice can colonize internal organs of the host [27,28]. Moreover, Guzman and co-workers [74] showed that growth of *E. faecalis* in serum enhanced its ability to bind Girardi Heart cells, an interaction which in a later study was shown to be inhibited by incubation of the target cells with specific sugar residues. This indicated that carbohydrate antigens were responsible for the enhanced binding [75]. We observed that transcription of two capsular polysaccharides encoded by *epa* and *cps* were down-regulated during growth in blood. Inactivation of these carbohydrate antigen loci cause reduced biofilm formation [20,76], increased susceptibility to phagocytosis [20,51], and attenuated virulence in mice [48]. More interestingly, these exopolysaccharides appear to play a role in adherence/colonization of tissues and organs, and in immune system evasion [10,11,20,50,51]. The involvement of carbohydrate antigens in binding has been further investigated by Singh *et al*, who showed that the *epa* locus is important for adherence/colonization of tissues and organs [50]. Capsule formation is crucial for virulence in *S. pneumoniae*, which undergoes phase variation in order to establish infection [77]. However, an *in vivo* transcriptome analysis of *S. pneumoniae* infection in mice has revealed that the *cps* transcription was not enhanced in blood. Furthermore, this study also demonstrated that the expression of a number of virulence traits in *S. pneumoniae* was body site dependent [72]. Hence, it is possible that the transcription of the *E. faecalis epa* and *cps* loci might be more pronounced in colonized organs.

The effect of serum on *E. faecalis* adherence has been studied further by Nallapareddy and Murray [78], who found that 46 different *E. faecalis* strains all showed enhanced binding to fibronectin and fibrinogen after exposure to 40% horse serum. This property was eliminated by protease treatment, which indicated that adherence was mediated by surface exposed proteins [78]. In our study most genes encoding identified or putative adhesive proteins (such as microbial surface components recognizing adhesive matrix molecules (MSCRAMMs), the collagen adhesin *ace* and aggregation substances) were either not regulated or down-regulated during growth in blood. However, we observed enhanced transcription of several genes encoding membrane proteins or lipoproteins. It is tempting to speculate that these cell envelope bound proteins also might play a role in promoting the adherence of *E. faecalis* during infection.

Analysis of the V583 transcriptome revealed signs of lytic stress in response to blood. Particularly interesting was the immense up-regulation of the *lrgAB* operon. In *S. aureus* it has been shown that the transcription of *lrgAB* is affected by carbohydrate metabolism, acid production or cell wall active antibiotics. LrgAB inhibit murein hydrolase activity, and thus counteract lysis of the cell [79,80]. The exact function of *lrgAB* in *E. faecalis* remains elusive. Even so, the immediate and continuously high level transcription of this operon observed in the present study, and the fact that the homologous genes in *S. agalactiae* also showed an enhanced transcription during growth in blood [43], suggests an important role for *lrgAB* upon blood exposure.

Biometal limitation constitutes a significant obstacle for bacterial pathogens to establish infection in vertebrate hosts [59,60,64]. Lactic acid bacteria (LAB) comprise one of the very

few groups of bacteria for which iron is not an essential growth factor [63,81]. In contrast, manganese is essential to the fermentative metabolism of LAB [62,63]. The enhanced transcription of the main manganese scavenging mechanism encoded by *efaCBA* is a clear indication that *E. faecalis* experiences manganese constraints that might restrict its growth in blood. The strong increase in the superoxide dismutase (*sodA*) expression further emphasizes the importance of an effective manganese uptake during growth in blood. These observations are particularly interesting as both *efaCBA* and *sodA* have been shown to be implicated in virulence [38,82]. In addition, our microarray data show that genes related to iron metabolism constituted one of the major changes of *E. faecalis* adaptation to blood, although iron is not an essential requirement for growth. This suggests a potential role of iron acquisition in virulence development of *E. faecalis*. Examination of the V583 genome has revealed approximately 53 genes with an apparent function in iron homeostasis and metabolism. It was recently demonstrated that many LAB can utilize heme to perform respiratory metabolism [83,84], and that the pathogen *S. agalactiae* requires heme for full virulence [85]. Thus, the significance of the biometals manganese and iron/heme in virulence of *E. faecalis* warrants further investigation.

Previous studies have demonstrated the ability of *E. faecalis* to sense its environment and modulate cytolysin production upon blood encounter [23]. The V583 strain is deficient of cytolysin production, but is classified as a pathogen since it was isolated from a patient with a persistent bloodstream infection [6] and belongs to a high risk clonal complex consisting of nosocomial isolates [7]. Consequently, it was relevant to look at the response of virulence related genes. The V583 strain harbors an incomplete (partially deleted) *E. faecalis* pathogenicity island (PAI) [86]. From the PAI of V583 only one gene, *dps* (EF0606) involved in oxidative stress management, was up-regulated. A few genes involved in pantothenate biosynthesis, amino acid metabolism and a transposase were down-regulated, while transcription of the remaining genes within the PAI were unaffected. It is important to note that most genes in the PAI were indeed expressed, and hence might contribute to the fitness of V583, but in the experimental growth conditions examined here, no major responses in transcription of PAI-genes were revealed.

A number of pathogens employ master regulatory systems to coordinate expression of virulence factors during infection e.g. *prfA* in *Listeria monocytogenes* [87], *agr* in *S. aureus* [88] and *cooRS* in *S. pyogenes* [89]. The equivalent system in *E. faecalis* is the *fsr* quorum sensing system, which controls the expression of several genes including *gelE* (encoding a gelatinase) and *sprE* (encoding a serine proteinase) [17,90,91]. It has previously been shown that expression of this system is sensitive to environmental conditions [24], and that its level of expression varies between different bacterial strains [92]. In our study we readily detected the *fsrB* and *gelE* transcripts by QPCR, but growth in blood caused a moderate down-regulation of these genes in V583. This appears to be the opposite effect from growth in serum by the MMH594 strain [24]. Our results demonstrate that blood did not act as a cue to enhance *fsr* expression, but rather interfered with its expression. It has been shown that the cytolysin quorum sensing pheromones interact with blood cells in a positive manner [23]. It is possible that the down-regulation was caused by the presence of blood cells in our experiments, which might have adsorbed the quorum sensing signaling pheromone. It has been proposed that the GelE and SprE proteases contribute to virulence by degrading infected tissue [93], and it is possible that *fsr* expression is more pronounced in colonized organs.

In conclusion, this study provides new insights into the adaptive process of *E. faecalis* to growth and persistence in blood. Bacteremia caused by *E. faecalis* represents a major clinical problem, thus the results presented here could be valuable for future studies devoted to the development of new therapeutic approaches for preventing or treating enterococcal infections.

Materials and Methods

Cultivation and Growth Measurement

The strain used in this study was the sequenced *E. faecalis* clinical isolate, V583 ([6,8]). For all experiments V583 was streaked on a 2xYT (1% (w/v) yeast extract, 1.6% (w/v) tryptone and 1% (w/v) NaCl) agar plate and incubated at 37°C o/n. Four individual colonies from the 2xYT plate were inoculated into the same tube of 5 ml 2xYT medium and grown for 17 hours without shaking at 37°C. The culture was then diluted 1000x in 2xYT medium (pre-warmed to 37°C) and incubated until the culture reached an optical density at 600 nm (OD₆₀₀) of 0.1.

Growth of the bacterium was monitored by counting colony forming units (CFU). A 150 ml culture grown to an OD₆₀₀ of 0.1 as described above was split in three and centrifuged at 8000 x g for 3 minutes at 37°C. The cells were resuspended in either (i) 50 ml defibrinated horse blood (TCS Biosciences Ltd.), (ii) 45 ml 2xYT medium and 5 ml blood (10% (v/v) blood) or (iii) 50 ml 2xYT medium (all pre-warmed to 37°C). After thorough mixing 5 ml from each culture was immediately transferred to a fresh tube and placed on ice and the remaining culture was incubated further at 37°C without shaking. The 5 ml cultures were sonicated at an amplitude of 25% 25 seconds (5 seconds on and 5 seconds off) using a Vibra-Cell VCX-500 ultrasonic processor (Sonics) with a microtip. Immediately after sonication 500 µl of each culture was serially diluted in 0.9% (w/v) NaCl (pre-chilled) and plated on 2xYT agar plates. The whole process was repeated after 30, 60, 90, 120, 240 and 360 minutes and the experiment was performed on three consecutive days.

Cultivation and Sampling Prior to Microarray Analysis

A pre-culture of V583 was prepared as described above. After 17 hours the culture was diluted 1000x in 450 ml 2xYT medium and incubated at 37°C until the culture reached an OD₆₀₀ of 0.1, when the culture was split in two. A volume of 25 ml pre-warmed 2xYT medium (37°C) was added to the control culture whereas 25 ml pre-warmed blood was added to the test culture, resulting in a final concentration of 10% (v/v) blood. Samples (25 ml) from each culture were collected by centrifugation (8000 x g for 2 minutes at 37°C) after 5, 15, 30 and 60 minutes, and the cell-pellets were immediately frozen in liquid Nitrogen and stored at -80°C prior to RNA extraction.

A parallel experiment was designed to investigate the transcriptional response in pure blood. A pre-culture of V583 was diluted 1000x in 200 ml 2xYT medium and incubated at 37°C until the culture reached an OD₆₀₀ of 0.1, when the culture was split in two and centrifuged (8000 x g for 3 minutes at 37°C). For the control culture the cells were resuspended in 100 ml pre-warmed 2xYT (37°C) whereas for the test culture the cells were resuspended in 100 ml pre-warmed blood (37°C), resulting in a final concentration of 100% (v/v) blood in the test culture. Samples (35 ml) of each culture were collected by centrifugation (8000 x g for 2 minutes at 37°C) 30 minutes after addition of blood, and the cell-pellets were immediately frozen in liquid Nitrogen and stored at -80°C prior to RNA extraction.

RNA Isolation

The bacterial cells were washed with 50 ml cold 0.1xTE (for 10% blood) and 3x50 ml cold 0.1xTE (for 100% blood) to remove blood constituents prior to RNA extraction. Then samples suspended in 700 μ l RLT buffer (Qiagen) were transferred to a 2 ml screw cap FastPrep tube (Qbiogene) containing 0.6 g of glass beads ($\leq 106 \mu\text{m}$) (Sigma) and 300 μ l chloroform (Merck). Cells were lysed by vigorous shaking for 20 seconds at 6.0 m/s in a FP120 FastPrep cell disruptor (Qbiogene). After lysis the samples were placed on ice for 5 minutes before glass beads and chloroform were removed by a brief centrifugation. The aqueous phase was transferred to a new tube and centrifuged at $\sim 8000 \times g$ for 2 minutes to remove cell debris and unlysed cells. The supernatants were removed and kept on ice in separate tubes, while the pellets were suspended in 350 μ l RLT buffer, transferred to new FastPrep tubes and subjected to another round of homogenization. The two supernatants from each sample were merged and added 750 μ l 96% EtOH. Total RNA was then isolated using the RNeasy Mini kit (Qiagen) according to the manufacturer's protocol. The integrity of the RNA samples was analyzed using the RNA 600 Nano LabChip kit and a Bioanalyzer 2100 (Agilent Technologies). The concentration and purity of the total RNA was measured using a NanoDrop ND-1000 spectrophotometer (NanoDrop Technologies, Inc.).

cDNA Synthesis and Fluorescent Labeling

Total RNA was reversed transcribed using a modified version of protocol #M007 from the Pathogen Functional Genomic Resource center at The Institute for Genomic Research (TIGR: <http://pfgrc.tigr.org/protocols/M007.pdf>). Accordingly, 5 μ g of total RNA and 20 μ g of random hexamers (Invitrogen) in a reaction volume of 17.3 μ l were denatured at 70°C for 10 minutes and cooled on ice for 5 minutes. Then, 6 μ l of 5x First Strand buffer (Invitrogen), 3 μ l of 0.1 M dithiothreitol, 20 U rRNasin (Promega), 1.2 μ l of a 12.5 mM dNTP (Invitrogen) and aminoallyl-dUTP (Ambion) labeling mixture (aa-dUTP-dTTP 2:3), and 400 U of SuperScript III reverse transcriptase (Invitrogen) were added in a total volume of 30 μ l. The labeling reaction mixture was incubated at 25°C for 5 minutes and then at 42°C for 16 hours. The RNA was hydrolyzed by adding 10 μ l of 0.5 M EDTA and 10 μ l of 1 M NaOH. The reaction mixture was incubated at 65°C for 15 minutes, and then neutralized by adding 25 μ l of 1 M Tris-HCl (pH 7.0). Purification of the cDNA was performed using Microcon YM-30 filters (Millipore) according to the manufacturer's protocol. The cDNA was dried in a vacuum centrifuge and stored at -20°C . Coupling of aminoallyl-labeled cDNA to Cy3 and Cy5 (Amersham Biosciences) was done by resuspending the cDNA in 9 μ l 0.1 M sodium carbonate buffer pH 9.3. The cDNA samples were transferred to dried Cy-dye aliquots (dissolved in DMSO and dried in a vacuum centrifuge prior to labeling), mixed and incubated for 1 hour. A volume of 35 μ l 100 mM sodium acetate pH 5.2 was added and unincorporated dye was removed using QIAquick PCR purification kit (Qiagen) according to the manufacturer's protocol. Finally the Cy3- and Cy5-labeled samples were mixed and dried in a vacuum centrifuge.

Hybridization and Microarray Data Analysis

Microarray experiments were performed using whole genome *E. faecalis* V583 PCR-based microarrays described by Aakra et al [94]. Prior to hybridization, the Ultra Gaps slides (Corning) were prehybridized according to the manufacturer's recommendations. Briefly, the arrays were incubated in a prehybridization solution (3x SSC, 0.1% (wt/vol) sodium dodecyl sulphate (SDS), 0.1 mg/

ml Bovine Serum Albumin (BSA), Sigma) at 50°C for 30 minutes. After prehybridization, the arrays were washed twice in distilled water (RT) for 1 minute, then at 95°C in distilled water for 2 minutes, followed by a 1 minute wash in isopropanol and the slides were dried by centrifugation (70 x g for 5 minutes) in an Eppendorf 5810R tabletop centrifuge. The Cy3- and Cy5-labeled cDNA samples were resuspended in 40 μ l hybridization solution (3x SSC, 0.1% (wt/vol) SDS, 1 mg/ml BSA, 0.1 mg/ml Salmon Sperm DNA (Invitrogen), 50% (vol/vol) Formamide), denatured by boiling for 2 minutes and cooled at room temperature for 5 minutes. The samples were centrifuged briefly and applied to the prehybridized microarray under a 25x60 mm LifterSlip (Eric Scientific Company). The microarrays were hybridized at 42°C for 16 hours. After hybridization, the slides were washed in 2x SSC buffer with 0.1% (wt/vol) SDS for 2 minutes, followed by a wash in 1x SSC for 2 minutes, then in 0.2x SSC for 2 minutes and finally in 0.05x SSC for 1 minute, each at room temperature. The slides were dried by centrifugation (70 x g for 5 minutes). Three biological replicates with one dyeswap were performed for all experiments. Microarray slides were scanned at 10 μm resolution using a Model G2505B (Agilent) microarrayscanner. Fluorescent intensities and morphologies were analyzed using GenePix Pro ver. 6.0 (Axon).

Normalization and Data Analysis

Raw data from each array was preprocessed independently. A lowess-smoothed background was subtracted from all foreground intensities, and a cross-validated lowess-method was used in an intensity-dependent normalization of every array. The \log_2 ratios for each spot were further analyzed using a mixed model [29] to detect differentially expressed genes. For YTB a mixed model was fitted to the data for each of the four sample times (5, 15, 30 and 60 minutes) separately and for blood a mixed model was fitted to the data from the 30 minutes time point. Data for the three arrays at every sample time were described by

$$y_{ijk} = \mu_i + u_{ij} + e_{ijk} \quad (1)$$

Where y_{ijk} is the observed \log_2 ratio of gene i (1,...,3502) on array j (1,2,3) and in spot k (1,...,5) on that array, μ_i is the expected \log_2 ratio for gene i , u_{ij} is a random effect of gene i on array j and e_{ijk} is the remaining noise. The variance components were estimated under the assumption of gaussian errors using a restricted maximum likelihood approach coping with the unbalanced data due to missing spots. Differentially expressed genes were identified by testing the hypothesis $H_0: \mu_i = 0$ against $H_1: \mu_i \neq 0$. A chi-square test for every gene resolves this for the model in [1,95], and a Bonferroni-corrected rejection level of $p < 0.05$ was used throughout. If $H_0: \mu_i = 0$ was rejected, and $\mu_i > 0$, genes were considered to be up-regulated in the cells grown in YTB and blood. If $H_0: \mu_i = 0$ was rejected, and $\mu_i < 0$, genes were considered to be down-regulated. All data analysis algorithms were programmed in MATLAB (MathWorks inc). A gene was discarded from the final results (designated NA in Table S1) if it was replicated in less than 8 spots after filtering of the data.

Microarray Data Accession Number

The microarray data obtained in this study has been deposited in the ArrayExpress database (<http://www.ebi.ac.uk/arrayexpress/>) according to the MIAME standard. The accession number is E-TABM-541.

Real-time Quantitative RT-PCR

Real time quantitative RT-PCR (QPCR) was used to validate the expression levels for selected genes. QPCR was performed on a Rotor-Gene 6000 centrifugal amplification system (Corbett Research). The genes of interest and the corresponding primers are listed in Table 2. Total RNA used for cDNA synthesis was the same as described above (RNA samples harvested after 30 minutes growth in YTB or blood and their corresponding control samples). Synthesis of cDNA was performed using 1 µg total RNA, 6 µg random primers (Invitrogen) and 40 U RNase OUT (Invitrogen) in a reaction volume of 12.5 µl. The reaction mixture was denatured at 65°C for 10 minutes and cooled at 4°C for 5 minutes. The reaction mixture was then added 4 µl of 5x First Strand buffer (Invitrogen), 1 µl of 0.1 M dithiothreitol (Invitrogen), 1 µl of 10 µM dNTP (Invitrogen) and 140 U of SuperScript III reverse transcriptase (Invitrogen) to a total volume of 20 µl. The reaction mixture was incubated at 25°C for 5 minutes, 2 hours at 50°C, and then for 15 minutes at 70°C. Finally, 20 ng RNaseA (Sigma-Aldrich) was added to the reaction followed by an incubation at 37°C for 20 minutes. PCR amplification was performed using the recommendation by the manufacturer (with an annealing temperature of 60°C) with 2.5 µl 100x diluted cDNA in a 25-µl reaction mixture containing 12.5 µl FastStart SYBR green Master (Roche) and 7.5 µM of each primer. Standard curves with four dilutions were made in duplicates for each primer pair to calculate the amplification efficiency, and all genes were quantified in triplicates. Since the standard curves indicated a slight difference in amplification efficiencies of the different target genes and reference, differential expression was calculated by the Pfaffl method. This is an optimal method to use for calculating relative gene expression when the amplification efficiencies of the target gene and the reference gene are different since the amplification efficiency is included in the calculation [96]. The obtained C_T value for 23S from each sample was used as a reference for each gene in the corresponding samples.

References

- Noble CJ (1978) Carriage of group D streptococci in the human bowel. *J Clin Pathol* 31: 1182–1186.
- Flahaut S, Laplace JM, Frere J, Auffray Y (1998) The oxidative stress response in *Enterococcus faecalis*: relationship between H₂O₂ tolerance and H₂O₂ stress proteins. *Lett Appl Microbiol* 26: 259–264.
- Sherman JM, Mauer JC, Stark P (1937) *Streptococcus faecalis*. *J Bacteriol* 33: 275–282.
- Richards MJ, Edwards JR, Culver DH, Gaynes RP (2000) Nosocomial infections in combined medical-surgical intensive care units in the United States. *Infect Control Hosp Epidemiol* 21: 510–515.
- Huycke MM, Sahn DF, Gilmore MS (1998) Multiple-drug resistant enterococci: the nature of the problem and an agenda for the future. *Emerg Infect Dis* 4: 239–249.
- Sahn DF, Kissinger J, Gilmore MS, Murray PR, Mulder R, et al. (1989) In vitro susceptibility studies of vancomycin-resistant *Enterococcus faecalis*. *Antimicrob Agents Chemother* 33: 1588–1591.
- Ruiz-Garbajosa P, Bonten MJ, Robinson DA, Top J, Nallapareddy SR, et al. (2006) Multilocus sequence typing scheme for *Enterococcus faecalis* reveals hospital-adapted genetic complexes in a background of high rates of recombination. *J Clin Microbiol* 44: 2220–2228.
- Paulsen IT, Banerjee L, Myers GS, Nelson KE, Seshadri R, et al. (2003) Role of mobile DNA in the evolution of vancomycin-resistant *Enterococcus faecalis*. *Science* 299: 2071–2074.
- Lowe AM, Lambert PA, Smith AW (1995) Cloning of an *Enterococcus faecalis* endocarditis antigen: homology with adhesins from some oral streptococci. *Infect Immun* 63: 703–706.
- Hancock LE, Gilmore MS (2002) The capsular polysaccharide of *Enterococcus faecalis* and its relationship to other polysaccharides in the cell wall. *Proc Natl Acad Sci U S A* 99: 1574–1579.
- Hancock LE, Shepard BD, Gilmore MS (2003) Molecular analysis of the *Enterococcus faecalis* serotype 2 polysaccharide determinant. *J Bacteriol* 185: 4393–4401.
- Xu Y, Jiang L, Murray BE, Weinstock GM (1997) *Enterococcus faecalis* antigens in human infections. *Infect Immun* 65: 4207–4215.

Supporting Information

Table S1 Microarray expression data from *E. faecalis* strain V583 during incubation in blood or 2xYT supplemented with 10% blood (YTB). Gene expression after 30 minutes (blood) or 5, 15, 30 and 60 minutes (YTB) of incubation is relative to the expression during growth in 2xYT for the corresponding time length. a) Genes comprising putative operon structures predicted by <http://biocyc.org> [1,2] are marked with one color (red or light red for genes on the leading strand, blue or light blue for genes in on the lagging strand). b) Log₂-values greater than 1 or less than -1 are highlighted in red or green respectively. Genes for which less than 8 spots were present were discarded from the analysis and are denoted “NA”. c) A significantly regulated gene (bonferroni corrected level of p<0.05) has the corresponding p-value written in bold. 1. Paley SM, Karp PD (2002) Evaluation of computational metabolic-pathway predictions for *Helicobacter pylori*. *Bioinformatics* 18: 715–724. 2. Romero PR, Karp PD (2004) Using functional and organizational information to improve genome-wide computational prediction of transcription units on pathway-genome databases. *Bioinformatics* 20: 709–717.

Found at: doi:10.1371/journal.pone.0007660.s001 (0.34 MB PDF)

Acknowledgments

We thank Bjorn E. Kristiansen, the Norwegian Microarray Consortium, Oslo, for printing the microarray slides and Linda H. Godager for performing the real-time quantitative RT-PCR.

Author Contributions

Conceived and designed the experiments: HCV DAB. Performed the experiments: HCV. Analyzed the data: HCV LS IFN DAB. Contributed reagents/materials/analysis tools: LS IFN. Wrote the paper: HCV LS IFN DAB.

- Xu Y, Murray BE, Weinstock GM (1998) A cluster of genes involved in polysaccharide biosynthesis from *Enterococcus faecalis* OG1RF. *Infect Immun* 66: 4313–4323.
- Mundy LM, Sahn DF, Gilmore M (2000) Relationships between enterococcal virulence and antimicrobial resistance. *Clin Microbiol Rev* 13: 513–522.
- Ike Y, Hashimoto H, Clewell DB (1984) Hemolysin of *Streptococcus faecalis* subspecies zymogenes contributes to virulence in mice. *Infect Immun* 45: 528–530.
- Chow JW, Thal LA, Perri MB, Vazquez JA, Donabedian SM, et al. (1993) Plasmid-associated hemolysin and aggregation substance production contribute to virulence in experimental enterococcal endocarditis. *Antimicrob Agents Chemother* 37: 2474–2477.
- Qin X, Singh KV, Weinstock GM, Murray BE (2000) Effects of *Enterococcus faecalis* *fsr* genes on production of gelatinase and a serine protease and virulence. *Infect Immun* 68: 2579–2586.
- Sifri CD, Mylonakis E, Singh KV, Qin X, Garsin DA, et al. (2002) Virulence effect of *Enterococcus faecalis* protease genes and the quorum-sensing locus *fsr* in *Caenorhabditis elegans* and mice. *Infect Immun* 70: 5647–5650.
- Schlievert PM, Gahr PJ, Assimacopoulos AP, Dinges MM, Stoehr JA, et al. (1998) Aggregation and binding substances enhance pathogenicity in rabbit models of *Enterococcus faecalis* endocarditis. *Infect Immun* 66: 218–223.
- Teng F, Jacques-Palaz KD, Weinstock GM, Murray BE (2002) Evidence that the enterococcal polysaccharide antigen gene (*epa*) cluster is widespread in *Enterococcus faecalis* and influences resistance to phagocytic killing of *E. faecalis*. *Infect Immun* 70: 2010–2015.
- Huycke MM, Abrams V, Moore DR (2002) *Enterococcus faecalis* produces extracellular superoxide and hydrogen peroxide that damages colonic epithelial cell DNA. *Carcinogenesis* 23: 529–536.
- Chandler JR, Hirt H, Dunny GM (2005) A paracrine peptide sex pheromone also acts as an autocrine signal to induce plasmid transfer and virulence factor expression in vivo. *Proc Natl Acad Sci U S A* 102: 15617–15622.
- Coburn PS, Pillar CM, Jett BD, Haas W, Gilmore MS (2004) *Enterococcus faecalis* senses target cells and in response expresses cytolysin. *Science* 306: 2270–2272.

24. Shepard BD, Gilmore MS (2002) Differential expression of virulence-related genes in *Enterococcus faecalis* in response to biological cues in serum and urine. *Infect Immun* 70: 4344–4352.
25. Kreft B, Marre R, Schramm U, Wirth R (1992) Aggregation substance of *Enterococcus faecalis* mediates adhesion to cultured renal tubular cells. *Infect Immun* 60: 25–30.
26. Hirt H, Schlievert PM, Dunny GM (2002) *In vivo* induction of virulence and antibiotic resistance transfer in *Enterococcus faecalis* mediated by the sex pheromone-sensing system of pCF10. *Infect Immun* 70: 716–723.
27. Gentry-Weeks C, Estay M, Loui C, Baker D (2003) Intravenous mouse infection model for studying the pathology of *Enterococcus faecalis* infections. *Infect Immun* 71: 1434–1441.
28. Wells CL, Jechorek RP, Erlandsen SL (1990) Evidence for the translocation of *Enterococcus faecalis* across the mouse intestinal tract. *J Infect Dis* 162: 82–90.
29. Wernisch L, Kendall SL, Sondej S, Wietzorrek A, Parish T, et al. (2003) Analysis of whole-genome microarray replicates using mixed models. *Bioinformatics* 19: 53–61.
30. Teng F, Nannini EC, Murray BE (2005) Importance of *gls24* in virulence and stress response of *Enterococcus faecalis* and use of the Gls24 protein as a possible immunotherapy target. *J Infect Dis* 191: 472–480.
31. Capiiaux H, Giard JC, Lemariniere S, Auffray Y (2000) Characterization and analysis of a new gene involved in glucose starvation response in *Enterococcus faecalis*. *Int J Food Microbiol* 55: 99–102.
32. Nannini EC, Teng F, Singh KV, Murray BE (2005) Decreased Virulence of a *gls24* Mutant of *Enterococcus faecalis* OG1RF in an Experimental Endocarditis Model. *Infect Immun* 73: 7772–7774.
33. Giard JC, Hartke A, Flahaut S, Boutibonnes P, Auffray Y (1997) Glucose starvation response in *Enterococcus faecalis* JH2-2: survival and protein analysis. *Res Microbiol* 148: 27–35.
34. Rince A, Flahaut S, Auffray Y (2000) Identification of general stress genes in *Enterococcus faecalis*. *Int J Food Microbiol* 55: 87–91.
35. Klebanoff SJ (1980) Oxygen metabolism and the toxic properties of phagocytes. *Ann Intern Med* 93: 480–489.
36. Thomas EL, Lehrer RI, Rest RF (1988) Human neutrophil antimicrobial activity. *Rev Infect Dis* 10 Suppl 2: S450–456.
37. La Carbona S, Sauvageot N, Giard JC, Benachour A, Posteraro B, et al. (2007) Comparative study of the physiological roles of three peroxidases (NADH peroxidase, Alkyl hydroperoxide reductase and Thiol peroxidase) in oxidative stress response, survival inside macrophages and virulence of *Enterococcus faecalis*. *Mol Microbiol* 66: 1148–1163.
38. Verneuil N, Maze A, Sanguinetti M, Laplace JM, Benachour A, et al. (2006) Implication of (Mn)superoxide dismutase of *Enterococcus faecalis* in oxidative stress responses and survival inside macrophages. *Microbiology* 152: 2579–2589.
39. Verneuil N, Rince A, Sanguinetti M, Posteraro B, Fadda G, et al. (2005) Contribution of a PerR-like regulator to the oxidative-stress response and virulence of *Enterococcus faecalis*. *Microbiology* 151: 3997–4004.
40. Giard JC, Riboulet E, Verneuil N, Sanguinetti M, Auffray Y, et al. (2006) Characterization of Ers, a PrfA-like regulator of *Enterococcus faecalis*. *FEMS Immunol Med Microbiol* 46: 410–418.
41. Riboulet E, Verneuil N, La Carbona S, Sauvageot N, Auffray Y, et al. (2007) Relationships between oxidative stress response and virulence in *Enterococcus faecalis*. *J Mol Microbiol Biotechnol* 13: 140–146.
42. Rince A, Giard JC, Pichereau V, Flahaut S, Auffray Y (2001) Identification and characterization of *gsp65*, an organic hydroperoxide resistance (*ohr*) gene encoding a general stress protein in *Enterococcus faecalis*. *J Bacteriol* 183: 1482–1488.
43. Mereghetti L, Sitkiewicz I, Green NM, Musser JM (2008) Extensive adaptive changes occur in the transcriptome of *Streptococcus agalactiae* (group B *Streptococcus*) in response to incubation with human blood. *PLoS ONE* 3: e3143.
44. Jordan S, Hutchings MI, Mascher T (2008) Cell envelope stress response in Gram-positive bacteria. *FEMS Microbiol Rev* 32: 107–146.
45. Rowley G, Spector M, Kormanec J, Roberts M (2006) Pushing the envelope: extracytoplasmic stress responses in bacterial pathogens. *Nat Rev Microbiol* 4: 383–394.
46. Solheim M, Aakra A, Vebo H, Snipen L, Nes IF (2007) Transcriptional responses of *Enterococcus faecalis* V583 to bovine bile and sodium dodecyl sulfate. *Appl Environ Microbiol* 73: 5767–5774.
47. Brunskill EW, Bayles KW (1996) Identification of LytSR-regulated genes from *Staphylococcus aureus*. *J Bacteriol* 178: 5810–5812.
48. Xu Y, Singh KV, Qin X, Murray BE, Weinstock GM (2000) Analysis of a gene cluster of *Enterococcus faecalis* involved in polysaccharide biosynthesis. *Infect Immun* 68: 815–823.
49. Teng F, Singh KV, Bourgogne A, Zeng J, Murray BE (2009) Further characterization of the *epa* gene cluster and Epa polysaccharides of *Enterococcus faecalis*. *Infect Immun* 77: 3759–3767.
50. Singh KV, Lewis RJ, Murray BE (2009) Importance of the *epa* Locus of *Enterococcus faecalis* OG1RF in a Mouse Model of Ascending Urinary Tract Infection. *J Infect Dis* 200: 417–420.
51. Huebner J, Wang Y, Krueger WA, Madoff LC, Martirosian G, et al. (1999) Isolation and chemical characterization of a capsular polysaccharide antigen shared by clinical isolates of *Enterococcus faecalis* and vancomycin-resistant *Enterococcus faecium*. *Infect Immun* 67: 1213–1219.
52. Fabretti F, Theilacker C, Baldassarri L, Kaczynski Z, Kropec A, et al. (2006) Alanine esters of enterococcal lipoteichoic acid play a role in biofilm formation and resistance to antimicrobial peptides. *Infect Immun* 74: 4164–4171.
53. Le Breton Y, Pichereau V, Sauvageot N, Auffray Y, Rince A (2005) Maltose utilization in *Enterococcus faecalis*. *J Appl Microbiol* 98: 806–813.
54. Hufnagel M, Koch S, Creti R, Baldassarri L, Huebner J (2004) A putative sugar-binding transcriptional regulator in a novel gene locus in *Enterococcus faecalis* contributes to production of biofilm and prolonged bacteremia in mice. *J Infect Dis* 189: 420–430.
55. Mohamed JA, Teng F, Nallapareddy SR, Murray BE (2006) Pleiotropic effects of 2 *Enterococcus faecalis* *sagA*-like genes, *salA* and *salB*, which encode proteins that are antigenic during human infection, on biofilm formation and binding to collagen type I and fibronectin. *J Infect Dis* 193: 231–240.
56. Hutchings MI, Palmer T, Harrington DJ, Sutcliffe IC (2009) Lipoprotein biogenesis in Gram-positive bacteria: knowing when to hold 'em, knowing when to fold 'em. *Trends Microbiol* 17: 13–21.
57. Hechard Y, Pelletier C, Cenatiempo Y, Frere J (2001) Analysis of sigma(54)-dependent genes in *Enterococcus faecalis*: a mannose PTS permease (EIP^{Mann}) is involved in sensitivity to a bacteriocin, mesentericin Y105. *Microbiology* 147: 1575–1580.
58. Deutscher J, Francke C, Postma PW (2006) How phosphotransferase system-related protein phosphorylation regulates carbohydrate metabolism in bacteria. *Microbiol Mol Biol Rev* 70: 939–1031.
59. Corbin BD, Seeley EH, Raab A, Feldmann J, Miller MR, et al. (2008) Metal chelation and inhibition of bacterial growth in tissue abscesses. *Science* 319: 962–965.
60. Torres V, Stauff LD, Pishchany G, Bezbradica JS, Gordy LE, et al. (2007) A *Staphylococcus aureus* regulatory system that responds to host heme and modulates virulence. *Cell Host & Microbe* 1: 109–119.
61. Andrews SC, Robinson AK, Rodriguez-Quinones F (2003) Bacterial iron homeostasis. *FEMS Microbiol Rev* 27: 215–237.
62. MacLeod RA (1951) Further mineral requirements of *Streptococcus faecalis*. *J Bacteriol* 62: 337–345.
63. MacLeod RA, Snell EE (1947) Some mineral requirements of the lactic acid bacteria. *J Biol Chem* 170: 351–365.
64. Low YL, Jakubovics NS, Flatman JC, Jenkinson HF, Smith AW (2003) Manganese-dependent regulation of the endocarditis-associated virulence factor EfaA of *Enterococcus faecalis*. *J Med Microbiol* 52: 113–119.
65. Bourgogne A, Garsin DA, Qin X, Singh KV, Sillanpaa J, et al. (2008) Large scale variation in *Enterococcus faecalis* illustrated by the genome analysis of strain OG1RF. *Genome Biol* 9: R110.
66. Murray BE, Singh KV, Ross RP, Heath JD, Dunny GM, et al. (1993) Generation of restriction map of *Enterococcus faecalis* OG1 and investigation of growth requirements and regions encoding biosynthetic function. *J Bacteriol* 175: 5216–5223.
67. Graham MR, Virtaneva K, Porcella SF, Barry WT, Gowen BB, et al. (2005) Group A *Streptococcus* transcriptome dynamics during growth in human blood reveals bacterial adaptive and survival strategies. *Am J Pathol* 166: 455–465.
68. Benachour A, Muller C, Dabrowski-Coton M, Le Breton Y, Giard JC, et al. (2005) The *Enterococcus faecalis* *sigV* protein is an extracytoplasmic function sigma factor contributing to survival following heat, acid, and ethanol treatments. *J Bacteriol* 187: 1022–1035.
69. Hancock LE, Perego M (2004) Systematic inactivation and phenotypic characterization of two-component signal transduction systems of *Enterococcus faecalis* V583. *J Bacteriol* 186: 7951–7958.
70. Teng F, Wang L, Singh KV, Murray BE, Weinstock GM (2002) Involvement of PhoP-PhoS homologs in *Enterococcus faecalis* virulence. *Infect Immun* 70: 1991–1996.
71. Brinster S, Lamberet G, Staels B, Trieu-Cuot P, Gruss A, et al. (2009) Type II fatty acid synthesis is not a suitable antibiotic target for Gram-positive pathogens. *Nature* 458: 83–86.
72. Orihuela CJ, Radin JN, Sublett JE, Gao G, Kaushal D, et al. (2004) Microarray analysis of pneumococcal gene expression during invasive disease. *Infect Immun* 72: 5582–5596.
73. Walecka E, Bania J, Dworniczek E, Ugorski M (2009) Genotypic characterization of hospital *Enterococcus faecalis* strains using multiple-locus variable-number tandem-repeat analysis. *Lett Appl Microbiol* 49: 79–84.
74. Guzman CA, Pruzzo C, LiPira G, Calegari L (1989) Role of adherence in pathogenesis of *Enterococcus faecalis* urinary tract infection and endocarditis. *Infect Immun* 57: 1834–1838.
75. Guzman CA, Pruzzo C, Plate M, Guardati MC, Calegari L (1991) Serum dependent expression of *Enterococcus faecalis* adhesins involved in the colonization of heart cells. *Microb Pathog* 11: 399–409.
76. Mohamed JA, Huang W, Nallapareddy SR, Teng F, Murray BE (2004) Influence of origin of isolates, especially endocarditis isolates, and various genes on biofilm formation by *Enterococcus faecalis*. *Infect Immun* 72: 3658–3663.
77. Weiser JN, Austrian R, Sreenivasan PK, Masure HR (1994) Phase variation in pneumococcal opacity: relationship between colonial morphology and nasopharyngeal colonization. *Infect Immun* 62: 2582–2589.
78. Nallapareddy SR, Murray BE (2008) Role played by serum, a biological cue, in the adherence of *Enterococcus faecalis* to extracellular matrix proteins, collagen, fibrinogen, and fibronectin. *J Infect Dis* 197: 1728–1736.

79. Rice KC, Nelson JB, Patton TG, Yang SJ, Bayles KW (2005) Acetic acid induces expression of the *Staphylococcus aureus* *cidABC* and *lrgAB* murein hydrolase regulator operons. *J Bacteriol* 187: 813–821.
80. Groicher KH, Firek BA, Fujimoto DF, Bayles KW (2000) The *Staphylococcus aureus* *lrgAB* operon modulates murein hydrolase activity and penicillin tolerance. *J Bacteriol* 182: 1794–1801.
81. Posey JE, Gherardini FC (2000) Lack of a role for iron in the Lyme disease pathogen. *Science* 288: 1651–1653.
82. Singh KV, Coque TM, Weinstock GM, Murray BE (1998) In vivo testing of an *Enterococcus faecalis* *efaA* mutant and use of *efaA* homologs for species identification. *FEMS Immunol Med Microbiol* 21: 323–331.
83. Duwat P, Sourice S, Cesselin B, Lamberet G, Vido K, et al. (2001) Respiration capacity of the fermenting bacterium *Lactococcus lactis* and its positive effects on growth and survival. *J Bacteriol* 183: 4509–4516.
84. Brooijmans R, Smit B, Santos F, van Riel J, de Vos WM, et al. (2009) Heme and menaquinone induced electron transport in lactic acid bacteria. *Microb Cell Fact* 8: 28.
85. Yamamoto Y, Poyart C, Trieu-Cuot P, Lamberet G, Gruss A, et al. (2005) Respiration metabolism of Group B *Streptococcus* is activated by environmental haem and quinone and contributes to virulence. *Mol Microbiol* 56: 525–534.
86. Shankar N, Baghdayan AS, Gilmore MS (2002) Modulation of virulence within a pathogenicity island in vancomycin-resistant *Enterococcus faecalis*. *Nature* 417: 746–750.
87. Chakraborty T, Leimeister-Wachter M, Domann E, Hartl M, Goebel W, et al. (1992) Coordinate regulation of virulence genes in *Listeria monocytogenes* requires the product of the *prfA* gene. *J Bacteriol* 174: 568–574.
88. Recsei P, Kreiswirth B, O'Reilly M, Schlievert P, Gruss A, et al. (1986) Regulation of exoprotein gene expression in *Staphylococcus aureus* by *agr*. *Mol Genet* 202: 58–61.
89. Graham MR, Smoot LM, Migliaccio CA, Virtaneva K, Sturdevant DE, et al. (2002) Virulence control in group A *Streptococcus* by a two-component gene regulatory system: global expression profiling and in vivo infection modeling. *Proc Natl Acad Sci U S A* 99: 13855–13860.
90. Qin X, Singh KV, Weinstock GM, Murray BE (2001) Characterization of *fsr*, a regulator controlling expression of gelatinase and serine protease in *Enterococcus faecalis* OG1RF. *J Bacteriol* 183: 3372–3382.
91. Bourgogne A, Hilsenbeck SG, Dunny GM, Murray BE (2006) Comparison of OG1RF and an isogenic *fsrB* deletion mutant by transcriptional analysis: the Fsr system of *Enterococcus faecalis* is more than the activator of gelatinase and serine protease. *J Bacteriol* 188: 2875–2884.
92. Roberts JC, Singh KV, Okhuysen PC, Murray BE (2004) Molecular epidemiology of the *fsr* locus and of gelatinase production among different subsets of *Enterococcus faecalis* isolates. *J Clin Microbiol* 42: 2317–2320.
93. Portenier I, Waltimo TMT, Haapasalo M (2003) *Enterococcus faecalis* - the root canal survivor and 'star' in post-treatment disease. *Endodontic Topics* 6: 135–159.
94. Aakra A, Vebø H, Snipen L, Hirt H, Aastveit A, et al. (2005) Transcriptional response of *Enterococcus faecalis* V583 to erythromycin. *Antimicrob Agents Chemother* 49: 2246–2259.
95. Diggle PJ, Liang K, Zeger SL (1994) Analysis of Longitudinal Data. Oxford: Clarendon Press. 253 p.
96. Pfaffl MW (2001) A new mathematical model for relative quantification in real-time RT-PCR. *Nucleic Acids Res* 29: e45.
97. Jaeger KE, Ransac S, Dijkstra BW, Colson C, van Heuvel M, et al. (1994) Bacterial lipases. *FEMS Microbiol Rev* 15: 29–63.
98. von Wallbrunn A, Heipieper HJ, Meinhardt F (2002) Cis/trans isomerisation of unsaturated fatty acids in a cardiolipin synthase knock-out mutant of *Pseudomonas putida* P8. *Appl Microbiol Biotechnol* 60: 179–185.

PAPER III

Comparative genomic analysis of pathogenic and probiotic *Enterococcus faecalis* isolates, and their transcriptional responses to growth in human urine.

Heidi C. Vebo^{1*}, Margrete Solheim¹, Lars Snipen², Ingolf F. Nes¹, and Dag A. Brede¹

¹ Laboratory of Microbial Gene Technology and Food Microbiology, ² Section for Biostatistics. Department of Chemistry, Biotechnology and Food Science, The Norwegian University of Life Sciences, N-1432 Ås, Norway

***Corresponding author:**

Heidi C. Vebo

The Norwegian University of Life Sciences

Laboratory of Microbial Gene Technology

P.O. Box 5003

N-1432 Ås, Norway

Phone: +47 64965912 Fax: +47 64941465 e-mail: heidi.vebo@umb.no

Abstract

Background:

Urinary tract infection (UTI) is the most common infection caused by enterococci, and *Enterococcus faecalis* accounts for the majority of enterococcal infections. Although a number of virulence related traits have been established, no comprehensive genomic or transcriptomic studies have been conducted to investigate how to distinguish pathogenic from non-pathogenic *E. faecalis* in their ability to cause UTI.

Methodology/Principal Findings:

In order to identify potential genetic traits or gene regulatory features that distinguish pathogenic from non-pathogenic *E. faecalis* with respect to UTI, we have performed comparative genomic analysis, and investigated growth capacity and transcriptome profiling in human urine *in vitro*.

Six strains of different origins were cultivated and all grew readily in human urine. The three strains chosen for transcriptional analysis showed an overall similar response with respect to energy and nitrogen metabolism, stress mechanism, cell envelope modifications, and trace metal acquisition. Our results suggest that citrate and aspartate are significant for growth of *E. faecalis* in human urine, and manganese appear to be a limiting factor. The majority of virulence factors were either not differentially regulated or down-regulated. Notably, the *fsrABC* operon important for virulence in *E. faecalis* was significantly up-regulated in the pathogenic MMH594 strain.

Conclusions/Significance:

Strains from different origins have similar capacity to grow in human urine. The overall similar transcriptional responses imply that the pathogenic potential of a certain *E. faecalis* strain is to a great extent determined by presence of fitness and virulence factors, rather than the level of expression of such traits. However, the significant differential expression of the *fsr*-system between MMH594 and OG1RF implies that the amount produced of certain virulence factor(s) can contribute to differences in virulence potential between strains.

Introduction

Once considered as harmless commensals of the intestinal tract, enterococci now rank among the leading causes of infections among hospital patients [1,2]. *Enterococcus faecalis* is among the most prevalent agents isolated from nosocomial urinary tract infections (UTIs), and is a common cause of chronic and recurrent UTIs, especially those associated with structural abnormalities and medical devices, such as urinary catheters [3]. The ability of *E. faecalis* to cause infection has been linked to inherent enterococcal traits, enabling the bacterium to tolerate harsh and diverse environments. In addition, several factors that may contribute to enterococcal virulence have been characterized (reviewed in [4]), and the role of these factors in pathogenicity have been further established in various animal models [5-8] and cultured cell lines [9,10]. However, a widespread distribution of putative virulence determinants in enterococcal isolates independent of origin has been reported [11-16], and to date, no single virulence factor has been demonstrated to be essential for enterococcal infections. The ability of *E. faecalis* to cause infection is therefore likely to involve an orchestrated interplay between the regulation of these putative virulence factors and various genetic determinants that govern adaptation of the bacterial cell physiology during the infection process. Cultivation in urine partly mimics the urinary tract environment, and identification of differentially expressed genes *in vitro* may therefore represent a potential means to identify novel fitness factors required for this particular ecological niche.

Shepard and Gilmore previously examined the effect of growth in urine on the expression of known and suspected enterococcal virulence factors by quantitative real-time PCR [17], and significant changes in *E. faecalis* virulence-associated gene expression were observed in response to the biological cues present in urine, compared to laboratory medium-growth. Furthermore, studies of other pathogens causing UTI have reported responses involving iron acquisition systems and genes involved in sugar and amino acid metabolism [18,19], which may indicate that bacteria suffer from glucose and iron limitation during growth in human urine.

In this report, we compare the global expression profiles of three *E. faecalis* strains during growth in human urine *in vitro*. The three strains were chosen based on their origins; the

Symbioflor 1 strain, included in a commercial probiotic product used for more than fifty years without any reports of infection [20], the hospital outbreak strain MMH594 holding most known virulence genes in its genetic repertoire [21,22], and finally the laboratory strain OG1RF which harbors some important virulence traits like *fsr* and *epa*, but is devoid of mobile genetic elements (MGEs) [23,24]. This latter strain is however capable of causing infection in *e.g.* mice [23,25], and has been extensively used as a model organism to investigate virulence ([4] and references therein) . The aim of this work was to gain insight into genetic factors that make *E. faecalis* such a potent cause of human UTI. The study was designed to identify traits that distinguish pathogenic from non-pathogenic *E. faecalis*. Identification of such traits may ultimately contribute to development of strategies for prevention and treatment of *E. faecalis* UTI.

Results and Discussion

Growth capacity of different *E. faecalis* strains in urine and 2xYT

Escherichia coli associated with UTI normally grow well in urine, while non-uropathogenic strains do not [26]. To examine whether this also could be true for *E. faecalis*, six strains of nosocomial, UTI, commensal or probiotic origin were cultivated in urine and colony forming unit (CFU) counts performed (Fig. 1). Only minor differences in growth capacity were observed between the various isolates, with generation times of around 48 minutes. MMH594 and V583 reached a slightly higher final cell density ($\sim 2.0 \times 10^8$ CFU/ml) compared to OG1RF and Symbioflor 1 ($\sim 1.2 \times 10^8$ CFU/ml), and even more so compared to Baby isolate 62 and 179Vet ($\sim 6.5 \times 10^7$ CFU/ml). These observations are consistent with a recent study by Carlos *et al.* [27], where strains from diverse origins, such as food and clinical strains, did not grow significantly different in urine. Furthermore, the growth capacity of MMH594 observed in the present study was in agreement with previous reports [17].

Comparative genomic hybridization

Since the initial growth experiments did not reveal any strains with a distinctively enhanced or reduced growth capacity in urine, two pathogenic strains MMH594 and OG1RF, and the

probiotic strain Symbioflor 1 were selected for further investigation by comparative genome hybridization and transcriptional analysis.

Comparative genome hybridization was performed to assess the variation in gene content between the three test strains, using V583 as reference (Fig. 2, Fig. 3 and Table S1). A total of 2284 genes were classified as present in all the strains tested. Not surprisingly, the clinical bacteremia isolate MMH594 showed the highest similarity to the reference strain V583 (94.7% genes in common). In addition, the CGH data confirmed the presence of all the ten additional enterococcal pathogenicity island (PAI) genes, covering a 17-kb deletion in V583, in MMH594 [22,28]. For the two other test strains the similarity to the reference strain was significantly lower, with 74.1 and 73.7% of V583 genes represented on the array classified as present in OG1RF and Symbioflor 1, respectively. The CGH analysis indicated that 12 PAI genes without apparent function in virulence were present in Symbioflor1, whereas three PAI genes were found in OG1RF (Fig. 3 and Table S1), but inspection of the published genome sequence [23] showed that these were false positive results. As expected, none of the additional PAI genes seemed to be present in the latter two strains. Altogether, MMH594 contains 596 genes that appear to be divergent in OG1RF and Symbioflor 1. Major variations in the presence of all the previously defined mobile genetic elements (MGEs) were observed between the three test strains. Except for *phage01* and *vanB*, all the MGEs seemed to be present in MMH594. *phage02* appear to be part of the *E. faecalis* core genome, but none of the other elements were found in OG1RF. This observation is consistent with the genome sequence available for OG1RF [23]. Symbioflor 1 appeared to contain certain genes/modules from *phage06*, but not the entire element. The rest of the MGEs were divergent by CGH in Symbioflor 1, which is consistent with previous reports [20]. Notably, Symbioflor 1 contains two major deletions in proximity to the *vanB* associated island and the *efaB5* element (Fig. 3 and Table S1). The latter deletion extends in the 5' direction of *efaB5* to EF1811 including the *fsr-gelE-sprE* virulence locus.

The number of on-going *E. faecalis* sequencing projects is rapidly increasing, and the recent publication of the OG1RF genome revealed a total of 2474 genes that were common for V583 and OG1RF. This number was based on the homology at the DNA level with the ORFs described for V583, with a BLASTN cutoff e-value of 1×10^{-5} [23]. The number of common

genes revealed by sequencing is slightly higher than the number of present genes obtained by CGH in this present study. When comparing the list of common genes between OG1RF and V583 obtained by Bourgogne *et al.* to the data presented here, a total of 168 genes deviate in their classification as present and divergent in OG1RF. In addition, 20 of 2474 genes identified as common to V583 and OG1RF by Bourgogne *et al.* were not represented on the array used in the present study. Moreover, the numbers reported by Bourgogne *et al.* did not include blasting with any of the plasmid ORFs described for V583, while 47 plasmid genes were classified as present in OG1RF by CGH. An interactive Genewiz map [29] of OG1RF CGH and BlastN (Genbank [ABPI00000000](#)) analysis compared with V583 is accessible at; <http://ws.cbs.dtu.dk/cgi-bin/gwBrowser-0.91/edit.cgi?hexkey=6561d07e713b77fe75aa3403798e36c1>.

Transcriptional analysis

A rich laboratory medium (2xYT) was used as the reference culture medium since it is considered to contain a minimum of infection relevant biological cues [17]. The growth capacity in urine was compared to that in the 2xYT medium by CFU counts (Fig. S1). We found that growth in urine was slightly slower, and the cell density obtained was about one log lower than in 2xYT. For the transcriptional analysis, the three strains were grown in 2xYT to a cell density $\sim 1 \times 10^7$ before exposure to either pre-warmed urine or 2xYT (control). Samples were collected after 5 (t_5) and 30 (t_{30}) minutes growth. The obtained \log_2 -ratios and q -values found in the three strains during growth in urine compared to 2xYT are listed in Table S1.

Growth in urine vs. 2xYT triggers global transcriptional changes for both pathogenic and probiotic *E. faecalis*

The microarray results revealed changed expression in most functional gene categories for all three strains. At t_5 , 713 genes were differentially expressed in MMH594, 735 in OG1RF and 730 in Symbioflor 1. 344 of these regulated genes were common for all three strains (Fig. 4A). At t_{30} , the number of regulated genes increased dramatically to 1212 genes in MMH594 and 979 in Symbioflor 1. However, in OG1RF the number of regulated genes decreased to 574 after 30 minutes growth in urine. It is possible that the reduced number of

regulated genes in OG1RF at t_{30} reflects a more rapid adjustment to the new growth environment, which potentially can be advantageous for the establishment of an infection. A total of 378 differentially expressed genes were common for MMH594, OG1RF and Symbioflor 1 at t_{30} (Fig. 4B). Of the 596 genes that appeared unique to MMH594, 153 were differentially expressed at one or both time points during growth in urine. None of the genes unique to OG1RF or Symbioflor 1 were differentially expressed.

The heat map in Figure 5 presents an overview of the regulated genes within each functional category for the three strains. The figure shows large similarities in the expression pattern between the strains, but also reveals an individual transcriptional signature for each strain. The similarities were particularly evident by the down-regulation of several genes involved in transcription and cell envelope functions, and in the up-regulation of several genes involved in fatty acid and phospholipid metabolism, energy metabolism, amino acid biosynthesis, and genes with a regulatory function. For OG1RF, it seems that the down-regulation of several genes involved in protein synthesis occurs at t_5 , while most of the down-regulation occurs at t_{30} in the two other strains. We also found several genes encoding proteins with a transport and/or binding function that are regulated in only two of the three strains, however, without any two strains showing a more similar pattern to each other. An overview of the number of regulated genes within each functional category is given in Figure S2.

Transcription of metabolic pathways during exponential growth in urine.

As described in Figure 1, all the investigated *E. faecalis* strains were capable of rapid growth (doubling time of 48.6 ± 3.7 min) in urine, with most strains reaching final cell densities of $\sim 1 \times 10^8$ cfu/mL (mean growth $1.3 \times 10^8 \pm 0.6 \times 10^8$). Prior to the current study, no comprehensive investigation regarding which substrates or metabolic processes that confer growth of *E. faecalis* in urine existed. The transcriptome data (Table S1) was thus examined to identify metabolic pathways that showed specific responses during exponential growth in urine.

With respect to carbon metabolism the genes encoding the main glucose uptake-system, mannose phosphoenolpyruvate phosphotransferase (PTS) *mptBACD* (EF0019-22) [30] were down-regulated in all three strains. This is consistent with a recent metabolomic investigation which showed that urine from healthy adults contains glucose concentrations in the range of 0.2-0.6 mM [31]. Such concentrations of glucose is below the threshold for release of carbon catabolite repression (CCR), and the cells thus initiate use of less preferred carbon and energy sources [32]. This implied that substrates besides glucose might play a role for growth of *E. faecalis* in urine. However, of the loci known to be subject to catabolite control protein A (CcpA) mediated CCR, only the genes encoding citrate metabolism (EF3322-15) [33] were positively modulated in MMH594 and OG1RF at both time points and at t_5 in Symbioflor1. At t_{30} EF3322-15 only showed a slightly (not statistically significant) enhanced expression in Symbioflor 1. The content of citrate in human urine is in the range of 1-2 mM [34], which suggests that citrate metabolism is important for *E. faecalis* during growth in urine.

PTS systems facilitate uptake of diverse sugars in *E. faecalis*. Two genes encoding PTS systems with unknown substrates (EF2438 and EF2603) showed an enhanced expression in all three strains. Both EF2438 and EF2603 are part of the *E. faecalis* core genome, and might be important for growth of *E. faecalis* in urine. Furthermore, two operons encoding a sucrose uptake PTS-system (EF1602-01) and sucrose metabolism (EF1603-04) showed consistent up-regulation in all three strains. Dietary sucrose is normally degraded in the intestinal lumen and absorbed as glucose and fructose, but a previous study has shown that even healthy individuals have μM sucrose content in their urine [35]. Moreover, the sugar content in urine increases with high sugar diet. Once sucrose is present in the bloodstream it is not metabolized further, but removed from the blood via the renal capillaries and excreted into the urine, reaching concentrations of 70 to 200 μM [35]. Interestingly, EF1603-04 knock-out mutants show reduced virulence in a *Caenorhabditis elegans* infection model [36,37]. All three strains showed elevated expression of the major facilitator family transporter (EF0082) proposed to function in import of phosphorylated sugars [38] and glycerol [39], which implies that such substrates might contribute to growth in urine.

Transcriptome analysis conducted on an *E. coli* asymptomatic bacteriuria strain revealed an important role of amino sugar and amino acids present in urine as growth substrates [40]. The

transcription of *nagB* (EF0466) and *nagA-1* (EF1317) involved in N-acetyl glucosamine metabolism was highly elevated, implying that these substrates were utilized by *E. faecalis* during growth in urine. A massive down-regulation of *glmS* (EF2151), which is responsible for conversion of fructose-6P into glucosamine-6P using glutamine as a nitrogen source, could signify glutamine constraints.

Growth in urine also had an impact on pyruvate metabolic pathways and certain changes were strain specific. For OG1RF and Symbioflor 1, we observed increased expression of L-lactate dehydrogenase (*ldh-1*; EF0255), whereas expression of *adhE* (EF0900), involved in ethanol formation was reduced. The *pflAB* (EF1612 and EF1613) genes responsible for formate formation were reduced in MMH594 and Symbioflor 1. In all strains the *lutABC* operon (EF1108-1110), involved in metabolism of L-lactate like substrates was up-regulated. The pyruvate dehydrogenase complex gene-cluster *pdhAB*, *aceF* and *lpdA* (EF1353-56) involved in acetyl-CoA biosynthesis showed consistent up-regulation in all three strains. Moreover, the *ackA* gene (EF1983) responsible for conversion of acetyl-phosphate to acetate and ATP was significantly down-regulated, perhaps as a consequence of increased acetate production due to elevated activity of the citrate metabolism (EF3322-15) [33]. It is thus conceivable that the increased acetyl-CoA formation serves to supply either the FASII biosynthesis, or the citrate metabolism.

Transport and biosynthesis

Compared to the rich 2xYT medium the growth rates were significantly lower in urine, and moreover, the growth halted one order of magnitude below that in 2xYT (Fig. S1). For *E. coli* it has been demonstrated that growth in urine is restricted by availability of one specific cofactor, namely iron [40]. We were thus interested to see whether the transcriptional responses with respect to transport and biosynthesis processes in *E. faecalis*, could reveal candidate nutrients or co-factors whose availability restrict growth of *E. faecalis* in urine.

Human urine contains significant amounts of creatine, creatinine, and glycine, while other amino acids like histidine, glutamine, methionine, proline, glutamate, arginine and branched chain amino acids (bcaa) are present at lower concentrations [41]. The CGH-results indicate

that MMH594 and Symbioflor 1 have similar requirements for amino acids as OG1RF, which was shown to be auxotrophic for amino acids like histidine, isoleucine, methionine, and tryptophan [24]. Also, some *E. faecalis* strains require arginine, glutamate, glycine, leucine, or valine [24], and are capable of utilizing certain amino acids as energy and carbon source [42,43]. However, transcription of the genes encoding catabolism of arginine (EF0104-7 and EF0108) and serine (EF0097-100) was significantly reduced, indicating a shift towards protein synthesis rather than energy metabolism.

According to our data, the transcription of several genes encoding oligo-peptide ABC-transporters (EF0907, EF0909-12 and EF3110-06) was enhanced at t_{30} , and the transcription of three amino acid permease genes (EF0635, EF0929 and EF2377) and two operons encoding amino acid transporters (EF0247-46 and EF0761-60) was reduced in all three strains (Table S1). These observations indicate that *E. faecalis* meets its demand for certain amino acids by acquiring oligo-peptides during growth in urine. However, the *gln*-operon encoding glutamine/glutamate transport system (EF1120-17) [44] were up-regulated in all strains at both time points, suggesting that glutamate/glutamine from urine were utilized. This was further supported by the observed reduced expression of the glutamine synthase operon *glnRA* (EF2160-59) and glutamate synthase *gltA* (EF2560). On the contrary, the increased expression of *cysK* (EF1584) implies that cysteine is scarce in urine, which also is in accordance to the metabolomic analysis of human urine [41].

An operon comprising a putative amino acid ABC transporter (EF0893-92) and a putative aspartate aminotransferase (EF0891) was highly up-regulated in all strains. The latter gene is predicted to facilitate the conversion of aspartate and alpha-ketoglutarate to oxaloacetate and glutamate, which may also in turn explain the down-regulation of the above mentioned *gltA*. Furthermore, the transcription of a gene encoding methionine synthase (EF0395) was enhanced in all three strains. These results are consistent with the metabolomic analysis of human urine which showed that aspartate is 5-fold more abundant than methionine [41]. These observations imply that aspartate might serve a key role for nitrogen metabolism of *E. faecalis* in urine.

We observed an enhanced transcription of an isochorismatase gene (EF3192) in all strains at t_{30} , and also a second isochorismatase gene (EF3090) in MMH594. This coincided with a down-regulation of the EF1561-68 operon in MMH594 and Symbioflor 1, responsible for biosynthesis of chorismate, which is a precursor of aromatic amino acids, folate and quinones. In agreement with this observation, we also noticed a reduced transcription of several genes responsible for biosynthesis of cofactors, prosthetic groups, and carriers including menaquinone and ubiquinone (EF3255-54 and EF3260). Interestingly, the genes encoding chorismate biosynthesis were not down-regulated in OG1RF. Moreover, another gene cluster encoding menaquinone biosynthesis (EF0446-50) showed a reduced transcription in MMH594 and Symbioflor 1, while an enhanced transcription in OG1RF, although the regulation of some of the genes were not statistically significant.

Thus, it appears that *E. faecalis* scavenge available peptides and amino acids, which in turn are sequentially hydrolyzed and transaminated in order to fuel the pool of depleted amino acids.

Urinary tract pathogenic bacteria like *E. coli* (UPEC), have pathogenic islands dedicated to acquisition of limited nutrients and biometals [45]. Manganese is one such factor which is essential for the fermentative metabolism of lactic acid bacteria (LAB) [46,47]. The up-regulation of the main manganese scavenging mechanism encoded by *efaCBA* (EF2074-76), accompanied by two other genes (EF1057 and EF1901) encoding Mn^{2+}/Fe^{2+} transporters in all strains at both time points is a clear indication that *E. faecalis* scavenged manganese. The content of manganese in human urine is in the nano molar range [48], while the optimal concentration for *E. faecalis* is in the micro molar range [49]. Thus manganese may be restrictive for the growth of *E. faecalis*. This in turn can affect the virulence of the bacterium and *efaCBA* has indeed been shown to be implicated in virulence [50]. Notably, for MMH594 a potential auxiliary Mn-uptake system (EF 0575-78) [51], located within the PAI also showed highly elevated expression, indicating that PAI harboring strains might be better equipped to cope with manganese deplete environments.

Furthermore, our experiments revealed an enhanced expression of several other genes involved in iron transport; the *feoAB* (EF0475-76) and *ceuBCD* and *fatB* (EF3085-82) operons were up-regulated in all strains at t_{30} . Another gene involved in iron transport, *feuA*

(EF0188) was down-regulated in all strains at t_5 , but was up-regulated in MMH594 and OG1RF at t_{30} . Interestingly, in OG1RF a third iron transport encoding operon (EF0191-93) was only up-regulated at t_{30} . Iron is one of the main limiting factors for *E. coli* growth in urine and the addition of iron to urine increased the maximum growth extensively [18,19]. LAB, on the other hand comprise one of the very few groups of bacteria for which iron is not an essential growth factor [47]. Even so, our data suggest a potentially important role of iron acquisition and metabolism during growth in urine.

Stress response of V583 towards exposure to urine

Proteomic analyses with systematic exposure to various stresses have previously identified six genes encoding general stress response proteins (GSPs) which were up-regulated in *E. faecalis* by a wide variety of environmental stimuli [52]. The enhanced expression of all the GSP-encoding genes at one or both time points in the present study indicates that the bacterium experienced a multitude of stress factors upon the encounter with urine. This impression was further substantiated by the significantly differential transcription of a large number of genes with a proven or predicted function in other stress responses in *E. faecalis* (Table 1 and S1).

The gene encoding Gsp62 (EF0770; hypothetical protein) was the only GSP which showed a significantly enhanced expression in all strain at both time points. The stress- and starvation inducible *gls24* operon (EF0076-81) was significantly up-regulated at both time points in OG1RF and Symbioflor 1, while partly up-regulated in MMH594. Inactivation of *gls24* and *glsB* (EF0079 and -80, respectively) has been reported to have a pleiotrophic effect on cell morphology and stress tolerance in *E. faecalis* [53]. A *gls24* disruption mutant has also been shown to be highly attenuated in animal infection models [54,55]. MMH594 contains two additional *gls24*-like genes within the PAI (EF0604 and PAIef0055). Both gene were up-regulated at t_{30} and might possibly contribute to the fitness of MHH594 during growth in urine.

An organic hydroperoxide resistance protein, *ohr* (Gsp65; EF0453) was up-regulated in MMH594 at t_5 , and in all three strains at t_{30} . An *ohr* mutant has previously been shown to be

less resistant to the oxidative stress generated by 20 mM Tertiary-Butylhydroperoxide, suggesting that Ohr may be implicated in oxidative stress resistance in *E. faecalis* [56]. Interestingly, the microarray data revealed differential expression of an arsenal of genes holding putative roles in oxidative stress response in *E. faecalis* (Table 1 and S1). Verneuil and co-workers previously reported the expression of *ahpCF* (EF2738 and -39), *sodA* (EF0463), *katA* (EF1597) and *gor* (EF3270) to be positively controlled by the transcriptional regulator *hypR* (EF2958) [57]. Indeed, *hypR* showed augmented expression in Symbioflor 1 at both t_5 and t_{30} . However, the lack of significant enhanced expression of *hypR* in the two remaining test strains, may suggest that these genes are under more complex regulation. In fact, *fur* (EF1525), *perR* (EF1585) and *zur* (EF2417), encoding other putative regulators of oxidative response in *E. faecalis* [58,59] were also up-regulated at t_{30} in MMH594, in all three strains and in Symbioflor 1, respectively. In addition, *ers* (EF0074), encoding another regulator involved in survival during oxidative stress [60] showed reduced transcription in OG1RF at t_5 and in Symbioflor 1 at both t_5 and t_{30} . The enhanced transcription of genes involved in oxidative stress response during exposure to urine is interesting. Especially in light of an observed adaptation to lethal challenges of H_2O_2 by pretreatment with sublethal concentrations of H_2O_2 [61], and a reported link between oxidative stress response and survival within macrophages in enterococci [60,62,63]. Furthermore, it has been demonstrated that purified lipoteichoic acids from *E. faecalis* induced proliferation and production of nitrous oxides and cytokines by a subpopulation of basal urothelial cells [64,65]. It is thus tempting to speculate that urine act as a cue to trigger oxidative stress-protection by *E. faecalis*, in order to render increased resistance against certain host defense mechanisms in the urinary tract.

Modifications to the cell envelope caused by growth in urine

When infecting a host, the integrity and composition of the cell envelope of the bacterium is important to avoid damage by the host defense systems [66,67]. In the case of *E. faecalis*, it has been demonstrated that important processes in the interaction with the host *e.g.* recognition by immune system mechanisms and innate immune evasion, involve specific cell envelope structures like lipoteichoic acids [68], and cell wall and capsular polysaccharide determinants [69,70].

During growth in urine, signs of adaptation to this new growth environment were evident for several genes important for the cell membrane composition and surface related structures (Table S1). We observed an immediate response to urine by the up-regulation of two gene clusters (EF0282-84 and EF2886-75) responsible for type II fatty acid biosynthesis (FASII) and isomerization of membrane phospholipids. Most of these genes were up-regulated in all strains at t_5 and t_{30} . Interestingly, these gene clusters have previously been shown to be up-regulated in response to growth in blood [71] and to exposure to the cell membrane detergent SDS [72]. Furthermore, the FASII genes were down-regulated in response to exposure to NaCl (Solheim *et al*, in preparation), bovine bile, and SDS and bovine bile in combination [72], indicating that several different external stressors triggers remodeling of the fatty acid composition in the cell membrane.

In addition to the FASII pathway, a regulation of three genes encoding lipases and two genes encoding cardiolipin synthetases further indicates both degradation and processing of fatty acids (Table S1). The cardiolipin synthetase gene (EF0631) was down-regulated in MMH594 and Symbioflor 1 at both t_5 and t_{30} , and in OG1RF at t_5 , whereas another cardiolipin synthetase gene (EF1608) was up-regulated at both time points in Symbioflor 1, at t_5 in OG1RF and t_{30} in MMH594. Of the lipase encoding genes, EF0169 was up-regulated in all strains at both time points, EF3191 was up-regulated in MMH594 and Symbioflor 1 at t_{30} , and EF1683 was only up-regulated in Symbioflor 1 at t_{30} . It is possible that the lipolytic activity is connected to a modulation of the FASII genes, as it recently was demonstrated that *E. faecalis* can utilize available fatty acids from the environment in their membrane biogenesis [73]. However, there are only trace amounts of free fatty acids in urine [34], and it is therefore more likely that the remodeling of the fatty acid composition in the cell membrane is a more general stress response in *E. faecalis*, while the lipases may play a more specialized role in virulence. A recent study by Walecka and co-workers revealed that a higher percentage of invasive *E. faecalis* isolates produce lipases compared to non-invasive isolates [74], indicating a central role for lipase activity during invasive infection. Notably, Symbioflor 1 showed a more enhanced expression of genes encoding lipases compared to the pathogenic strains.

The ability of *E. faecalis* to adhere to and develop biofilm is thought to be important for its potential to cause UTI and other infections [75]. In our experimental design, the cells were cultivated planktonically. We were thus interested in assessing whether genes implicated in

adherence or biofilm formation would be modulated by human urine. The gene encoding the maltose PTS system *malT* (EF0958) and the cognate operon *bopABCD/malPBMR* (EF0957-54), which are involved in biofilm formation [76,77], were partly up-regulated in OG1RF at t_5 . Another gene important for biofilm production and the initial attachment stage for binding to abiotic surfaces is a sortase A encoding gene, *srtA* (EF3056) [78,79]. This gene showed an enhanced expression in MMH594 at t_{30} and Symbioflor 1 at both time points. Interestingly, an *srtA* mutant showed a slightly attenuated virulence during UTI in mice [80]. However, among the genes encoding potential substrate proteins of SrtA, only EF2713 was up-regulated at t_5 in MMH594, whereas EF3314 showed an enhanced expression in Symbioflor 1 at both t_5 and t_{30} . This latter gene encodes a protein recently shown to be important for the pathogenicity of *E. faecalis* [81], and it is noteworthy that the only strain which showed an enhanced expression of this gene, was the probiotic strain.

Bacterial surface proteins are key players in host-pathogen interactions [80]. Therefore, the change of membrane bound proteins might alter the bacterium's potential of causing an infection. Regulation of several genes encoding proteins bound to the cell membrane or cell surface *i.e.* membrane proteins and lipoproteins was observed for all three strains (Table S1). Moreover, most microbial surface components recognizing adhesive matrix molecules (MSCRAMMs) and cell-wall anchor family proteins [82] including the endocarditis- and biofilm-associated pilus (*ebp*) [83,84] were either down-regulated, or not differential regulated (Table S1).

A gene encoding a chitin binding protein (EF0362) and one encoding a chitinase (EF0361) were up-regulated in all three strains at t_5 . The direct function for these genes in response to urine is not clear, however a homologous protein GbpA in *Vibrio cholerae* was shown to facilitate binding to the chitin monomer N-acetylglucosamine [85], a sugar residue found on the surface of epithelial cells [86-88], which line the cavities and surfaces of structures including the urinary tract. Hence, it is possible that biological cues in urine trigger the up-regulation of these genes as an initial step of adherence to uroepithelial cells. Interestingly, growth of *E. faecalis* V583 in blood triggered an even more enhanced transcription of these two genes [71], but a functional study of these genes would be required to elucidate any function related to enterococcal virulence.

Mohamed *et al.* [89] demonstrated that a knockout mutant of the secreted antigen *salB* (EF0394) in OG1RF showed reduced biofilm formation in BHI, but enhanced biofilm production in the presence of serum or fibronectin. They also showed that the *salB* mutant was able to bind to the extra cellular matrix (ECM) proteins collagen type I and fibronectin, whereas wild type OG1RF did not bind these ECM proteins [89]. Furthermore they showed that a *sala* (EF3060; secreted lipase) mutant also produced slightly less biofilm than wild type OG1RF, while binding to ECM was unaffected. During growth in urine *sala* was down-regulated in all strains at both time points, while *salB* was down-regulated in all strains at t_5 , and in MMH594 at t_{30} . Mohamed and co-workers [89] speculated that under certain conditions a down-regulation of *salB* would be sufficient to see similar effects as was seen for the *salB* mutant, thus it is possible that the expression of *salB* and possibly also *sala* is reduced in response to urine in order to promote colonization of the urinary tract.

At t_5 , a gene encoding the major autolysin of *E. faecalis*, *atla* (EF0799) was down-regulated in all three strains. An *atla* deletion mutant of OG1RF showed delayed biofilm formation, reduced attachment on plastic surfaces and longer chains than the wild type OG1RF [78,90]. Atla is also essential for DNA release and biofilm accumulation, which is needed for the development of a mature biofilm in *E. faecalis* [78,91]. MMH594 and Symbioflor 1 contain a second peptidoglycan hydrolase encoding gene *atlB* (EF0355), which have been shown to compensate for the absence of Atla in autolysis and cell separation [90]. *atlB* was down-regulated at t_{30} in MMH594, while not differentially expressed in Symbioflor 1. Two putative holin genes (EF0353-54) up-stream of *atlB* were also down-regulated in MMH594, however these two genes were absent in Symbioflor 1. The lowered expression of *atla* and *atlB* may also be connected to reduced cell wall synthesis indicated by down regulation of several genes responsible for peptidoglycan biosynthesis (Table S1), which again is consistent with the significantly lower growth rate in urine compared to 2xYT.

Most of the genes within a cluster responsible for the production of a serotype-determining exopolysaccharide (EF2198-2177; *epa*) [92,93] were down-regulated both at t_5 and t_{30} in the three strains. An OG1RF $\Delta epaB$ mutant has previously been reported to show reduced virulence in mice [94], higher susceptibility to phagocytic killing [70], and decreased biofilm formation compared to the wild type [70,95]. Furthermore, Singh *et al.* recently showed that the *epaB* mutant was less competitive compared to the wild type in a model of UTI in mouse [96]. However, it is possible that the exopolysaccharide production is body-site dependent,

and could be more pronounced in *E. faecalis* that have reached the glomerular basement membrane in kidneys, which is a preferred site for *E. faecalis* colonization [96,97].

The serotype 2 capsular polysaccharide (*cps*) [98], which constitutes an important virulence factor that enables *E. faecalis* to evade phagocytic killing, by masking the lipoteichoic acids [69], is absent in both OG1RF and Symbioflor 1 (Fig. 3 and Table S1). Intriguingly, the *cps* gene cluster (EF2495-85) was down-regulated in MMH594 at t_{30} , which is similar to the response observed in V583 growing in blood [71]. It is tempting to speculate that a basal capsular polysaccharide production could be sufficient to protect *E. faecalis* from complement-mediated opsonophagocytosis, especially in infected tissues where micro-colonies or biofilm develop.

Glycosyl transferase family proteins are involved in formation of a number of cell surface structures including glycolipids, glycoproteins and polysaccharides [99]. Apart from the *epa* and *cps* loci, five genes encoding glycosyl transferases were also down-regulated in one or more strains in response to growth in urine. Neighboring the *epa* cluster, two glycosyl transferases (EF2170 and EF2167) proposed as potential virulence factors [100], together with the EF2172-66 genes, comprise a three operon locus possibly connected to lipopolysaccharide production. This gene cluster is present in MMH594 only (Fig. 3 and Table S1), and most genes within this locus were down-regulated at both t_5 and t_{30} . The gene encoding the glycosyl transferase *bgsA* (EF2891), responsible for the biosynthesis of the major cell membrane glycolipid in *E. faecalis*, was down-regulated in OG1RF at t_{30} . A *bgsA* mutant showed a reduced ability to accumulate biofilm, reduced virulence in a mouse bacteremia model and reduced ability to adhere to colonic epithelial cells [101]. The *bgsA* mutant also displayed a reduced cell surface hydrophobicity compared to the wild type, an effect which may also contribute to the impaired biofilm production [101]. The neighboring gene, EF2890, also encodes a glycosyl transferase that was down-regulated in OG1RF and MMH594 at t_{30} . The function of EF2890 remains to be elucidated, but it could possibly be connected to that of *bgsA*. A putative n-acetylglucosamine glycosyl transferase-encoding gene (EF0887) implicated in peptidoglycan-biosynthesis was up-regulated in all three strains at t_{30} , while down-regulated at t_5 in Symbioflor and OG1RF.

In sum, the human urine milieu appears to instigate a drastically altered composition of the cell envelope and cell surface structures, some of which might be advantageous or required for establishment of *E. faecalis* UTI.

Virulence traits and Regulatory genes

A number of genetic traits have been identified to contribute to virulence in *E. faecalis* [5,6,8,25,55,70,98,102,103]. The expression of selected virulence genes in MMH594 during growth in urine have previously been examined by real-time quantitative PCR (QPCR) [17]. More recently, a new QPCR study of the expression in several strains including MMH594 during growth in urine was published [104]. The two studies show some differences in gene expression in MMH594, e.g. of a gene encoding the enterococcal surface protein Esp (PAIef0056). Shepard and Gilmore found an enhanced expression of *esp*, while Carlos *et al.* found a reduced expression of the same gene. In the present study, we found that the *esp* gene was not significantly differentially expressed. Indeed, QPCR appear to be more sensitive and have a broader detection range than microarray, but the deviating results still seem to imply a problem when comparing these types of experiments. Shepard and Gilmore reported a growth-phase dependent difference in the expression of the virulence genes tested. Hence, the differences observed between these three similar experiments are most likely due to the different methods used for cultivation. Our aim was to investigate the immediate effect on actively growing *E. faecalis* cells upon the first encounter of urine. We revealed a significant impact on the transcription of a number of virulence related traits connected to stress, co-factor acquisition, and cell surface structures (described above), and a summary of these genes can be found in Table 2.

The *fsr* quorum sensing system has been shown to coordinate expression of the virulence factors *gelE* (encoding a gelatinase) and *sprE* (encoding a serine protease) during infection of *C. elegans* and in mouse preitonitis models [25,103], and several other genes were differentially expressed in wild type OG1RF compared to an *fsrB* mutant, indicating a more complex regulatory network [105]. Consistent with previous observations [17], we detected an up-regulation of the *fsrABC* genes (EF1822-20) in MMH594 at t_{30} . The *fsrA* gene was also up-regulated at t_5 . Interestingly, we found no regulation of these genes in OG1RF (the genes are divergent in Symbioflor 1). In MMH594, the downstream *gelE* (EF1818) was down-regulated at t_5 , but not at t_{30} . The *sprE* gene (EF1817) was not significantly regulated at either

time points. These results show that for MMH594, transfer to urine immediately triggers an elevated transcription of the *fsr*-quorum sensing system. Quorum sensing regulatory cascades are characteristically initiated by elevated expression of a regulatory unit, in this case *fsr*-operon, of which the most likely consequence would be the subsequent induction of the *fsr*-regulon.

The *fsr*-quorum sensing system is one of in total 18 two component systems in the V583 genome [106,107]. Of these two component systems, MMH594 contains 17, and OG1RF and Symbioflor contain 14. The histidine kinase -response regulator system 17 (hk-rr17; EF1633-32) has been suggested to be negatively regulated by *fsr* [105]. This could be consistent with modulation of *fsr* in MMH594. However, the absence of *fsr* in Symbioflor 1, and the observed down-regulation of hk-rr 17 system in this strain demonstrate that *fsr* can not be responsible for the repression of the hk-rr17 system in Symbioflor 1. Several of the other two-component systems were also affected by growth in urine such as hk-rr02 (EF3197-96) and hk-rr04 (EF1703-04), which were up-regulated in all strains at t_{30} and hk-rr10 (EF1050-51), which was down-regulated in all strains at both time points (although not significantly in all strains). The latter system encodes *etaRS*, which has been shown to be involved in stress response and virulence in *E. faecalis* [108].

The PAI is significantly more prevalent among infection-derived isolates compared to *E. faecalis* from other sources [22,28,109,110]. Moreover, the contribution of PAI-related genes to the pathogenicity of *E. faecalis* has been experimentally determined for certain traits (*araC*, cytolysin, *esp*)[5,102,111]. In the genome of the three strains used in this study only MMH594 contains the entire enterococcal PAI (129 genes), and 125 of the PAI genes are represented on the array. Fifteen genes including the *araC*-regulator (EF0524) were down-regulated, while twenty genes including manganese transporter (EF0575-77), *gls24* (EF0604) and a bile salt hydrolase (EF0521) were up-regulated at t_{30} in MMH594. The latter gene was also the only PAI gene which showed an enhanced expression in Symbioflor 1. These results demonstrate that potential virulence-, stress- and fitness-genes located in the PAI do in fact respond to an infection-relevant milieu like urine. However, the exact function of these genes in the pathogenicity of *E. faecalis* remains to be elucidated. Moreover, transcripts were detected for a substantial number of PAI genes, implying that their mere presence and basal expression might also be important during UTI.

Although the PAI is found in high frequency in infection derived isolates [22,28,109,110], the contribution of many PAI-genes to the pathogenicity of *E. faecalis* remains to be elucidated. Our experiment revealed that approximately one third of the PAI genes were differentially expressed in MMH594. In addition, transcripts were detected for a substantial number of PAI genes, implying that their mere presence and basal expression might be important also during UTI.

In conclusion, a significant proportion of the transcriptional responses seen during growth in urine were common for the three different strains examined, and the main differential regulation was observed among genes related to stress responses, energy metabolism, acquisition of trace metals, and a drastic modification of the cell envelope. One intriguing difference between the two pathogenic strains OG1RF and MMH594 was the regulation of the *fsr*-system, which could signify a difference in the virulence potential between these two strains.

Materials and methods

Bacterial strains and growth conditions: Bacterial strains used in this study are listed in Table 3. The growth capacity of six *Enterococcus faecalis* strains was examined. Three of these strains were selected for transcriptional profiling based on their origin. For all experiments *E. faecalis* strains were streaked on a 2xYT agar plate (1% (w/v) yeast extract, 1.6% (w/v) tryptone and 1% (w/v) NaCl) and incubated at 37°C over night (ON). Four individual colonies were then inoculated into the same tube of 5 ml 2xYT medium and grown ON without shaking at 37°C. For growth in urine, human urine was collected from four healthy men and women who had no history of UTI or antibiotic use in the last 6 months. The urine was pooled with equal amounts from each volunteer, centrifuged at 12000 x g and sterilized twice by filtration (0.22 µm-pore size). Since the composition of human urine may potentially be variable, samples were collected on three separate days for three replicate experiments and used within the next day.

Growth measurement: The six *E. faecalis* strains were pre-cultured as described above. ON cultures were diluted 1000x in either preheated urine (37°C) or in preheated 2xYT medium and incubated ON. These cultures were then diluted 1000x in either preheated urine or 2xYT, and cell growth was measured spectrophotometrically with a Bioscreen instrument (Bioscreen

C) and by plating and colony forming units (cfu) counts. Growth experiments measured spectrophotometrically were performed in triplicates with a total volume of 300 μ l of bacterial inoculum in fresh urine or 2xYT medium. Wells containing sterile urine / 2xYT were used as negative controls. Cultures were incubated at 37°C and optical density 600 nm (OD₆₀₀) was measured at 15-min intervals for 24 hours. To determine CFU/ml, viable cell counts were performed as follows: ON cultures were inoculated (1000x dilution) in preheated urine. Samples were collected immediately after inoculation, and after 2, 4, 6, 8, 10 and 24 hours for 2xYT, and also after 15 hours for urine. The number of CFU/ml was estimated by averaging the colony count values in two replicates per strain after ON incubation at 37 °C.

Cultivation and sampling prior to microarray analysis: The three selected *E. faecalis* strains, MMH594, OG1RF and Symbioflor 1 were pre-cultured as described above. The cultures were then diluted 1000x in 250 ml pre-warmed 2xYT medium and incubated further at 37°C. When the culture reached OD₆₀₀ = 0.1 the cultures from each strain was split in two and centrifuged (10000 x g for 3 min at 37°C). For the control cultures the pellets were resuspended in 100 ml pre-warmed 2xYT (37°C) whereas for the test culture the pellet was resuspended in 100 ml pre-warmed urine (37°C). Samples (45 ml) of each culture were collected immediately after the resuspension in urine (t₅), and after 30 min (t₃₀) by centrifugation (8000 x g for 2 min at 37°C), and the pellets were immediately frozen in liquid nitrogen and kept at -80°C prior to RNA extraction.

RNA isolation, cDNA synthesis, fluorescent labeling and hybridization: Total RNA was isolated by FastPrep (Bio 101/Savant) and RNeasy Mini kit (QIAGEN) as previously described [71]. The concentrations of the RNA samples were measured by using the NanoDrop (NanoDrop Technologies), and the quality was assessed by using the RNA 600 Nano LabChip kit and the Bioanalyzer 2100 (Agilent Technologies). cDNA was synthesized and labeled with the Fairplay III Microarray labeling kit (Stratagene) according to the manufacturer's protocol, with the following modifications: For each labeling reaction, 10 μ g of total RNA and 500 ng of random primers were initially preheated at 70°C for 10 min. A reverse transcription-PCR mixture (10x AffinityScript RT buffer, a 20x deoxynucleoside triphosphate mixture, 0.1 M dithiothreitol, 20 U RNase block, and AffinityScript HC RT) was added to the annealed primers and RNA, and the reaction mixture was further incubated for 3 h at 42°C. After labeling, 1 μ L of hydroxylamine (Sigma Aldrich) was added to quench the coupling reaction, and the reaction mixture was incubated 10 min. at room temperature. 70 μ L RNase-free water was then added, and unincorporated dyes were removed

from the samples by using the QIAquick PCR purification kit (QIAGEN). Labeled samples were then dried, prior to resuspension in 140 μ l hybridization solution (5x SSC, 0.1 % (w/v) SDS, 1.0 % (w/v) bovine serum albumin, 50 % (v/v) formamide and 0.01 % (w/v) single-stranded salmon sperm DNA) and hybridized for 16 h at 42 °C to the array in a Tecan HS 400 pro hybridization station (Tecan). Arrays were washed twice at 42 °C with 2x SSC + 0.2 % SDS, and twice at 23 °C with 2x SSC, followed by more stringent washes at 23 °C with 0.2x SSC and with filtrated H₂O. Three replicate hybridizations were performed with three separate batches of RNA. The three batches of RNA were obtained in three separate growth experiments. The Cy3 and Cy5 dyes (Amersham) used during cDNA synthesis were swapped in two of the three replicate hybridizations. All samples in the three experiments were co-hybridized with control samples collected at equal time points (*e.g.* t_5 was hybridized along with t_5). Hybridized arrays were scanned at wavelengths of 532 nm (Cy3) and 635 nm (Cy5) with a Tecan scanner LS (Tecan). Fluorescent intensities and spot morphologies were analyzed using GenePix Pro 6.0 (Molecular Devices), and spots were excluded based on slide or morphology abnormalities.

Microarrays: The microarray used in this work has been described previously [112]. The microarray designs have been deposited in the ArrayExpress database with the accession numbers A-MEXP-1688 and A-MEXP-1765.

Data analysis: Downstream analysis was done by the LIMMA package (www.bioconductor.org) in the R computing environment (www.r-project.org). Preprocessing and normalization followed a standard procedure using methods described by Smyth & Speed [113]. Testing for differential expressed gene was done by using a linear mixed model as described in Smyth [114]. A mixed-model approach was chosen to adequately describe between-array variation and still utilize probe-replicates (3 replicates of each probe in each array). An empirical Bayes smoothing of gene-wise variances was conducted according to Smyth *et al* [115]. For each gene, the p-value was adjusted to control the false discovery rate; hence, all p-values displayed are FDR-adjusted (and referred to as *q*-values). A gene was found to be significantly regulated if $q < 0.01$ and the log₂-ratio was similar or above 0.5, or similar or below -0.5. A gene represented with less than 1 spot on one or more arrays was excluded from the final results and marked as NA.

Comparative genomic hybridization: Genomic DNA was isolated by using the FP120 FastPrep bead-beater (BIO101/Savent) and the QiaPrep MiniPrep kit (Qiagen), as previously

described [112], and then labeled and purified with the BioPrime Array CGH Genomic labeling System (Invitrogen) and Cyanine Smart Pack dUTP (PerkinElmer Life Sciences), according to the manufacturer's protocol. Standard methods in the LIMMA package [113] in R (<http://www.r-project.org/>), available from the Bioconductor (<http://www.bioconductor.org>) were employed for preprocessing and normalization. Within-array normalization was first conducted by subtracting the median from the log-ratios for each array. A standard loess-normalization was then performed, where smoothing was based only on spots with $\text{abs}(\log\text{-ratio}) < 2.0$ to avoid biases due to extreme skewness in the log-ratio distribution. For the determination of present and divergent genes a method that predicts sequence identity based on array signals was used, as described by Snipen et al. [116]. A threshold of 0.75 was used in order to obtain a categorical response of presence or divergence, *i.e.* genes with *Sb*-value > 0.75 were classified as present, while genes with *Sb*-value < 0.75 were classified as divergent. Genes with *Sb*-value = 0.75 remained unclassified.

Microarray data accession number: The microarray data have been deposited in the ArrayExpress database with the series accession number E-TABM-885.

Acknowledgments

We thank the Norwegian Microarray Consortium, Trondheim, for printing the microarray slides. HCV was financially supported by grant 142656 from The Research Council of Norway, and MS by a grant from the European Union Sixth Framework Program under contract LSHE-CT-2007-037410.

References

1. Richards MJ, Edwards JR, Culver DH, Gaynes RP (2000) Nosocomial infections in combined medical-surgical intensive care units in the United States. *Infect Control Hosp Epidemiol* 21: 510-515.
2. Wisplinghoff H, Bischoff T, Tallent SM, Seifert H, Wenzel RP, et al. (2004) Nosocomial bloodstream infections in US hospitals: analysis of 24,179 cases from a prospective nationwide surveillance study. *Clin Infect Dis* 39: 309-317.
3. Gross PA, Harkavy LM, Barden GE, Flower MF (1976) The epidemiology of nosocomial enterococcal urinary tract infection. *Am J Med Sci* 272: 75-81.

4. Hancock LE, Gilmore MS (2006) Pathogenicity of enterococci. In: Fischetti VA, Novick RP, Ferretti JJ, Portnoy DA, Rood JI, editors. Gram-positive pathogens. Washington DC: ASM Press. pp. p. 299-311.
5. Shankar N, Lockett CV, Baghdayan AS, Drachenberg C, Gilmore MS, et al. (2001) Role of *Enterococcus faecalis* surface protein Esp in the pathogenesis of ascending urinary tract infection. *Infect Immun* 69: 4366-4372.
6. Chow JW, Thal LA, Perri MB, Vazquez JA, Donabedian SM, et al. (1993) Plasmid-associated hemolysin and aggregation substance production contribute to virulence in experimental enterococcal endocarditis. *Antimicrob Agents Chemother* 37: 2474-2477.
7. Jett BD, Jensen HG, Nordquist RE, Gilmore MS (1992) Contribution of the pAD1-encoded cytolysin to the severity of experimental *Enterococcus faecalis* endophthalmitis. *Infect Immun* 60: 2445-2452.
8. Schlievert PM, Gahr PJ, Assimacopoulos AP, Dinges MM, Stoehr JA, et al. (1998) Aggregation and binding substances enhance pathogenicity in rabbit models of *Enterococcus faecalis* endocarditis. *Infect Immun* 66: 218-223.
9. Kreft B, Marre R, Schramm U, Wirth R (1992) Aggregation substance of *Enterococcus faecalis* mediates adhesion to cultured renal tubular cells. *Infect Immun* 60: 25-30.
10. Olmsted SB, Dunny GM, Erlandsen SL, Wells CL (1994) A plasmid-encoded surface protein on *Enterococcus faecalis* augments its internalization by cultured intestinal epithelial cells. *J Infect Dis* 170: 1549-1556.
11. Eaton TJ, Gasson MJ (2001) Molecular screening of *Enterococcus* virulence determinants and potential for genetic exchange between food and medical isolates. *Appl Environ Microbiol* 67: 1628-1635.
12. Lempiainen H, Kinnunen K, Mertanen A, von Wright A (2005) Occurrence of virulence factors among human intestinal enterococcal isolates. *Lett Appl Microbiol* 41: 341-344.
13. Semedo T, Santos MA, Lopes MF, Figueiredo Marques JJ, Barreto Crespo MT, et al. (2003) Virulence factors in food, clinical and reference Enterococci: A common trait in the genus? *Syst Appl Microbiol* 26: 13-22.
14. Creti R, Imperi M, Bertuccini L, Fabretti F, Orefici G, et al. (2004) Survey for virulence determinants among *Enterococcus faecalis* isolated from different sources. *J Med Microbiol* 53: 13-20.
15. Franz CM, Muscholl-Silberhorn AB, Yousif NM, Vancanneyt M, Swings J, et al. (2001) Incidence of virulence factors and antibiotic resistance among Enterococci isolated from food. *Appl Environ Microbiol* 67: 4385-4389.

16. Mannu L, Paba A, Daga E, Comunian R, Zanetti S, et al. (2003) Comparison of the incidence of virulence determinants and antibiotic resistance between *Enterococcus faecium* strains of dairy, animal and clinical origin. *Int J Food Microbiol* 88: 291-304.
17. Shepard BD, Gilmore MS (2002) Differential expression of virulence-related genes in *Enterococcus faecalis* in response to biological cues in serum and urine. *Infect Immun* 70: 4344-4352.
18. Alteri CJ, Mobley HL (2007) Quantitative profile of the uropathogenic *Escherichia coli* outer membrane proteome during growth in human urine. *Infect Immun* 75: 2679-2688.
19. Russo TA, Carlino UB, Mong A, Jodush ST (1999) Identification of genes in an extraintestinal isolate of *Escherichia coli* with increased expression after exposure to human urine. *Infect Immun* 67: 5306-5314.
20. Domann E, Hain T, Ghai R, Billion A, Kuenne C, et al. (2007) Comparative genomic analysis for the presence of potential enterococcal virulence factors in the probiotic *Enterococcus faecalis* strain Symbioflor 1. *Int J Med Microbiol* 297: 533-539.
21. Huycke MM, Spiegel CA, Gilmore MS (1991) Bacteremia caused by hemolytic, high-level gentamicin-resistant *Enterococcus faecalis*. *Antimicrob Agents Chemother* 35: 1626-1634.
22. Shankar N, Baghdayan AS, Gilmore MS (2002) Modulation of virulence within a pathogenicity island in vancomycin-resistant *Enterococcus faecalis*. *Nature* 417: 746-750.
23. Bourgogne A, Garsin DA, Qin X, Singh KV, Sillanpaa J, et al. (2008) Large scale variation in *Enterococcus faecalis* illustrated by the genome analysis of strain OG1RF. *Genome Biol* 9: R110.
24. Murray BE, Singh KV, Ross RP, Heath JD, Dunny GM, et al. (1993) Generation of restriction map of *Enterococcus faecalis* OG1 and investigation of growth requirements and regions encoding biosynthetic function. *J Bacteriol* 175: 5216-5223.
25. Sifri CD, Mylonakis E, Singh KV, Qin X, Garsin DA, et al. (2002) Virulence effect of *Enterococcus faecalis* protease genes and the quorum-sensing locus *fsr* in *Caenorhabditis elegans* and mice. *Infect Immun* 70: 5647-5650.
26. Stamey TA, Mihara G (1980) Observations on the growth of urethral and vaginal bacteria in sterile urine. *J Urol* 124: 461-463.
27. Carlos AR, Santos J, Semedo-Lemsaddek T, Barreto-Crespo MT, Tenreiro R (2009) Enterococci from artisanal dairy products show high levels of adaptability. *Int J Food Microbiol* 129: 194-199.
28. McBride SM, Coburn PS, Baghdayan AS, Willems RJ, Grande MJ, et al. (2009) Genetic variation and evolution of the pathogenicity island of *Enterococcus faecalis*. *J Bacteriol* 191: 3392-3402.

29. Hallin PF, Binnewies TT, Ussery DW (2008) The genome BLASTatlas-a GeneWiz extension for visualization of whole-genome homology. *Mol Biosyst* 4: 363-371.
30. Hechard Y, Pelletier C, Cenatiempo Y, Frere J (2001) Analysis of sigma(54)-dependent genes in *Enterococcus faecalis*: a mannose PTS permease (EII^{Man}) is involved in sensitivity to a bacteriocin, mesentericin Y105. *Microbiology* 147: 1575-1580.
31. Shaykhutdinov R, MacInnis G, Dowlatabadi R, Weljie A, Vogel H (2009) Quantitative analysis of metabolite concentrations in human urine samples using ¹³C{¹H} NMR spectroscopy. *Metabolomics* 5: 307-317.
32. Deutscher J, Francke C, Postma PW (2006) How phosphotransferase system-related protein phosphorylation regulates carbohydrate metabolism in bacteria. *Microbiol Mol Biol Rev* 70: 939-1031.
33. Blancato VS, Repizo GD, Suarez CA, Magni C (2008) Transcriptional regulation of the citrate gene cluster of *Enterococcus faecalis* Involves the GntR family transcriptional activator CitO. *J Bacteriol* 190: 7419-7430.
34. Wishart D, Knox C, Guo A (2009) HMDB: a knowledgebase for the human metabolome. 37(Database issue):D603-610. *Nucleic Acids Res.*
35. Tasevska N, Runswick SA, McTaggart A, Bingham SA (2005) Urinary sucrose and fructose as biomarkers for sugar consumption. *Cancer Epidemiol Biomarkers Prev* 14: 1287-1294.
36. Maadani A, Fox KA, Mylonakis E, Garsin DA (2007) *Enterococcus faecalis* mutations affecting virulence in the *Caenorhabditis elegans* model host. *Infect Immun* 75: 2634-2637.
37. Garsin DA, Sifri CD, Mylonakis E, Qin X, Singh KV, et al. (2001) A simple model host for identifying Gram-positive virulence factors. *Proc Natl Acad Sci U S A* 98: 10892-10897.
38. Riboulet-Bisson E, Sanguinetti M, Budin-Verneuil A, Auffray Y, Hartke A, et al. (2008) Characterization of the Ers regulon of *Enterococcus faecalis*. *Infect Immun* 76: 3064-3074.
39. Riboulet-Bisson E, Hartke A, Auffray Y, Giard JC (2009) Ers controls glycerol metabolism in *Enterococcus faecalis*. *Curr Microbiol* 58: 201-204.
40. Roos V, Klemm P (2006) Global gene expression profiling of the asymptomatic bacteriuria *Escherichia coli* strain 83972 in the human urinary tract. *Infect Immun* 74: 3565-3575.
41. Guo K, Li L (2009) Differential ¹²C-/¹³C-isotope dansylation labeling and fast liquid chromatography/mass spectrometry for absolute and relative quantification of the metabolome. *Anal Chem* 81: 3919-3932.
42. Deibel RH (1964) Utilization of arginine as an energy source for the growth of *Streptococcus faecalis*. *J Bacteriol* 87: 988-992.
43. Roon RJ, Barker HA (1972) Fermentation of agmatine in *Streptococcus faecalis*: occurrence of putrescine transcarbamoylase. *J Bacteriol* 109: 44-50.

44. Le Breton Y, Muller C, Auffray Y, Rince A (2007) New insights into the *Enterococcus faecalis* CroRS two-component system obtained using a differential-display random arbitrarily primed PCR approach. *Appl Environ Microbiol* 73: 3738-3741.
45. Lloyd AL, Rasko DA, Mobley HL (2007) Defining genomic islands and uropathogen-specific genes in uropathogenic *Escherichia coli*. *J Bacteriol* 189: 3532-3546.
46. MacLeod RA (1951) Further mineral requirements of *Streptococcus faecalis*. *J Bacteriol* 62: 337-345.
47. MacLeod RA, Snell EE (1947) Some mineral requirements of the lactic acid bacteria. *J Biol Chem* 170: 351-365.
48. Jarvisalo J, Olkinuora M, Kiilunen M, Kivisto H, Ristola P, et al. (1992) Urinary and blood manganese in occupationally nonexposed populations and in manual metal arc welders of mild steel. *Int Arch Occup Environ Health* 63: 495-501.
49. Low YL, Jakubovics NS, Flatman JC, Jenkinson HF, Smith AW (2003) Manganese-dependent regulation of the endocarditis-associated virulence factor EfaA of *Enterococcus faecalis*. *J Med Microbiol* 52: 113-119.
50. Singh KV, Coque TM, Weinstock GM, Murray BE (1998) In vivo testing of an *Enterococcus faecalis* *efaA* mutant and use of *efaA* homologs for species identification. *FEMS Immunol Med Microbiol* 21: 323-331.
51. Manson J, Gilmore M (2006) 7. Pathogenomics of *Enterococcus faecalis*. In: Hacker J, Dobrindt U, editors. *Pathogenomics: genome analysis of pathogenic microbes*. Weinheim: WILEY-VCH Verlag GmbH & Co. KGaA. pp. 125-148.
52. Rince A, Flahaut S, Auffray Y (2000) Identification of general stress genes in *Enterococcus faecalis*. *Int J Food Microbiol* 55: 87-91.
53. Giard JC, Rince A, Capioux H, Auffray Y, Hartke A (2000) Inactivation of the stress- and starvation-inducible *gls24* operon has a pleiotrophic effect on cell morphology, stress sensitivity, and gene expression in *Enterococcus faecalis*. *J Bacteriol* 182: 4512-4520.
54. Nannini EC, Teng F, Singh KV, Murray BE (2005) Decreased virulence of a *gls24* mutant of *Enterococcus faecalis* OG1RF in an experimental endocarditis model. *Infect Immun* 73: 7772-7774.
55. Teng F, Nannini EC, Murray BE (2005) Importance of *gls24* in virulence and stress response of *Enterococcus faecalis* and use of the Gls24 protein as a possible immunotherapy target. *J Infect Dis* 191: 472-480.
56. Rince A, Giard JC, Pichereau V, Flahaut S, Auffray Y (2001) Identification and characterization of *gsp65*, an organic hydroperoxide resistance (*ohr*) gene encoding a general stress protein in *Enterococcus faecalis*. *J Bacteriol* 183: 1482-1488.

57. Verneuil N, Rince A, Sanguinetti M, Auffray Y, Hartke A, et al. (2005) Implication of *hypR* in the virulence and oxidative stress response of *Enterococcus faecalis*. FEMS Microbiol Lett 252: 137-141.
58. Riboulet E, Verneuil N, La Carbona S, Sauvageot N, Auffray Y, et al. (2007) Relationships between oxidative stress response and virulence in *Enterococcus faecalis*. J Mol Microbiol Biotechnol 13: 140-146.
59. Verneuil N, Rince A, Sanguinetti M, Posteraro B, Fadda G, et al. (2005) Contribution of a PerR-like regulator to the oxidative-stress response and virulence of *Enterococcus faecalis*. Microbiology 151: 3997-4004.
60. Giard JC, Riboulet E, Verneuil N, Sanguinetti M, Auffray Y, et al. (2006) Characterization of Ers, a PrfA-like regulator of *Enterococcus faecalis*. FEMS Immunol Med Microbiol 46: 410-418.
61. Flahaut S, Hartke A, Giard JC, Benachour A, Boutibonnes P, et al. (1996) Relationship between stress response toward bile salts, acid and heat treatment in *Enterococcus faecalis*. FEMS Microbiol Lett 138: 49-54.
62. La Carbona S, Sauvageot N, Giard JC, Benachour A, Posteraro B, et al. (2007) Comparative study of the physiological roles of three peroxidases (NADH peroxidase, Alkyl hydroperoxide reductase and Thiol peroxidase) in oxidative stress response, survival inside macrophages and virulence of *Enterococcus faecalis*. Mol Microbiol 66: 1148-1163.
63. Verneuil N, Maze A, Sanguinetti M, Laplace JM, Benachour A, et al. (2006) Implication of (Mn)superoxide dismutase of *Enterococcus faecalis* in oxidative stress responses and survival inside macrophages. Microbiology 152: 2579-2589.
64. Elgavish A, Lloyd K, Reed R (1996) A subpopulation of human urothelial cells is stimulated to proliferate by treatment *in vitro* with lipoteichoic acid, a cell wall component of *Streptococcus faecalis*. J Cell Physiol 169: 42-51.
65. Elgavish A (2000) NF-kappaB activation mediates the response of a subpopulation of basal uroepithelial cells to a cell wall component of *Enterococcus faecalis*. J Cell Physiol 182: 232-238.
66. Jordan S, Hutchings MI, Mascher T (2008) Cell envelope stress response in Gram-positive bacteria. FEMS Microbiol Rev 32: 107-146.
67. Rowley G, Spector M, Kormanec J, Roberts M (2006) Pushing the envelope: extracytoplasmic stress responses in bacterial pathogens. Nat Rev Microbiol 4: 383-394.
68. Theilacker C, Kaczynski Z, Kropec A, Fabretti F, Sange T, et al. (2006) Opsonic antibodies to *Enterococcus faecalis* strain 12030 are directed against lipoteichoic acid. Infect Immun 74: 5703-5712.

69. Thurlow LR, Thomas VC, Fleming SD, Hancock LE (2009) *Enterococcus faecalis* capsular polysaccharide serotypes C and D and their contributions to host innate immune evasion. *Infect Immun* 77: 5551-5557.
70. Teng F, Jacques-Palaz KD, Weinstock GM, Murray BE (2002) Evidence that the enterococcal polysaccharide antigen gene (*epa*) cluster is widespread in *Enterococcus faecalis* and influences resistance to phagocytic killing of *E. faecalis*. *Infect Immun* 70: 2010-2015.
71. Vebø HC, Snipen L, Nes IF, Brede DA (2009) The transcriptome of the nosocomial pathogen *Enterococcus faecalis* V583 reveals adaptive responses to growth in blood. *PLoS One* 4: e7660.
72. Solheim M, Aakra A, Vebo H, Snipen L, Nes IF (2007) Transcriptional responses of *Enterococcus faecalis* V583 to bovine bile and sodium dodecyl sulfate. *Appl Environ Microbiol* 73: 5767-5774.
73. Brinster S, Lamberet G, Staels B, Trieu-Cuot P, Gruss A, et al. (2009) Type II fatty acid synthesis is not a suitable antibiotic target for Gram-positive pathogens. *Nature* 458: 83-86.
74. Walecka E, Bania J, Dworniczek E, Ugorski M (2009) Genotypic characterization of hospital *Enterococcus faecalis* strains using multiple-locus variable-number tandem-repeat analysis. *Lett Appl Microbiol* 49: 79-84.
75. Mohamed JA, Huang DB (2007) Biofilm formation by enterococci. *J Med Microbiol* 56: 1581-1588.
76. Hufnagel M, Koch S, Creti R, Baldassarri L, Huebner J (2004) A putative sugar-binding transcriptional regulator in a novel gene locus in *Enterococcus faecalis* contributes to production of biofilm and prolonged bacteremia in mice. *J Infect Dis* 189: 420-430.
77. Le Breton Y, Pichereau V, Sauvageot N, Auffray Y, Rince A (2005) Maltose utilization in *Enterococcus faecalis*. *J Appl Microbiol* 98: 806-813.
78. Guiton PS, Hung CS, Kline KA, Roth R, Kau AL, et al. (2009) Contribution of autolysin and Sortase a during *Enterococcus faecalis* DNA-dependent biofilm development. *Infect Immun* 77: 3626-3638.
79. Kristich CJ, Nguyen VT, Le T, Barnes AM, Grindle S, et al. (2008) Development and use of an efficient system for random mariner transposon mutagenesis to identify novel genetic determinants of biofilm formation in the core *Enterococcus faecalis* genome. *Appl Environ Microbiol* 74: 3377-3386.
80. Kemp KD, Singh KV, Nallapareddy SR, Murray BE (2007) Relative contributions of *Enterococcus faecalis* OG1RF sortase-encoding genes, *srtA* and *bps* (*srtC*), to biofilm formation and a murine model of urinary tract infection. *Infect Immun* 75: 5399-5404.
81. Creti R, Fabretti F, Koch S, Huebner J, Garsin DA, et al. (2009) Surface protein EF3314 contributes to virulence properties of *Enterococcus faecalis*. *Int J Artif Organs* 32: 611-620.

82. Sillanpaa J, Xu Y, Nallapareddy SR, Murray BE, Hook M (2004) A family of putative MSCRAMMs from *Enterococcus faecalis*. *Microbiology* 150: 2069-2078.
83. Singh KV, Nallapareddy SR, Murray BE (2007) Importance of the *ebp* (endocarditis- and biofilm-associated pilus) locus in the pathogenesis of *Enterococcus faecalis* ascending urinary tract infection. *J Infect Dis* 195: 1671-1677.
84. Nallapareddy SR, Singh KV, Sillanpaa J, Garsin DA, Hook M, et al. (2006) Endocarditis and biofilm-associated pili of *Enterococcus faecalis*. *J Clin Invest* 116: 2799-2807.
85. Jude BA, Martinez RM, Skorupski K, Taylor RK (2009) Levels of the secreted *Vibrio cholerae* attachment factor GbpA are modulated by quorum-sensing-induced proteolysis. *J Bacteriol* 191: 6911-6917.
86. Bjork S, Breimer ME, Hansson GC, Karlsson KA, Leffler H (1987) Structures of blood group glycosphingolipids of human small intestine. A relation between the expression of fucolipids of epithelial cells and the ABO, Le and Se phenotype of the donor. *J Biol Chem* 262: 6758-6765.
87. Finne J, Breimer ME, Hansson GC, Karlsson KA, Leffler H, et al. (1989) Novel polyfucosylated N-linked glycopeptides with blood group A, H, X, and Y determinants from human small intestinal epithelial cells. *J Biol Chem* 264: 5720-5735.
88. Kirn TJ, Jude BA, Taylor RK (2005) A colonization factor links *Vibrio cholerae* environmental survival and human infection. *Nature* 438: 863-866.
89. Mohamed JA, Teng F, Nallapareddy SR, Murray BE (2006) Pleiotrophic effects of 2 *Enterococcus faecalis* *sagA*-like genes, *salA* and *salB*, which encode proteins that are antigenic during human infection, on biofilm formation and binding to collagen type i and fibronectin. *J Infect Dis* 193: 231-240.
90. Mesnage S, Chau F, Dubost L, Arthur M (2008) Role of N-acetylglucosaminidase and N-acetylmuramidase activities in *Enterococcus faecalis* peptidoglycan metabolism. *J Biol Chem* 283: 19845-19853.
91. Thomas VC, Hiromasa Y, Harms N, Thurlow L, Tomich J, et al. (2009) A fratricidal mechanism is responsible for eDNA release and contributes to biofilm development of *Enterococcus faecalis*. *Mol Microbiol* 72: 1022-1036.
92. Xu Y, Murray BE, Weinstock GM (1998) A cluster of genes involved in polysaccharide biosynthesis from *Enterococcus faecalis* OG1RF. *Infect Immun* 66: 4313-4323.
93. Teng F, Singh KV, Bourgogne A, Zeng J, Murray BE (2009) Further characterization of the *epa* gene cluster and Epa polysaccharides of *Enterococcus faecalis*. *Infect Immun* 77: 3759-3767.
94. Xu Y, Singh KV, Qin X, Murray BE, Weinstock GM (2000) Analysis of a gene cluster of *Enterococcus faecalis* involved in polysaccharide biosynthesis. *Infect Immun* 68: 815-823.

95. Mohamed JA, Huang W, Nallapareddy SR, Teng F, Murray BE (2004) Influence of origin of isolates, especially endocarditis isolates, and various genes on biofilm formation by *Enterococcus faecalis*. *Infect Immun* 72: 3658-3663.
96. Singh KV, Lewis RJ, Murray BE (2009) Importance of the *epa* Locus of *Enterococcus faecalis* OG1RF in a Mouse Model of Ascending Urinary Tract Infection. *J Infect Dis* 200: 417-420.
97. Kau AL, Martin SM, Lyon W, Hayes E, Caparon MG, et al. (2005) *Enterococcus faecalis* tropism for the kidneys in the urinary tract of C57BL/6J mice. *Infect Immun* 73: 2461-2468.
98. Hancock LE, Gilmore MS (2002) The capsular polysaccharide of *Enterococcus faecalis* and its relationship to other polysaccharides in the cell wall. *Proc Natl Acad Sci U S A* 99: 1574-1579.
99. Paulson JC, Colley KJ (1989) Glycosyltransferases. Structure, localization, and control of cell type-specific glycosylation. *J Biol Chem* 264: 17615-17618.
100. Paulsen IT, Banerjee L, Myers GS, Nelson KE, Seshadri R, et al. (2003) Role of mobile DNA in the evolution of vancomycin-resistant *Enterococcus faecalis*. *Science* 299: 2071-2074.
101. Theilacker C, Sanchez-Carballo P, Toma I, Fabretti F, Sava I, et al. (2009) Glycolipids are involved in biofilm accumulation and prolonged bacteraemia in *Enterococcus faecalis*. *Mol Microbiol* 71: 1055-1069.
102. Ike Y, Hashimoto H, Clewell DB (1984) Hemolysin of *Streptococcus faecalis* subspecies zymogenes contributes to virulence in mice. *Infect Immun* 45: 528-530.
103. Qin X, Singh KV, Weinstock GM, Murray BE (2000) Effects of *Enterococcus faecalis* *fsr* genes on production of gelatinase and a serine protease and virulence. *Infect Immun* 68: 2579-2586.
104. Carlos AR, Semedo-Lemsaddek T, Barreto-Crespo MT, Tenreiro R (2009) Transcriptional analysis of virulence-related genes in enterococci from distinct origins. *J Appl Microbiol*.
105. Bourgogne A, Hilsenbeck SG, Dunny GM, Murray BE (2006) Comparison of OG1RF and an isogenic *fsrB* deletion mutant by transcriptional analysis: the *Fsr* system of *Enterococcus faecalis* is more than the activator of gelatinase and serine protease. *J Bacteriol* 188: 2875-2884.
106. Hancock L, Perego M (2002) Two-component signal transduction in *Enterococcus faecalis*. *J Bacteriol* 184: 5819-5825.
107. Hancock LE, Perego M (2004) Systematic inactivation and phenotypic characterization of two-component signal transduction systems of *Enterococcus faecalis* V583. *J Bacteriol* 186: 7951-7958.
108. Teng F, Wang L, Singh KV, Murray BE, Weinstock GM (2002) Involvement of PhoP-PhoS homologs in *Enterococcus faecalis* virulence. *Infect Immun* 70: 1991-1996.

109. Nallapareddy SR, Wenxiang H, Weinstock GM, Murray BE (2005) Molecular characterization of a widespread, pathogenic, and antibiotic resistance-receptive *Enterococcus faecalis* lineage and dissemination of its putative pathogenicity island. *J Bacteriol* 187: 5709-5718.
110. McBride SM, Fischetti VA, Leblanc DJ, Moellering RC, Jr., Gilmore MS (2007) Genetic diversity among *Enterococcus faecalis*. *PLoS One* 2: e582.
111. Coburn PS, Baghdayan AS, Dolan GT, Shankar N (2008) An AraC-type transcriptional regulator encoded on the *Enterococcus faecalis* pathogenicity island contributes to pathogenesis and intracellular macrophage survival. *Infect Immun* 76: 5668-5676.
112. Solheim M, Aakra A, Snipen LG, Brede DA, Nes IF (2009) Comparative genomics of *Enterococcus faecalis* from healthy Norwegian infants. *BMC Genomics* 10: 194.
113. Smyth GK, Speed T (2003) Normalization of cDNA microarray data. *Methods* 31: 265-273.
114. Smyth GK (2004) Linear models and empirical bayes methods for assessing differential expression in microarray experiments. *Stat Appl Genet Mol Biol* 3: Article3.
115. Smyth GK, Michaud J, Scott HS (2005) Use of within-array replicate spots for assessing differential expression in microarray experiments. *Bioinformatics* 21: 2067-2075.
116. Snipen L, Nyquist OL, Solheim M, Aakra A, Nes IF (2009) Improved analysis of bacterial CGH data beyond the log-ratio paradigm. *BMC Bioinformatics* 10: 91.
117. Giard JC, Laplace JM, Rince A, Pichereau V, Benachour A, et al. (2001) The stress proteome of *Enterococcus faecalis*. *Electrophoresis* 22: 2947-2954.
118. Laport MS, Lemos JA, Bastos Md Mdo C, Burne RA, Giambiagi-De Marval M (2004) Transcriptional analysis of the *groE* and *dnaK* heat-shock operons of *Enterococcus faecalis*. *Res Microbiol* 155: 252-258.
119. Giard JC, Hartke A, Flahaut S, Boutibonnes P, Auffray Y (1997) Glucose starvation response in *Enterococcus faecalis* JH2-2: survival and protein analysis. *Res Microbiol* 148: 27-35.
120. Verneuil N, Sanguinetti M, Le Breton Y, Posteraro B, Fadda G, et al. (2004) Effects of the *Enterococcus faecalis hypR* gene encoding a new transcriptional regulator on oxidative stress response and intracellular survival within macrophages. *Infect Immun* 72: 4424-4431.
121. Laplace JM, Hartke A, Giard JC, Auffray Y (2000) Cloning, characterization and expression of an *Enterococcus faecalis* gene responsive to heavy metals. *Appl Microbiol Biotechnol* 53: 685-689.
122. Giard JC, Verneuil N, Auffray Y, Hartke A (2002) Characterization of genes homologous to the general stress-inducible gene *gls24* in *Enterococcus faecalis* and *Lactococcus lactis*. *FEMS Microbiol Lett* 206: 235-239.

123. Sahm DF, Kissinger J, Gilmore MS, Murray PR, Mulder R, et al. (1989) In vitro susceptibility studies of vancomycin-resistant *Enterococcus faecalis*. *Antimicrob Agents Chemother* 33: 1588-1591.
124. Aakra A, Nyquist OL, Snipen L, Reiersen TS, Nes IF (2007) Survey of genomic diversity among *Enterococcus faecalis* strains by microarray-based comparative genomic hybridization. *Appl Environ Microbiol* 73: 2207-2217.
125. Paley SM, Karp PD (2002) Evaluation of computational metabolic-pathway predictions for *Helicobacter pylori*. *Bioinformatics* 18: 715-724.
126. Romero PR, Karp PD (2004) Using functional and organizational information to improve genome-wide computational prediction of transcription units on pathway-genome databases. *Bioinformatics* 20: 709-717.

Tables

Table 1: Differentially expressed genes with a proven or predicted function in various stress responses in *E. faecalis*.

ORF	Other names	Gene product	Log ₂ -ratio*						Reference
			MMH594		OG1RF		Symbioflor 1		
			<i>t</i> ₅	<i>t</i> ₃₀	<i>t</i> ₅	<i>t</i> ₃₀	<i>t</i> ₅	<i>t</i> ₃₀	
<i>General stress response</i>									
EF0453	Gsp65	OsmC/Ohr family protein	0.9	3.0	0.7	3.1	0.9	3.8	[52,56]
EF0770	Gsp62	Conserved hypothetical protein	1.1	4.0	0.9	2.0	0.9	3.1	[52,117]
EF1058		Universal stress protein family	-	-	-	-	2.2	2.3	[100]
EF1084		Universal stress protein family	-	-	-	-	-	0.7	[100]
EF1308	Gsp66	DnaK protein	0.8	-	-	- 1.1	-	- 0.6	[52,118]
EF1560	Gsp63	Hypothetical protein	1.2	3.1	-	-	-	1.3	[52,117]
EF1744		General stress protein, putative	0.6	1.8	-	-	0.8	1.8	[100]
EF1810		General stress protein A (<i>gspA-1</i>)	-	2.1	-	-	-	-	[100]
EF1811		General stress protein A (<i>gspA-2</i>)	1.1	4.1	1.3	-	-	2.7	[100]
EF1982	Gls33	Universal stress protein family	1.1	1.6	1.4	-	1.6	2.3	[100,117,119]
EF2633	Gsp67	Chaperonin, 60 kDa (<i>groEL</i>)	0.8	-	-	- 1.1	-	-	[52,118]
EF2797	Gsp64	Hypothetical protein	-	-	-	-	0.6	1.1	[52,117]
EF2894		General stress protein 13, putative	-	-	-	-	0.6	-	[100]
EF3035		Universal stress protein family	-	-	0.7	-	0.5	-	[100]
<i>Oxidative stress</i>									
EF0074		Transcriptional regulator, Crp/Fnr family (<i>ers</i>)	-	-	- 0.9	-	- 1.1	- 0.8	[60]
EF0463		Superoxide dismutase (<i>sodA</i>)	1.0	3.5	-	2.1	-	3.3	[100]
EF1211		NADH peroxidase (<i>npr</i>)	0.7	3.0	0.8	2.2	0.5	3.7	[100]
EF1338		Thioredoxine reductase (<i>trxB</i>)	-	1.2	-	1.4	-	1.1	[100]
EF1405		Thioredoxine (<i>trx</i>)	-	1.6	-	-	-	1.3	[58]
EF1525		Transcriptional regulator, Fur family (<i>fur</i>)	-	- 0.6	-	-	-	-	[58,59]
EF1585		Transcriptional regulator, Fur family (<i>perR</i>)	-	1.3	-	0.7	-	1.5	[58,59]
EF1586		NADH oxidase (<i>nox</i>)	- 1.2	-	- 1.2	0.9	- 0.8	0.7	[100]
EF1597		Catalase (<i>katA</i>)	- 0.8	2.0	- 1.2	- 1.0	-	1.1	[100]
EF1681		Peptide methionine-S-sulfoxide reductase (<i>msrA</i>)	-	3.2	-	-	-	1.7	[58]
EF2055		Oxidoreductase, pyridine nucleotide-disulfide family	- 0.7	1.4	- 0.8	1.5	- 0.9	0.8	[100]
EF2417		Transcriptional regulator, Fur family (<i>zur</i>)	-	-	-	-	-	0.6	[58,59]
EF2958		Transcriptional regulator, LysR family (<i>hypR</i>)	-	-	-	-	0.5	0.9	[57,120]
EF3164		Peptide methionine-R-sulfoxide reductase (<i>msrB/ csrA</i>)	- 0.6	4.4	-	1.7	-	3.3	[57,121]
EF3233		Dps family protein (<i>dps</i>)	0.7	2.0	0.6	1.6	1.0	2.1	[100]
EF3257		Oxidoreductase, pyridine nucleotide-disulfide family	-	- 1.0	-	-	-	- 1.6	[100]
EF3270		Glutathione reductase (<i>gor</i>)	-	0.7	-	1.1	0.5	1.5	[100]
<i>Osmotic stress</i>									
EF0295		V-type ATPase, subunit J	-	- 0.7	-	-	- 0.5	- 1.0	[100]
EF0402		Na ⁺ /H ⁺ antiporter	- 0.8	-	-	-	-	- 0.9	[100]
EF0636		Na ⁺ /H ⁺ antiporter	1.0	- 4.5	1.2	- 5.3	-	- 3.8	[100]
<i>Metal-ion resistance</i>									
EF0298		Copper-translocating P-type ATPase	-	2.3	1.2	2.1	1.7	3.4	[100]
EF0299		Copper transport protein CopZ	-	2.3	-	2.7	-	4.1	[100]

EF0758		Cadmium-translocating P-type ATPase	-	-	-	-	1.0	[100]	
EF0871		Cation-transporting ATPase, E1-E2 family	0.7	0.7	-	-	1.1	[100]	
EF0875		Copper-translocating P-type ATPase	1.7	2.2	1.4	1.0	1.5	2.0	[100]
EF1268		Cation-transporting ATPase, E1-E2 family	-	1.2	-	-	-	0.6	[100]
EF1352		Magnesium-translocating P-type ATPase	-	-	1.3	-0.7	1.5	-	[100]
EF1400		Cadmium-translocating P-type ATPase	-	-0.8	-	-	-	-	[100]
EF1519		Cation-transporting ATPase, E1-E2 family	-0.9	-1.0	-	-	-0.7	-0.8	[100]
EF2623		Cadmium-translocating P-type ATPase	-	-1.3	-	-1.6	0.6	-	[100]
<i>Heat shock</i>									
EF1306		Heat-inducible transcription repressor HrcA (<i>hrcA</i>)	1.0	-0.8	0.7	-1.1	0.8	-	[100,118]
EF1307		Heat shock protein GrpE (<i>grpE</i>)	1.0	-	-	-1.2	-	-	[100,118]
EF1646		Heat shock protein HslVU, ATPase subunit (<i>hslU</i>)	1.2	1.7	0.5		0.7	0.9	[100,118]
EF1647		Heat shock protein HslV (<i>hslV</i>)	1.5	1.7	0.9		1.1	0.6	[100,118]
EF2634		Chaperonin, 10 kDa (<i>groES</i>)	0.9	-	0.7	-	-	-0.7	[118]
<i>Glucose starvation</i>									
EF0079		Gls24 protein (<i>gls24</i>)	-	-	0.5	3.1	-	3.1	[119,122]
EF0080		Gls24 protein (<i>glsB</i>)	0.7	4.4	0.9	3.3	0.8	3.0	[119,122]
EF0099	Gls17	L-serine dehydratase, iron-sulfur-dependent	-	-3.8	1.6	-5.0	0.8	-2.9	[117,119]
EF0106	Gls14	Carbamate kinase		-2.2			-	-2.9	[117,119]
EF1359	Gls40	Conserved hypothetical protein		1.0	0.6	-	0.6	-0.6	[117,119]
EF1360	Gls10	Dihydroxyacetone kinase family protein		0.7	0.6		0.6	-	[117,119]
EF1361	Gls27	Dihydroxyacetone kinase family protein	0.7	2.0	0.9	1.3	0.9	-	[117,119]
EF1962	Gls23	Triosephosphate isomerase (<i>tpiA</i>)	-	-1.3	-	-	-	-	[117,119]
EF2500	Gls37	GcvH family protein	-	-1.0	-0.8	-	-0.6	-1.0	[117,119]
PAlef0055 ^a		Stress-induced protein Gls24	-	0.9	-	-	-	-	[22]

*Only significant log₂-ratios are listed.

^a Present in MMH594 only.

Table 2: Differentially expressed genes with a proven or predicted virulence function in *E. faecalis*. Only significant log₂-ratios are listed.

ORF	Gene name	Gene product	Log ₂ -ratio*						Reference
			MMH594		OG1RF		Symbioflor 1		
			t5	t30	t5	t30	t5	t30	
EF0031		membrane protein, putative	-	0,6	-	-	-	1,0	[100]
EF0032		membrane protein, putative	-	-	-	0,9	-	0,7	[100]
EF0055		adhesion lipoprotein	-0,7	-0,7	-1,3	-	-	-0,8	[100]
EF0079	<i>gls24</i>	gls24 protein	-	-	0,5	3,1	-	3,1	[54,55]
EF0080	<i>glsB</i>	gls24 protein	0,7	4,4	0,9	3,3	0,8	3,0	[54,55]
EF0089		conserved domain protein	-0,8	-1,3	-1,4	-2,1	-1,0	-1,3	[100]
EF0123		conserved hypothetical protein	-	-0,5	-	-	-	-	[100]
EF0355	<i>atlB</i>	endolysin, putative	-	-1,0	-	-	-	-	[90,100]
EF0361		chitinase, family 2	0,9	-	0,6	-	0,7	-	[100]
EF0362		chitin binding protein, putative	1,1	-	0,8	-	0,7	-	[100]
EF0394	<i>salB</i>	secreted antigen, putative	-1,5	-0,7	-3,4	-	-1,1	-	[89,100]
EF0463	<i>sodA</i>	superoxide dismutase, Mn	1,0	3,5	-	2,1	0,4	3,3	[63]
EF0468		LemA family protein	4,0	-	5,7	-	3,8	1,4	[100]
EF0526	<i>cylL-S</i>	cylL-S protein	-	2,2	-	-	-	-	[100]
EF0527	<i>cylM</i>	cylM protein	-	-0,5	-	-	-	-	[100]
EF0577		adhesion lipoprotein	3,4	4,1	-	-	-	-	[100]
EF0645		exfoliative toxin A, putative	-0,6	-	-	-	-0,8	-	[100]
EF0680		penicillin-binding protein 2A	-	-	0,6	-	-	-	[100]
EF0700		hemolysin	-	-	-	-	-0,7	-	[100]
EF0746		penicillin-binding protein, putative	-	0,7	1,2	-	0,8	-	[100]
EF0799	<i>atlA</i>	autolysin	-1,7	-	-1,3	-	-1,9	-	[78,90,91,100]
EF0927		sensor histidine kinase	-	-0,4	0,6	-	-	-	[100]
EF0941		ABC transporter, ATP-binding	-	-	-	-	-0,6	-	[100]
EF0944		extracellular protein, putative	-1,7	-1,5	-1,6	-0,9	-2,1	-1,8	[100]
EF0955	<i>bopC</i>	aldose 1-epimerase, putative	-	-	1,2	-	-	-	[76]
EF0956	<i>bopB</i>	beta-phosphoglucomutase	-	-	1,2	-	-	-	[76]
EF0991	<i>pbpC</i>	penicillin-binding protein C	-0,6	-1,1	-1,1	-1,2	-1,0	-1,8	[100]
EF1032	<i>drrC</i>	daunorubicin resistance protein	-	1,4	1,1	1,2	0,5	1,7	[100]
EF1038		lipoprotein, putative	0,7	-	1,2	1,7	-	-	[100]
EF1050	<i>etaR</i>	DNA-binding response regulator	-	-	-1,0	-	-0,8	-1,0	[108]
EF1051	<i>etaS</i>	sensor histidine kinase	-	-0,9	-0,7	-	-0,8	-	[108]
EF1057		Mn ²⁺ /Fe ²⁺ transporter	4,1	3,9	4,0	4,6	4,0	3,9	[100]
EF1148		penicillin-binding protein 1A	-	-0,4	-	-	-0,7	-0,9	[100]
EF1340		pheromone cAM373 prec.lipoprot.	-0,8	-1,5	-1,6	-0,9	-0,8	-1,1	[100]
EF1502		beta-lactamase, putative	-1,2	-3,0	-0,8	-1,6	-0,7	-1,7	[100]
EF1603	<i>scrB-1</i>	sucrose-6-phosphate dehydrogenase	2,0	3,8	2,3	-	3,2	4,4	[36,37]
EF1604	<i>scrR-1</i>	sucrose operon repressor ScrR	-	1,7	-	-	-	3,1	[36,37]
EF1740		penicillin-binding protein 1B, putative	-	-0,5	-	-	-	-	[100]
EF1818	<i>gelE</i>	coccolysin	-0,5	-	-	-	-	-	[25,100,103]
EF1820	<i>fsrC</i>	histidine kinase, putative	-	1,0	-	-	-	-	[25,103]
EF1821	<i>fsrB</i>	agrBfs protein	-	1,0	-	-	-	-	[25,103]
EF1822	<i>fsrA</i>	response regulator	0,7	0,8	-	-	-	-	[25,103]
EF2074	<i>efaC</i>	ABC transporter	2,8	3,0	3,1	3,1	3,7	3,4	[50]
EF2075	<i>efaB</i>	ABC transporter	2,7	3,5	3,0	3,7	3,6	3,4	[50]
EF2076	<i>efaA</i>	endocarditis specific antigen	2,3	2,9	2,8	3,4	3,0	3,2	[100] [50]
EF2093		endolysin domain protein	-	-0,6	-	-	-	-	[100]
EF2167		glycosyl transferase	-1,1	-2,6	-	-	-	-	[100]
EF2170		glycosyl transferase	-	-1,3	-	-	-	-	[100]
EF2174		conserved domain protein	-0,9	-	-	-	-1,0	-0,9	[100]
EF2176		glycosyl transferase	-0,8	-1,0	-1,7	-	-1,1	-0,5	[100]
EF2177	<i>epaR</i>	bacterial sugar transferase	-1,1	-	-	-	-0,8	-1,6	[70,95,96]
EF2178	<i>epaQ</i>	membrane protein, putative	-1,3	-1,9	-0,9	-1,2	-0,5	-2,1	[70,95,96]
EF2179	<i>epaP</i>	conserved hypothetical protein	-2,5	-3,6	-1,2	-0,9	-1,0	-2,7	[70,95,96]
EF2180	<i>epaO</i>	glycosyl transferase,	-1,4	-1,5	-0,8	-	-0,6	-1,6	[70,95,96]

EF2181	<i>epaN</i>	glycosyl transferase	-0,8	-2,1	-0,5	-0,7	-0,9	-1,7	[70,95,96]
EF2182	<i>epaM</i>	ABC transporter	-0,9	-1,5	-	-	-0,9	-2,1	[70,95,96]
EF2183	<i>epaL</i>	ABC transporter	-0,9	-1,9	-	-	-1,1	-2,3	[70,95,96]
EF2184	<i>epaK</i>	hypothetical protein	-0,7	-1,6	-1,2	-	-0,8	-1,8	[70,95,96]
EF2186		conserved domain protein	-1,2	-2,9	-	-	-	-	[70,95,96]
EF2188		racemase domain protein	-1,2	-1,8	-	-	-	-	[70,95,96]
EF2189	<i>epaJ</i>	conserved hypothetical protein	-1,2	-2,2	-1,1	-	-0,9	-1,4	[70,95,96]
EF2190	<i>epaI</i>	glycosyl transferase	-1,1	-2,0	-0,9	-	-0,6	-1,6	[70,95,96]
EF2191	<i>epaH</i>	dTDP-4-dehydrorhamnose reductase	-1,0	-1,5	-0,7	-	-0,8	-1,5	[70,95,96]
EF2192	<i>epaG</i>	dTDP-glucose 4,6-dehydratase	-0,7	-	-	-	-0,8	-	[70,95,96]
EF2193	<i>epaF</i>	dTDP-4-dehydrorhamnose	-0,7	-	-0,7	-	-0,6	-0,9	[70,95,96]
EF2194	<i>epaE</i>	glucose-1-ph thymidyltransferase	-0,9	-1,1	-0,7	-0,6	-0,9	-1,8	[70,95,96]
EF2195	<i>epaD</i>	glycosyl transferase	-1,2	-1,7	-	-0,5	-0,9	-1,2	[70,95,96]
EF2196	<i>epaC</i>	glycosyl transferase	-1,0	-	-	-	-0,5	-0,5	[70,95,96]
EF2197	<i>epaB</i>	glycosyl transferase	-0,9	-	-	-	-0,5	-0,5	[70,95,96]
EF2198	<i>epaA</i>	glycosyl transferase	-0,8	-0,9	-	-	-0,5	-1,0	[70,95,96]
EF2202		tspO protein, putative	1,0	2,8	-	-	-	1,7	[100]
EF2439		undecaprenol kinase, putative	-1,4	-1,3	-1,5	-	-1,5	-1,8	[100]
EF2485	<i>cpsK</i>	ABC transporter	-	-3,1	-	-	-	-	[69,98]
EF2486	<i>cpsJ</i>	ABC transporter	-	-3,1	-	-	-	-	[69,98]
EF2487	<i>cpsI</i>	UDP-galactopyranose mutase	-	-2,7	-	-	-	-	[69,98]
EF2488	<i>cpsH</i>	lipoprotein, putative	-	-2,1	-	-	-	-	[69,98]
EF2489	<i>cpsG</i>	MurB family protein	-	-2,8	-	-	-	-	[69,98]
EF2490	<i>cpsF</i>	conserved hypothetical protein	-	-3,1	-	-	-	-	[69,98]
EF2491	<i>cpsE</i>	glycosyl transferase	-	-2,5	-	-	-	-	[69,98]
EF2492	<i>cpsD</i>	glycosyl transferase	-0,7	-2,3	-	-	-	-	[69,98]
EF2493	<i>cpsC</i>	teichoic acid biosynth. prot., putative	-	-1,0	-	-	-	-	[69,98]
EF2496		pheromone cOB1 precursor/lipoprot.	-	-	-	-	-0,9	-	[100]
EF2658		FemAB family protein	-	-	-0,8	-	-	-	[100]
EF2680		ABC transporter	-	0,7	-	-	-	-	[100]
EF2713		cell wall surface anchor family protein	2,2	-	-	-	-	-	[100]
EF2751		permease protein, putative	0,7	1,2	-	-	-	0,8	[100]
EF2802		endolysin	-	-	-	-	-	-0,6	[100]
EF2890		glycosyl transferase	-	-0,6	-	-1,2	-	-	[100]
EF2891	<i>bgsA</i>	glycosyl transferase	-	-	-	-1,1	-	-	[100,101]
EF2929		membrane protein, putative	-	0,8	-	-	-0,5	-	[100]
EF3023		polysaccharide lyase, family 8	-	-	-	-	-	-1,1	[100]
EF3056		sortase family protein	-	1,1	-	-	0,8	1,1	[78-80]
EF3060	<i>salA</i>	secreted lipase, putative	-3,5	-0,7	-3,6	-1,1	-3,1	-0,7	[89,100]
EF3076		cell wall surface anchor family protein	-	-0,5	-	-	-	-	[100]
EF3082		iron compound ABC transporter	-	2,3	-	3,4	-0,6	1,7	[100]
EF3083		iron compound ABC transporter	-	2,3	-	3,3	-	1,5	[100]
EF3084		iron compound ABC transporter	-0,7	2,7	-	4,0	-0,6	1,7	[100]
EF3085		iron compound ABC transporter	-	2,0	-	2,7	-	2,5	[100]
EF3106		peptide ABC transporter	0,6	5,1	0,8	3,4	-	-	[100]
EF3129		D-alanyl-D-alanine carboxypeptidase	-0,6	-0,8	-0,7	-0,7	-	-	[100]
EF3183		cell wall surface anchor family protein	-0,7	-1,1	-	-	-	-	[100]
EF3187		cell wall surface anchor family protein	-0,8	-0,9	-	-	-	-	[100]
EF3191		lipase, putative	-	0,6	-	-	-	0,8	[100]
EF3198		lipoprotein, YaeC family	-	0,7	-	0,7	-	1,5	[100]
EF3206		adhesion lipoprotein	-	-	-	1,1	-	-	[100]
EF3245		cell-envelope ass. acid phosphatase	-	-0,5	-	-	-	-	[100]
EF3256		pheromone cAD1 precursor lipoprot.	-0,6	-2,4	-	-	-	-0,6	[100]
EF3314		cell wall surface anchor family protein	-	-	-	-	0,9	3,5	[81,100]
EF3331		pheromone cCF10 precursor/lipoprot.	-	-1,5	-	-1,1	-	-1,4	[100]
PAlef0049 ^a	<i>cyl I</i>	CylI protein	-	-0,9	-	-	-	-	[6,7,102]

*Only significant log₂-ratios are listed.

^a Present in MMH594 only.

Table 3. Bacterial strains used in this study.

Strain	Country	Source	Isolation site	MLST		Characteristics	Reference
				CC	ST		
Baby isolate 62	Norway	Non-hospitalized person < 1 year	Feces	S	66	Tet ^R	[112]
MMH594	USA	Hospitalized patient	Blood	6	6	Ery ^R , Gen ^R , hospital outbreak	[21]
OG1RF	USA	Laboratory strain		21	1	Rif ^R , Fus ^R	[24]
Symbioflor 1	Germany	Non-hospitalized person	Feces	25	248	Probiotic	[20]
V583	USA	Hospitalized patient	Blood	6	6	Ery ^R , Gen ^R , Van ^R	[123]
179Vet	Norway	Animal_dog	Urine	9	9	Multi-resistant*	[124]

* Tested against 16 different antibiotics, of which it was susceptible only to ampicillin.

Figure Legends

Figure 1: Characterization of growth of *E. faecalis* MMH594 (black circle), OG1RF (red triangle), Symbioflor 1 (green square) Baby isolate 62 (yellow diamond), V583 (blue triangle) and 179Vet (pink hexagon) in urine. The growth curves are represented by colony forming units per millilitre (CFU/ml) on the Y-axis, and hours as indicated on the X-axis. The growth curves correspond to the mean \pm STD of two parallels.

Figure 2. Venn diagram showing the distribution of genes classified as present in the three test strains. The percentages indicated for each strain specify how large part of total probe set represented on the array that was classified as present in the corresponding strain.

Figure 3. *E. faecalis* V583 genome-atlas [29] presentation of CGH analysis and transcriptional responses to growth (t_{30}) in urine compared to 2xYT. Mobile genetic elements [100,110] are indicated by brackets. From outer to inner lanes: 1) V583 annotated CDS, 2) CGH MMH594, 3) CGH Symbioflor1, 4) CGH OG1RF, 5) Urine transcriptome MMH594, 6) Urine transcriptome Symbioflor1, 7) Urine transcriptome OG1RF, 8) Intrinsic curvature, 9) Stacking energy, 10) Position Preference, 11) Global direct repeats, 12) Global inverted repeats, 13) GC skew, 14) AT percent. Interactive Genewiz atlases of CGH and transcriptome data are available at; <http://ws.cbs.dtu.dk/cgi-bin/gwBrowser/edit.cgi?hexkey=603430a081eb5be3f306e744e94b151a>, and <http://ws.cbs.dtu.dk/cgi-bin/gwBrowser/edit.cgi?hexkey=3068b894fb6b9e44d9c135a210950f52>, respectively.

Figure 4: Venn diagram showing the number of unique and common up- and down-regulated genes in MMH594, OG1RF and Symbioflor 1 when grown in urine compared to 2xYT after A: 5 minutes and B: 30 minutes.

Figure 5: Heat map visualizing the regulated genes in MMH594 (M), Symbioflor 1 (S) and OG1RF (O) when grown in urine compared to in 2xYT. The comparative genome hybridization (CGH) results for the respective regulated genes are shown in columns 1-3 (light blue: present gene, white: divergent gene). Genes found to be significantly regulated are indicated by either red (up-regulated), or blue (down-regulated). Genes regulated after growth for 5 minutes in urine compared to in 2xYT are listed in columns 4-6 and after 30 minutes in columns 7-9. The functional categories are sorted alphabetically (column 10). Significantly regulated hypothetical genes and genes encoding proteins with unknown function are not included in this heat map.

Figure 1

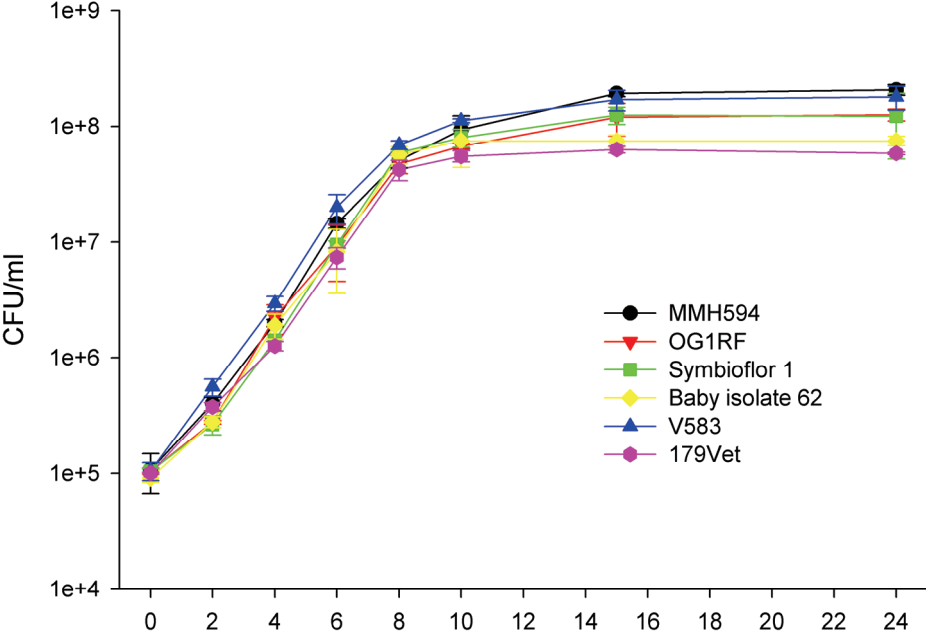


Figure 2

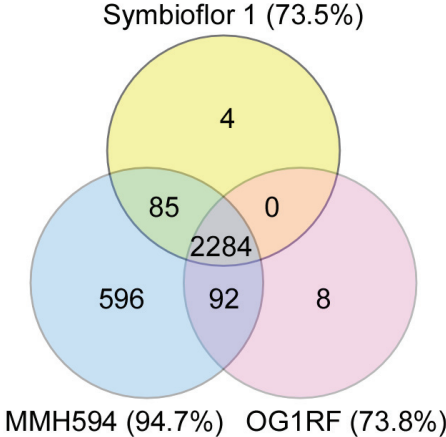


Figure 3

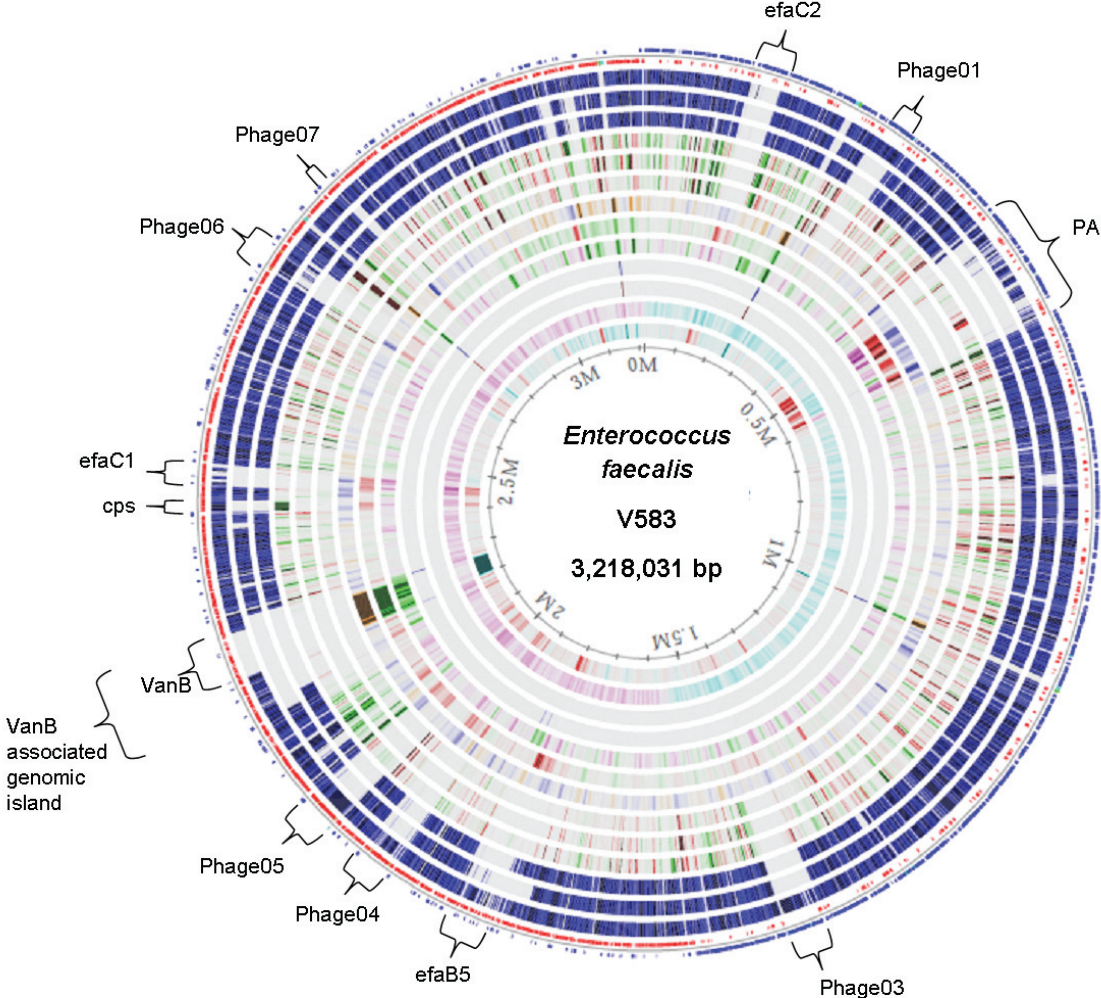


Figure 4

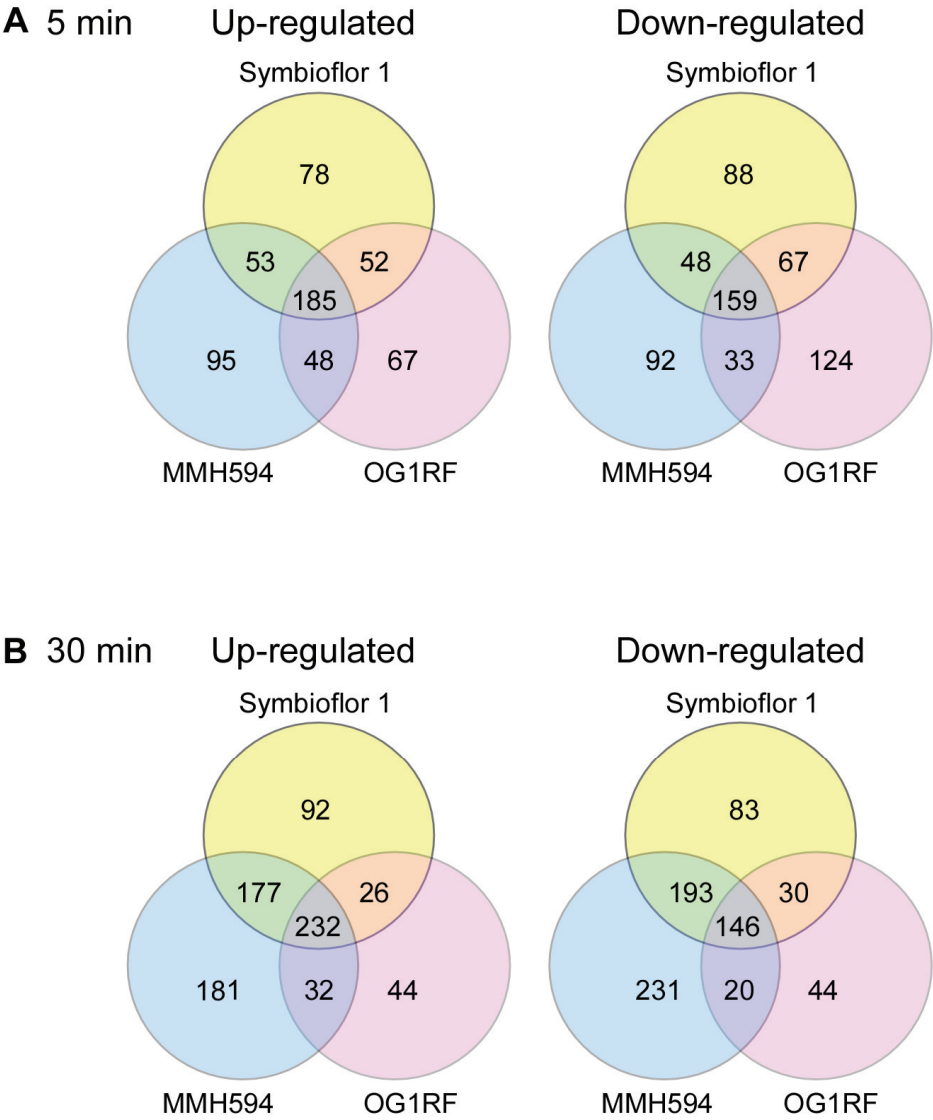
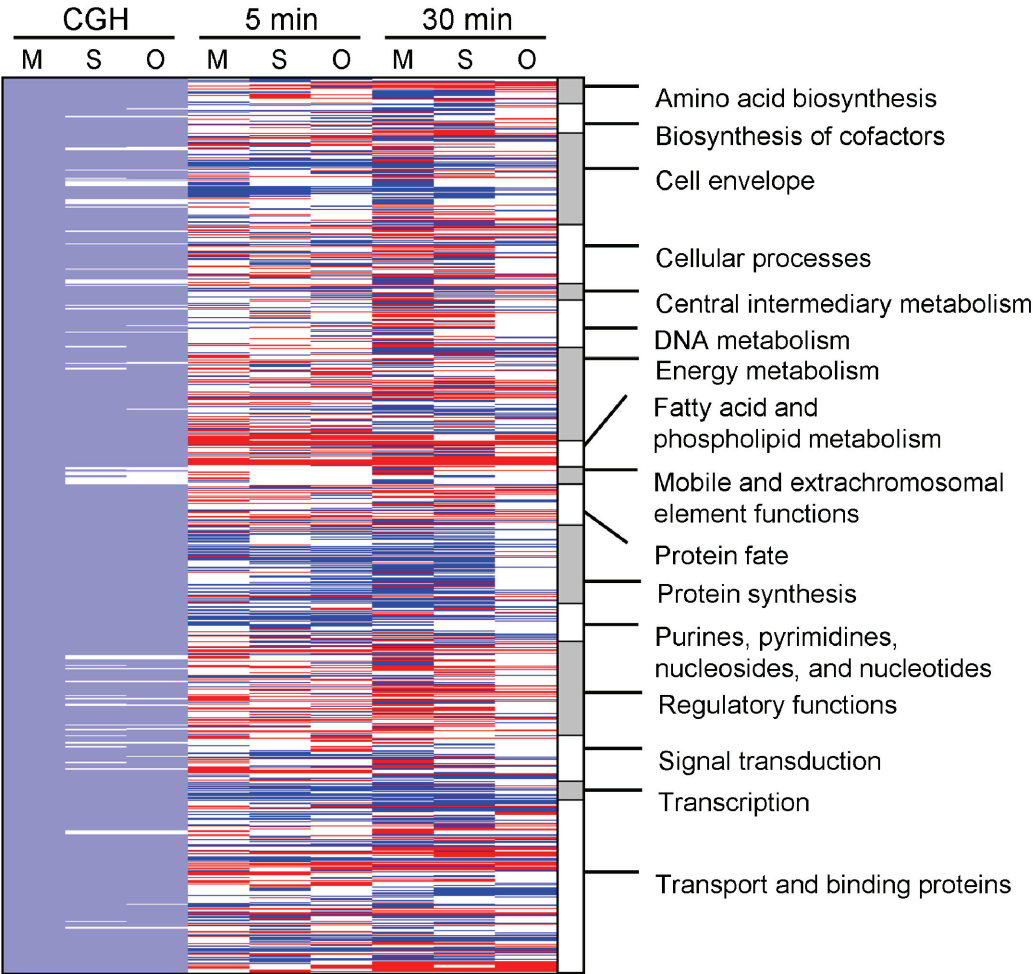


Figure 5



Supplementary material

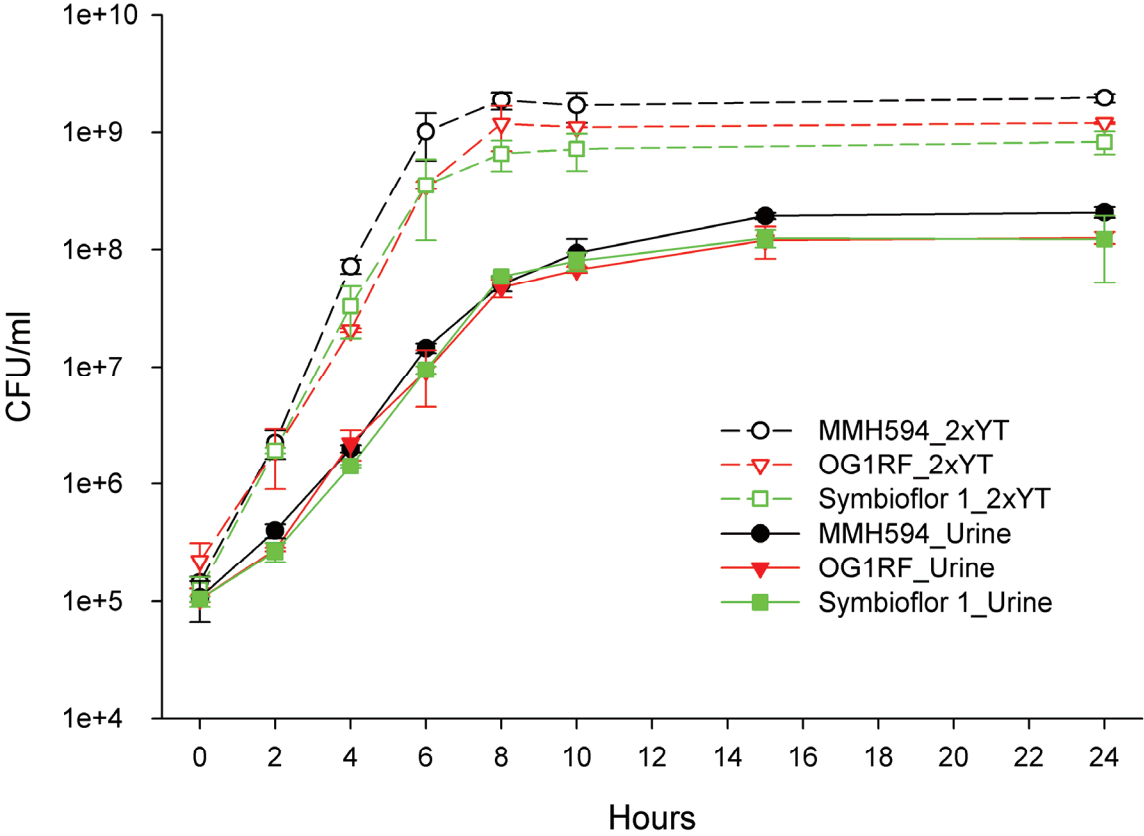


Figure S1: Characterization of growth of *E. faecalis* MMH594 (black circle), OG1RF (red triangle) and Symbioflor 1 (green square) in 2xYT (stippled lines) and urine (solid lines). The growth curves are represented by colony forming units per millilitre (CFU/ml) on the Y-axis, and hours as indicated on the X-axis. The growth curves correspond to the mean \pm STD of two parallels

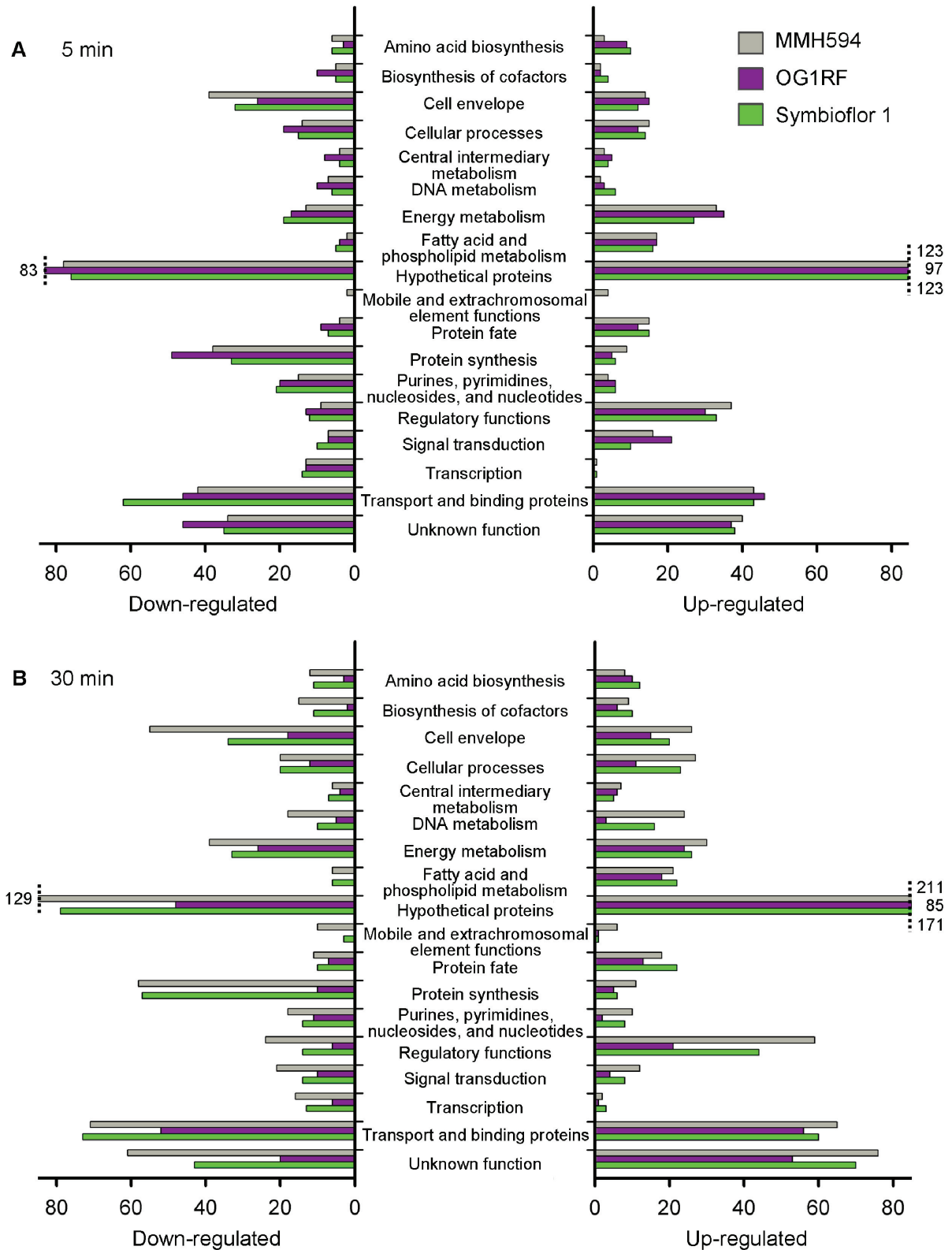


Figure S2: Overview of the number of up- and down-regulated genes in MMH594 (grey), OG1RF (purple) and Symbioflor 1 (green) at A) 5 minutes and B) 30 minutes. The functional categories are listed between the two bar-charts.

Table S1. Microarray expression data and comparative genome hybridization of *E. faecalis* strains MMH594 (M), OG1RF (O) and Symbioflor 1 (S). Differences in gene content were analyzed using comparative genomic hybridization (*): present (1), divergent (0), unclassified (U). Gene expression after 5 (t5) or 30 (t30) minutes of growth in urine is relative to 2xYT. Significantly regulated genes are $q < 0.01$ (bold), and $\log_2\text{-ratio} > \pm 0.5$. "NA" denotes non expressed or excluded genes.

Locus	Gene	Functional category	Subcategory	M	T	O	S	T	M	I	S	q-value	M_130	q-value	O_15	q-value	O_30	q-value	S_15	q-value	S_30	q-value			
EF1093	<i>ebpC</i>	Cell envelope	cell wall surface anchor family protein	1	1	1	1	1	1	1	1	0.18960	-0.7	0.79230	-0.1	0.18960	-0.7	0.79230	-0.1	0.18960	-0.7	0.79230	-0.2	0.35540	
EF1094	<i>srfC</i>	Surface structures	Other	1	1	1	1	1	1	1	1	0.40380	0.5	0.01659	0.2	0.40380	0.5	0.01659	0.2	0.40380	0.5	0.01659	0.4	0.05280	
EF1095		Hypothetical proteins	Surface structures	1	1	1	1	1	1	1	1	0.05650	0.5	0.01659	0.2	0.05650	0.5	0.01659	0.2	0.05650	0.5	0.01659	0.4	0.05280	
EF1096		Hypothetical proteins	Conserved	1	1	1	1	1	1	1	1	0.14620	3.5	0.00000	0.7	0.14620	3.5	0.00000	0.7	0.14620	3.5	0.00000	0.9	0.00088	
EF1097		Hypothetical proteins	Conserved	1	1	1	1	1	1	1	1	0.37920	0.2	0.37920	0.2	0.37920	0.2	0.37920	0.2	0.37920	0.2	0.37920	0.2	0.00000	
EF1098		Hypothetical proteins	Conserved	1	1	1	1	1	1	1	1	0.06885	0.2	0.37980	0.4	0.06885	0.2	0.37980	0.4	0.06885	0.2	0.37980	0.6	0.00104	
EF1099		Cell envelope	collagen adhesin protein	1	1	1	1	1	1	1	1	0.78950	0.2	0.05743	0.4	0.78950	0.2	0.05743	0.4	0.78950	0.2	0.05743	0.6	0.00170	
EF1100	<i>ace</i>	Transport and binding proteins	ABC transporter, ATP-binding/permease protein	1	1	1	1	1	1	1	1	0.32410	-0.4	0.32410	-0.4	0.32410	-0.4	0.32410	-0.4	0.32410	-0.4	0.32410	0.0	0.37880	
EF1101		Hypothetical proteins	conserved hypothetical protein	1	1	1	1	1	1	1	1	0.89630	-0.2	0.89630	-0.2	0.89630	-0.2	0.89630	-0.2	0.89630	-0.2	0.89630	0.6	0.00165	
EF1102		Hypothetical proteins	conserved hypothetical protein	1	1	1	1	1	1	1	1	0.00000	0.3	0.00000	0.3	0.00000	0.3	0.00000	0.3	0.00000	0.3	0.00000	0.2	0.00000	
EF1103		Transport and binding proteins	amino acid permease family protein	1	1	1	1	1	1	1	1	0.29790	0.1	0.29790	0.1	0.29790	0.1	0.29790	0.1	0.29790	0.1	0.29790	0.2	0.63010	
EF1104		Hypothetical proteins	conserved domain protein	1	1	1	1	1	1	1	1	0.34460	-0.2	0.34460	-0.2	0.34460	-0.2	0.34460	-0.2	0.34460	-0.2	0.34460	0.7	0.01272	
EF1105		Hypothetical proteins	hypothetical protein	1	1	1	1	1	1	1	1	0.94670	0.5	0.00054	0.4	0.94670	0.5	0.00054	0.4	0.94670	0.5	0.00054	0.6	0.00165	
EF1106		Hypothetical proteins	hypothetical protein	1	1	1	1	1	1	1	1	0.00000	0.4	0.00000	0.4	0.00000	0.4	0.00000	0.4	0.00000	0.4	0.00000	0.0	0.00000	
EF1107	<i>lurA</i>	Enzymes of unknown specificity	oxidoreductase, putative	1	1	1	1	1	1	1	1	0.87150	0.1	0.87150	0.1	0.87150	0.1	0.87150	0.1	0.87150	0.1	0.87150	0.9	0.00025	
EF1108	<i>lurB</i>	Electron transport	iron-sulfur cluster binding protein	1	1	1	1	1	1	1	1	0.36540	0.9	0.36540	0.9	0.36540	0.9	0.36540	0.9	0.36540	0.9	0.36540	0.6	0.00148	
EF1109	<i>lurC</i>	General	YkgG family protein	1	1	1	1	1	1	1	1	0.00006	0.1	0.00006	0.1	0.00006	0.1	0.00006	0.1	0.00006	0.1	0.00006	0.3	0.00025	
EF1110		Unknown function	signal peptidase I	1	1	1	1	1	1	1	1	0.11940	1.0	0.02747	0.4	0.11940	1.0	0.02747	0.4	0.11940	1.0	0.02747	0.4	0.00000	
EF1111		Protein fate	exonuclease RexA	1	1	1	1	1	1	1	1	0.57270	-0.5	0.00071	0.4	0.57270	-0.5	0.00071	0.4	0.57270	-0.5	0.00071	0.2	0.32630	
EF1112	<i>rexB</i>	DNA metabolism	conserved hypothetical protein	1	1	1	1	1	1	1	1	0.77340	0.2	0.65710	0.4	0.77340	0.2	0.65710	0.4	0.77340	0.2	0.65710	0.3	0.19690	
EF1113	<i>rexA</i>	DNA metabolism	conserved hypothetical protein	1	1	1	1	1	1	1	1	0.00023	1.6	0.00000	-0.1	0.00023	1.6	0.00000	-0.1	0.00023	1.6	0.00000	0.4	0.01741	
EF1114	<i>pheS</i>	Protein synthesis	RNA aminoacylation	1	1	1	1	1	1	1	1	0.65188	-1.0	0.00000	-0.4	0.65188	-1.0	0.00000	-0.4	0.65188	-1.0	0.00000	0.3	0.00476	
EF1115	<i>pheT</i>	Protein synthesis	phenylalanyl-tRNA synthetase, alpha subunit	1	1	1	1	1	1	1	1	0.33630	-1.2	0.00039	-0.6	0.33630	-1.2	0.00039	-0.6	0.33630	-1.2	0.00039	0.0	0.00144	
EF1116		Transport and binding proteins	phenylalanyl-tRNA synthetase, beta subunit	1	1	1	1	1	1	1	1	0.4	0.28100	1.8	0.00000	0.4	0.28100	1.8	0.00000	0.4	0.28100	1.8	0.00000	0.0	0.05007
EF1117		Transport and binding proteins	amino acid ABC transporter, permease protein	1	1	1	1	1	1	1	1	0.9	0.00545	2.6	0.00001	0.9	0.00545	2.6	0.00001	0.9	0.00545	2.6	0.00001	1.8	0.00001
EF1118		Transport and binding proteins	amino acid ABC transporter, permease protein	1	1	1	1	1	1	1	1	0.28100	1.8	0.00000	0.4	0.28100	1.8	0.00000	0.4	0.28100	1.8	0.00000	0.2	0.00001	
EF1119		Transport and binding proteins	amino acid ABC transporter, permease protein	1	1	1	1	1	1	1	1	0.9	0.00545	2.6	0.00001	0.9	0.00545	2.6	0.00001	0.9	0.00545	2.6	0.00001	1.8	0.00001
EF1120		Transport and binding proteins	amino acid ABC transporter, permease protein	1	1	1	1	1	1	1	1	0.28100	1.8	0.00000	0.4	0.28100	1.8	0.00000	0.4	0.28100	1.8	0.00000	0.2	0.00001	
EF1121	<i>murI</i>	Cell envelope	amino acid ABC transporter, amino acid-binding protein	1	1	1	1	1	1	1	1	0.00036	2.3	0.00000	2.9	0.00036	2.3	0.00000	2.9	0.00036	2.3	0.00000	0.2	0.00000	
EF1122	<i>mph</i>	Transcription	RNA processing	1	1	1	1	1	1	1	1	0.10424	2.6	0.00000	3.5	0.10424	2.6	0.00000	3.5	0.10424	2.6	0.00000	0.6	0.05028	
EF1123	<i>mph</i>	Hypothetical proteins	ribonuclease PH/Ham1 protein	1	1	1	1	1	1	1	1	0.32610	0.3	0.18600	0.3	0.32610	0.3	0.18600	0.3	0.32610	0.3	0.18600	0.2	0.56260	
EF1124		Regulatory functions	conserved hypothetical protein	1	1	1	1	1	1	1	1	0.49480	0.4	0.40250	0.3	0.49480	0.4	0.40250	0.3	0.49480	0.4	0.40250	0.2	0.19620	
EF1125		Hypothetical proteins	transcriptional regulator, DeoR family	1	1	1	1	1	1	1	1	0.3	0.49480	0.4	0.40250	0.3	0.49480	0.4	0.40250	0.3	0.49480	0.4	0.40250	0.4	0.19700
EF1126		Hypothetical proteins	conserved hypothetical protein	1	1	1	1	1	1	1	1	0.80560	0.1	0.80560	0.1	0.80560	0.1	0.80560	0.1	0.80560	0.1	0.80560	0.4	0.19700	
EF1127	<i>sgpT</i>	Transport and binding proteins	PTS system, IIA component	1	1	1	1	1	1	1	1	0.00000	0.1	0.00000	0.1	0.00000	0.1	0.00000	0.1	0.00000	0.1	0.00000	0.2	0.00000	
EF1128	<i>sgpB</i>	Transport and binding proteins	putative transport protein SgpT protein	1	1	1	1	1	1	1	1	0.00000	0.1	0.00000	0.1	0.00000	0.1	0.00000	0.1	0.00000	0.1	0.00000	0.2	0.00000	
EF1129		Energy metabolism	phosphotransferase enzyme II, B component SgpB	1	1	1	1	1	1	1	1	0.50760	0.1	0.50760	0.1	0.50760	0.1	0.50760	0.1	0.50760	0.1	0.50760	0.2	0.28230	
EF1130		Energy metabolism	hexulose-6-phosphate synthase, putative	1	1	1	1	1	1	1	1	0.48950	-0.1	0.48950	-0.1	0.48950	-0.1	0.48950	-0.1	0.48950	-0.1	0.48950	0.0	0.40890	
EF1131		Energy metabolism	L-xylulose-5-phosphate isomerase SgpU, putative	1	1	1	1	1	1	1	1	0.74410	1.4	0.00000	-0.6	0.74410	1.4	0.00000	-0.6	0.74410	1.4	0.00000	0.0	0.86260	
EF1132		Energy metabolism	peptide, M20M25M40 family	1	1	1	1	1	1	1	1	0.58220	2.2	0.00000	-0.6	0.58220	2.2	0.00000	-0.6	0.58220	2.2	0.00000	0.0	0.86260	
EF1133		Amino acid biosynthesis	2,3,4,5-tetrahydroxyindole-2,6-dicarboxylate N-succinyltransferase	1	1	1	1	1	1	1	1	0.40481	-0.1	0.40481	-0.1	0.40481	-0.1	0.40481	-0.1	0.40481	-0.1	0.40481	0.0	0.99897	
EF1134		Protein fate	conserved hypothetical protein	1	1	1	1	1	1	1	1	0.5	0.00000	0.1	0.5	0.00000	0.1	0.5	0.00000	0.1	0.5	0.00000	0.0	0.99897	
EF1135		Hypothetical proteins	hypothetical protein	1	1	1	1	1	1	1	1	0.57560	-0.4	0.07432	0.3	0.57560	-0.4	0.07432	0.3	0.57560	-0.4	0.07432	0.2	0.09920	
EF1136		Hypothetical proteins	hypothetical protein	1	1	1	1	1	1	1	1	0.6	0.03289	1.7	0.00002	0.8	0.6	0.03289	1.7	0.00002	0.8	0.6	0.03289	0.8	0.00380
EF1137		Hypothetical proteins	aldoketoreductase, aldoketoreductase family	1	1	1	1	1	1	1	1	0.32610	1.2	0.00000	0.3	0.32610	1.2	0.00000	0.3	0.32610	1.2	0.00000	0.1	0.00000	
EF1138		Unknown function	glutamine amidotransferase, class I	1	1	1	1	1	1	1	1	0.2	0.32610	1.2	0.00000	0.2	0.2	0.32610	1.2	0.00000	0.2	0.2	0.32610	0.6	0.00694
EF1139		Unknown function	glutamine amidotransferase, class I	1	1	1	1	1	1	1	1	0.4	0.85503	4.2	0.00000	0.4	0.4	0.85503	4.2	0.00000	0.4	0.4	0.85503	0.1	0.00001
EF1140	<i>glbA</i>	Energy metabolism	lactoylglutathione lyase	1	1	1	1	1	1	1	1	0.4	0.85503	4.2	0.00000	0.4	0.4	0.85503	4.2	0.00000	0.4	0.4	0.85503	0.1	0.00001
EF1141		DNA metabolism	MutT/nudix family protein	1	1	1	1	1	1	1	1	0.68960	1.7	0.00001	0.3	0.68960	1.7								

Locus	Gene	Functional category	Subcategory	M	T	O	S	T	M	I	S	q-value	M_130	q-value	O_15	q-value	O_30	q-value	S_15	q-value	S_30	q-value
EF1608		Fatty acid and phospholipid metabolism	Biosynthesis	1	1	1	1	1	0.7	0.05110	1.4	0.00000	1.4	0.00276	1.1	0.00276	0.4	0.17610	0.8	0.00009	1.4	0.00000
EF1609		Hypothetical proteins	Conserved	1	1	1	1	1	0.1	0.74200	0.3	0.03396	0.3	0.03396	-0.6	0.04077	-0.6	0.18030	0.8	0.00000	-0.2	0.48350
EF1610		Hypothetical proteins	Domain	1	1	1	1	1	-0.6	0.06988	-1.3	0.00000	-1.3	0.00000	0.1	0.64010	0.3	0.03452	-0.2	0.22270	-0.4	0.02756
EF1611	<i>ppaC</i>	Central intermediary metabolism	Phosphonous compounds	1	1	1	1	1	0.3	0.19040	-0.9	0.00021	-0.9	0.00021	0.4	0.03947	-0.4	0.03287	-0.2	0.18680	-0.4	0.05459
EF1612	<i>pflA</i>	Energy metabolism	Fermentation	1	1	1	1	1	0.1	0.68750	-2.7	0.00000	-2.7	0.00000	0.2	0.22200	-0.7	0.1521	-0.2	0.12660	-0.7	0.00064
EF1613	<i>pflB</i>	Energy metabolism	Fermentation	1	1	1	1	1	0.2	0.33810	-2.1	0.00000	-2.1	0.00000	0.4	0.02059	-0.4	0.12240	-0.2	0.07672	-0.6	0.00580
EF1614	<i>pflC</i>	DNA replication, recombination, and repair	DNA replication, recombination, and repair	1	1	1	1	1	0.7	0.00225	-1.7	0.00000	-1.7	0.00000	-0.3	0.33910	-0.1	0.87010	-0.3	0.05851	-0.9	0.00507
EF1615	<i>pflE</i>	DNA replication, recombination, and repair	DNA replication, recombination, and repair	1	1	1	1	1	-0.4	0.11750	-0.7	0.00000	-0.7	0.00000	0.4	0.16370	0.0	0.92900	-0.2	0.21660	-0.5	0.01238
EF1616		Unknown function	General	1	1	1	1	1	0.7	0.00734	1.5	0.00000	1.5	0.00000	NA	NA	NA	NA	NA	NA	NA	NA
EF1617		Hypothetical proteins	Conserved	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1618		Energy metabolism	Amino acids and amines	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1619		Cellular processes	Other	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1620		Hypothetical proteins	Conserved	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1621		Hypothetical proteins	Domain	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1622		Hypothetical proteins	Other	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1623		Central intermediary metabolism	Fermentation	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1624		Energy metabolism	General	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1625		Unknown function	General	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1626	<i>eulL</i>	Energy metabolism	Amino acids and amines	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1627	<i>eulC</i>	Energy metabolism	Amino acids and amines	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1629	<i>eulB</i>	Energy metabolism	Amino acids and amines	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1632		Signal transduction	Two-component systems	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1633		Signal transduction	Two-component systems	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1634		Unknown function	General	1	1	1	1	1	-0.5	0.05218	-1.1	0.00001	-1.1	0.00001	-0.5	0.09343	-0.7	0.00868	-0.4	0.05460	-0.4	0.33260
EF1635		Energy metabolism	Fermentation	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1637	<i>eupP</i>	Energy metabolism	Other	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1638		Unknown function	General	1	1	1	1	1	-0.2	0.33480	-0.4	0.06620	-0.4	0.06620	NA	NA	NA	NA	NA	NA	NA	NA
EF1639		Transport and binding proteins	Caltons and iron carrying compounds	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1640		Transport and binding proteins	Caltons and iron carrying compounds	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1641		Transport and binding proteins	Caltons and iron carrying compounds	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1643		Hypothetical proteins	Conserved	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1644		Unknown function	General	1	1	1	1	1	-1.4	0.00012	-1.7	0.00000	-1.7	0.00005	-0.1	0.00005	-0.5	0.06518	-1.4	0.00001	-1.5	0.00000
EF1645		Regulatory functions	DNA interactions	1	1	1	1	1	0.1	0.12570	2.2	0.00000	2.2	0.00000	0.7	0.00000	0.1	0.46320	0.1	0.46320	0.1	0.46320
EF1646	<i>codY</i>	Cellular processes	Adaptations to atypical conditions	1	1	1	1	1	0.8	0.00126	1.3	0.00008	1.3	0.00008	0.5	0.00512	0.7	0.01105	0.5	0.00486	1.0	0.00051
EF1647	<i>hslU</i>	Cellular processes	Protein folding and stabilization	1	1	1	1	1	0.2	0.00004	1.7	0.00000	1.7	0.00000	0.5	0.00526	0.6	0.01105	0.7	0.00445	0.9	0.00501
EF1648	<i>hslV</i>	Protein fold	DNA replication, recombination, and repair	1	1	1	1	1	1.5	0.00011	1.7	0.00000	1.7	0.00000	0.9	0.00006	0.6	0.05979	1.1	0.00003	0.6	0.00639
EF1649		DNA metabolism	General	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1649	<i>gid</i>	Unknown function	General	1	1	1	1	1	-0.6	0.00709	-0.3	0.06903	-0.3	0.06903	NA	NA	NA	NA	NA	NA	NA	NA
EF1660	<i>topA</i>	DNA metabolism	DNA replication, recombination, and repair	1	1	1	1	1	0.0	0.01377	-0.4	0.02783	-0.4	0.02783	-0.5	0.01358	0.0	0.86590	-0.2	0.44540	-0.1	0.63610
EF1661		Unknown function	General	1	1	1	1	1	0.0	0.93630	0.7	0.00011	0.7	0.00011	0.5	0.17390	1.1	0.00215	0.1	0.76200	0.8	0.02193
EF1662		Cellular processes	DNA transformation	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1663		Transcription	Degradation of rRNA	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1664		Unknown function	General	1	1	1	1	1	-0.4	0.06850	-0.8	0.00748	-0.8	0.00748	NA	NA	NA	NA	NA	NA	NA	NA
EF1665		Biosynthesis of cofactors, prosthetic groups, carriers	Pantothenate and coenzyme A	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1666		Regulatory functions	DNA interactions	1	1	1	1	1	-0.8	0.04429	-1.1	0.00665	-1.1	0.00665	-0.7	0.04154	-0.6	0.19560	-0.9	0.00010	-1.1	0.00011
EF1667		Cell envelope	Other	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1668	<i>bldC</i>	Energy metabolism	Amino acids and amines	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1669	<i>bldB</i>	Energy metabolism	Amino acids and amines	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1670	<i>bldA</i>	Energy metabolism	Amino acids and amines	1	1	1	1	1	0.1	0.68300	1.3	0.00001	1.3	0.00001	NA	NA	NA	NA	NA	NA	NA	NA
EF1671	<i>bldD</i>	Energy metabolism	Amino acids and amines	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1672	<i>buk</i>	Energy metabolism	Other	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1662	<i>pb</i>	Fatty acid and phospholipid metabolism	Degradation	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1664		Hypothetical proteins	Conserved	1	1	1	1	1	-0.7	0.01535	0.9	0.00004	0.9	0.00004	-0.7	0.00435	-0.4	0.36850	-0.7	0.00413	0.3	0.09860
EF1665		Hypothetical proteins	Conserved	1	1	1	1	1	-0.5	0.03698	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1666		Hypothetical proteins	Conserved	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1667		Unknown function	Enzymes of unknown specificity	1	1	1	1	1	1.1	0.00023	3.6	0.00000	3.6	0.00000	0.3	0.04448	2.2	0.00001	0.8	0.00006	2.9	0.00000
EF1668		Regulatory functions	DNA interactions	1	1	1	1	1	0.3	0.04853	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1669		Unknown function	Enzymes of unknown specificity	1	1	1	1	1	0.1	0.83610	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1670		Unknown function	Enzymes of unknown specificity	1	1	1	1	1	0.1	0.68300	1.3	0.00001	1.3	0.00001	-0.1	0.85690	0.5	0.13370	0.6	0.01039	NA	NA
EF1671		Unknown function	Enzymes of unknown specificity	1	1	1	1	1	0.1	0.68300	1.3	0.00001	1.3	0.00001	0.9	0.00084	1.6	0.0102	1.3	0.00000	NA	NA
EF1672		Transport and binding proteins	Unknown substrate	1	1	1	1	1	1.7	0.00000	3.4	0.00000	3.4	0.00000	0.9	0.00084	1.6	0.0102	1.3	0.00000	NA	NA
EF1673		Transport and binding proteins</																				

Locus	Gene	Functional category	Subcategory	Putative function	M	O	T	S	T	M	I	L	q-value	M	I	S	Q	Q	S	q-value	S	Q	q-value
EF1786	<i>purK-1</i>	Purines, pyrimidines, nucleosides, and nucleotides	Purine ribonucleotide biosynthesis	phosphoribosylaminimidazole carboxylase, ATPase subunit	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1787	<i>purE</i>	Purines, pyrimidines, nucleosides, and nucleotides	Purine ribonucleotide biosynthesis	phosphoribosylaminimidazole carboxylase, catalytic subunit	1	1	1	1	1	1	1	1	1.5	0.00012	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1789		Unknown function	General	SPFH domain/Band 7 family protein	1	1	1	1	1	1	1	1	0.6	0.00194	0.8	0.00194	0.6	0.09652	1.1	0.00005	NA	NA	0.00000
EF1790		Transport and binding proteins	General	phormone binding protein	1	1	1	1	1	1	1	1	0.4	0.00290	0.7	0.00177	0.1	0.67360	0.6	0.00276	1.0	NA	0.00052
EF1791		Transport and binding proteins	Conserved	conserved hypothetical protein	1	1	1	1	1	1	1	1	0.1	0.00380	0.5	0.00190	NA	NA	NA	NA	NA	NA	NA
EF1792		Amino acid biosynthesis	Conserved	branched-chain amino acid aminotransferase	1	1	1	1	1	1	1	1	-0.3	0.18830	-0.1	0.22110	-0.2	0.49250	-0.1	0.73340	0.1	NA	0.62400
EF1793	<i>ilvE</i>	Amino acid biosynthesis	Conserved	conserved hypothetical protein	1	1	1	1	1	1	1	1	1.2	0.00040	2.8	0.00000	0.5	0.12920	0.7	0.03425	NA	NA	NA
EF1794		Hypothetical proteins	Conserved	conserved hypothetical protein	1	1	1	1	1	1	1	1	0.5	0.06853	3.6	0.00000	0.2	0.43230	2.5	0.00000	0.5	0.00927	3.4
EF1797		Cell envelope	Other	lipoprotein, putative	1	1	1	1	1	1	1	1	-0.3	0.06840	-0.2	0.66276	-0.4	0.68370	0.1	0.69310	0.0	0.99310	0.2
EF1798		Hypothetical proteins	Conserved	conserved hypothetical protein	1	1	1	1	1	1	1	1	-0.2	0.35270	0.0	0.65000	-0.2	0.50310	0.5	0.03563	0.0	0.94560	0.0
EF1800		Hypothetical proteins	Conserved	conserved hypothetical protein	1	1	1	1	1	1	1	1	0.7	0.00717	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1801		Hypothetical proteins	PTS	PTS system, IIA component	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1802		Hypothetical proteins	PTS	PTS system, IIB component	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1803		Hypothetical proteins	PTS	PTS system, IIC component	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1804		Hypothetical proteins	PTS	PTS system, IID component	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1805		Energy metabolism	Biosynthesis and degradation of polysaccharides	glycosyl hydrolase, family 35	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1806	<i>lacC</i>	Energy metabolism	Biosynthesis and degradation of polysaccharides	tagatase-6-phosphate kinase	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1807	<i>lacD-2</i>	Energy metabolism	Biosynthesis and degradation of polysaccharides	tagatase 1,6-diphosphate aldolase	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1808		Energy metabolism	Enzymes of unknown specificity	agatS protein	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1809		Regulatory functions	DNA interactions	transcriptional regulator, GntR family	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1810	<i>gspA-1</i>	Regulatory functions	Adaptations to atypical conditions	general stress protein A	1	1	1	1	1	1	1	1	0.6	0.04494	2.1	0.00000	NA	NA	NA	NA	NA	NA	NA
EF1811	<i>gspA-2</i>	Regulatory functions	Adaptations to atypical conditions	general stress protein A	1	1	1	1	1	1	1	1	1.1	0.00023	4.1	0.00001	1.3	0.00001	0.5	0.00927	0.0	0.99310	0.2
EF1812		Hypothetical proteins	Other	hypothetical protein	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1813		Transport and binding proteins	General	sulfatase domain protein	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1814		Regulatory functions	DNA interactions	drug resistance transporter, EmrB/QacA family protein	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1815		Regulatory functions	Conserved	transcriptional regulator, LysR family, putative	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1816		Hypothetical proteins	Other	conserved hypothetical protein	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1817	<i>spoE</i>	Protein fate	Degradation of proteins, peptides, and glycopeptides	serine proteinase, V8 family	1	1	1	1	1	1	1	1	-0.1	0.46100	-0.1	0.56920	0.2	0.42790	0.2	0.32080	NA	NA	NA
EF1818	<i>gene</i>	Cell envelope	Other	coccolysin	1	1	1	1	1	1	1	1	-0.5	0.02068	0.2	0.15500	0.4	0.03869	NA	NA	NA	NA	NA
EF1820	<i>fsxC</i>	Signal transduction	Two-component systems	histidine kinase, putative	1	1	1	1	1	1	1	1	0.2	0.52410	1.0	0.00007	0.3	0.16920	0.2	0.34860	NA	NA	NA
EF1821	<i>fsxB</i>	Signal transduction	Two-component systems	agfB1s protein	1	1	1	1	1	1	1	1	0.2	0.20280	1.0	0.00001	0.4	0.13650	0.4	0.23300	NA	NA	NA
EF1822	<i>fsxA</i>	Signal transduction	Two-component systems	response regulator	1	1	1	1	1	1	1	1	0.7	0.00179	0.8	0.00002	0.4	0.13650	0.4	0.23300	NA	NA	NA
EF1824		Cell envelope	Biosynthesis and degradation of polysaccharides	N-acetyl/muramoyl-L-alanine amidase, family 4	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1825		Hypothetical proteins	Biosynthesis and degradation of polysaccharides	glycosyl hydrolase, family 31/fibrinectin type III domain protein	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1826		Energy metabolism	Domain	conserved domain protein	1	1	1	1	1	1	1	1	0.6	0.01791	0.1	0.56000	0.1	0.67370	NA	NA	NA	NA	NA
EF1827		Energy metabolism	Conserved	alcohol dehydrogenase, zinc-containing	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1828		Hypothetical proteins	Other	conserved hypothetical protein	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1829		Transport and binding proteins	Other	glycerol uptake facilitator protein, putative	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1830		Signal transduction	PTS	PTS system, IIC component	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1833		Hypothetical proteins	PTS	hypothetical protein	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1834	<i>lacB</i>	Energy metabolism	Biosynthesis and degradation of polysaccharides	galactose-6-phosphate isomerase, LacB subunit	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1835	<i>lacA</i>	Energy metabolism	Biosynthesis and degradation of polysaccharides	galactose-6-phosphate isomerase, LacA subunit	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1836		Signal transduction	PTS	PTS system, IIA component, putative	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1837		Signal transduction	PTS	PTS system, IIB component, putative	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1838		Signal transduction	PTS	PTS system, IIC component	1	1	1	1	1	1	1	1	0.5	0.12390	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1839		Regulatory functions	DNA interactions	lactose phosphotransferase system repressor LacR	1	1	1	1	1	1	1	1	0.3	0.28740	0.6	0.00665	0.1	0.78660	0.1	0.78660	NA	NA	NA
EF1841	<i>lacR</i>	Regulatory functions	DNA interactions	HD domain protein	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1843		Energy metabolism	Biosynthesis and degradation of polysaccharides	polysaccharide deacetylase family protein	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1844		Hypothetical proteins	Other	hypothetical protein	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1846		Hypothetical proteins	DNA replication, recombination, and repair	site-specific recombinase, phage integrase family	1	1	1	1	1	1	1	1	0.2	0.36640	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1847		DNA metabolism	Conserved	hypothetical protein	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1848		Hypothetical proteins	Conserved	conserved hypothetical protein	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1849		Hypothetical proteins	Biosynthesis and degradation of polysaccharides	conserved hypothetical protein	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1850		Hypothetical proteins	Conserved	conserved hypothetical protein	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1851		Energy metabolism	Biosynthesis and degradation of polysaccharides	glycosyl hydrolase, family 35	1	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1853		Hypothetical proteins	Transposon functions	hypothetical protein	1	1	1	1	1	1	1	1	0.0	0.15380	0.0	0.96190	NA	NA	NA	NA	NA	NA	NA
EF1855	<i>pand</i>	Mobile and extrachromosomal element functions	Transposon functions	transposase, IS256 family	1	1	1	1	1	1	1	1	0.0	0.17960	0.2	0.16200	NA	NA	NA	NA	NA	NA	NA
EF1856		Biosynthesis of cofactors, prosthetic groups, carriers	Pantothenate and coenzyme A	aspartate 1-decarboxylase	1	1	1	1	1	1	1	1	0.4	0.13910	-1.1	0.00000	NA	NA	NA	NA	NA	NA	NA
EF1859	<i>panC</i>	Biosynthesis of cofactors, prosthetic groups, carriers	Pantothenate and coenzyme A	panicate--beta-alanine ligase	1	1	1	1	1	1	1	1	0.3	0.23890	-0.9	0.00002	NA	NA	NA	NA	NA	NA	NA
EF1860	<i>panB</i>	Biosynthesis of cofactors, prosthetic groups, carriers	Pantothenate and coenzyme A	3-methyl-2-oxobutanoate hydroxymethyltransferase	1	1	1	1	1	1	1	1	0.4	0.03017	-0.4	0.01840	NA	NA	NA	NA	NA		

Locus	Gene	Functional category	Subcategory	M	O	S	T	M	I	S	q-value	M	I	S	q-value	O	S	q-value	S	I	S	q-value	S	I	S	q-value	
EF1887		Hypothetical proteins	Conserved	1	0	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1888		Hypothetical proteins	Conserved	1	0	0	0	0.39500	0.5	0.01350	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1889		Hypothetical proteins	Domain	1	0	0	0.2	0.03544	0.0	0.89100	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1890		Hypothetical proteins	Domain	1	1	0	0	0.00551	2.5	0.00000	1.3	0.00030	0.7	0.00165	0.8	0.00005	1.8	0.00005	1.6	0.00006	1.5	0.00000	1.5	0.00000	1.5	0.00000	1.5
EF1891		Cellular processes	Cell division	1	0	0	0	0.02110	1.4	0.00000	0.3	0.02946	0.6	0.00846	0.6	0.00026	1.0	0.00026	1.5	0.00000	1.5	0.00000	1.5	0.00000	1.5	0.00000	1.5
EF1892		Hypothetical proteins	Conserved	1	0	0	0	0.02526	0.5	0.02374	1.3	0.00171	1.0	0.00171	1.0	0.00426	0.6	0.00426	0.6	0.00426	0.6	0.00426	0.6	0.00426	0.6	0.00426	0.6
EF1893		Hypothetical proteins	Conserved	1	1	0	0	0.18330	0.9	0.00058	1.0	0.00426	0.6	0.00426	0.6	0.00426	0.6	0.00426	0.6	0.00426	0.6	0.00426	0.6	0.00426	0.6	0.00426	0.6
EF1894		Fatty acid and phospholipid metabolism	Degradation	1	1	0	0	0.18330	0.9	0.00058	1.0	0.00426	0.6	0.00426	0.6	0.00426	0.6	0.00426	0.6	0.00426	0.6	0.00426	0.6	0.00426	0.6	0.00426	0.6
EF1895		Hypothetical proteins	Conserved	1	1	0	0	0.04904	0.1	0.44800	0.6	0.00317	0.1	0.44800	0.6	0.00317	0.1	0.44800	0.6	0.00317	0.1	0.44800	0.6	0.00317	0.1	0.44800	0.6
EF1896		Cell envelope	Other	1	0	1	0	0.52290	0.7	0.00013	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1897	<i>ripS</i>	Hypothetical proteins	Ribosomal proteins: synthesis and modification	1	1	0	0	0.16770	0.8	0.00047	-1.3	0.00002	0.6	0.18720	-0.4	0.00414	-0.9	0.00414	-0.9	0.00414	-0.9	0.00414	-0.9	0.00414	-0.9	0.00414	-0.9
EF1898	<i>rimD</i>	Protein synthesis	RNA and rRNA base modification	1	1	0	0	0.91310	-0.5	0.00088	-0.2	0.12650	-0.1	0.59310	-0.3	0.03326	-0.4	0.03326	-0.4	0.03326	-0.4	0.03326	-0.4	0.03326	-0.4	0.03326	-0.4
EF1900	<i>rimM</i>	Transcription	RNA processing	1	1	0	0	0.03544	0.0	0.89100	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1901		Transport and binding proteins	Cations and iron carrying compounds	1	1	1	0	0.00551	2.5	0.00000	1.3	0.00030	0.7	0.00165	0.8	0.00005	1.8	0.00005	1.6	0.00006	1.5	0.00000	1.5	0.00000	1.5	0.00000	1.5
EF1902		Unknown function	Enzymes of unknown specificity	1	1	0	0	0.02110	1.4	0.00000	0.3	0.02946	0.6	0.00846	0.6	0.00026	1.0	0.00026	1.5	0.00000	1.5	0.00000	1.5	0.00000	1.5	0.00000	1.5
EF1903		Hypothetical proteins	Conserved	1	1	0	0	0.02526	0.5	0.02374	1.3	0.00171	1.0	0.00171	1.0	0.00426	0.6	0.00426	0.6	0.00426	0.6	0.00426	0.6	0.00426	0.6	0.00426	0.6
EF1904		Fatty acid and phospholipid metabolism	Degradation	1	1	0	0	0.18330	0.9	0.00058	1.0	0.00426	0.6	0.00426	0.6	0.00426	0.6	0.00426	0.6	0.00426	0.6	0.00426	0.6	0.00426	0.6	0.00426	0.6
EF1906		Hypothetical proteins	Conserved	1	1	0	0	0.18330	0.9	0.00058	1.0	0.00426	0.6	0.00426	0.6	0.00426	0.6	0.00426	0.6	0.00426	0.6	0.00426	0.6	0.00426	0.6	0.00426	0.6
EF1907		Unknown function	General	1	1	0	0	0.04904	0.1	0.44800	0.6	0.00317	0.1	0.44800	0.6	0.00317	0.1	0.44800	0.6	0.00317	0.1	0.44800	0.6	0.00317	0.1	0.44800	0.6
EF1908	<i>murC</i>	Cell envelope	Biosynthesis/degradation of murein sacculus/peptidoglycan	1	1	0	0	0.52290	0.7	0.00013	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1909		Hypothetical proteins	Conserved	1	1	1	0	0.34670	0.8	0.00002	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1910		Hypothetical proteins	Conserved	1	1	0	0	0.34670	0.8	0.00002	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1911		Hypothetical proteins	Conserved	1	1	0	0	0.38920	0.4	0.02158	0.2	0.44550	0.7	0.09135	0.2	0.44870	0.0	0.44870	0.0	0.44870	0.0	0.44870	0.0	0.44870	0.0	0.44870	0.0
EF1912		Unknown function	General	1	1	0	0	0.23890	-0.6	0.00311	-0.1	0.55970	-0.3	0.28030	-0.2	0.06230	-0.2	0.06230	-0.2	0.06230	-0.2	0.06230	-0.2	0.06230	-0.2	0.06230	-0.2
EF1913		Hypothetical proteins	Conserved	1	1	0	0	0.27580	0.0	0.99160	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1914		Hypothetical proteins	Conserved	1	1	0	0	0.27580	0.0	0.99160	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1915		Hypothetical proteins	Conserved	1	1	0	0	0.27580	0.0	0.99160	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1916		Unknown function	General	1	1	0	0	0.00125	1.1	0.00001	0.3	0.14120	0.5	0.01772	0.6	0.00665	1.5	0.00000	1.5	0.00000	1.5	0.00000	1.5	0.00000	1.5	0.00000	1.5
EF1917		Protein fate	Degradation of proteins, peptides, and glycopeptides	1	1	0	0	0.23980	-0.9	0.00008	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1918		Hypothetical proteins	Conserved	1	1	0	0	0.16360	-0.2	0.09392	-0.6	0.00917	-0.1	0.66400	-0.4	0.02711	-0.1	0.66400	-0.4	0.02711	-0.1	0.66400	-0.4	0.02711	-0.1	0.66400	-0.4
EF1919		Unknown function	Conserved	1	1	0	0	0.22680	-0.1	0.28760	-0.1	0.39230	-0.4	0.07876	-0.3	0.04376	-0.1	0.04376	-0.1	0.04376	-0.1	0.04376	-0.1	0.04376	-0.1	0.04376	-0.1
EF1920		Transport and binding proteins	Enzymes of unknown specificity	1	1	0	0	0.00424	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1921		Carbohydrates, nucleosides, and nucleotides	Carbohydrates, organic alcohols, and acids	1	1	0	0	0.04481	-0.8	0.00264	-0.4	0.03934	-0.1	0.77110	0.9	0.00076	0.7	0.00076	0.7	0.00076	0.7	0.00076	0.7	0.00076	0.7	0.00076	0.7
EF1922		Regulatory functions	Salvage of nucleosides and nucleotides	1	1	0	0	0.98020	0.1	0.36430	0.1	0.41220	0.4	0.60220	0.4	0.04672	0.2	0.04672	0.2	0.04672	0.2	0.04672	0.2	0.04672	0.2	0.04672	0.2
EF1923		Hypothetical proteins	DNA interactions	1	1	0	0	0.14720	0.4	0.02688	0.4	0.01684	-0.1	0.60680	0.6	0.03089	0.6	0.03089	0.6	0.03089	0.6	0.03089	0.6	0.03089	0.6	0.03089	0.6
EF1925		Hypothetical proteins	Conserved	1	1	0	0	0.06229	0.0	0.76590	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1926		Hypothetical proteins	Conserved	1	1	0	0	0.16860	0.4	0.00710	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1927		Transport and binding proteins	Other	1	1	0	0	0.00001	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1928		Energy metabolism	Other	1	1	1	0	0.00001	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1929		Energy metabolism	Other	1	1	1	0	0.00073	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1930		Hypothetical proteins	Conserved	1	1	0	0	0.00832	0.4	0.01293	0.1	0.70680	0.2	0.44650	0.6	0.00893	0.9	0.00893	0.9	0.00893	0.9	0.00893	0.9	0.00893	0.9	0.00893	0.9
EF1932		Hypothetical proteins	Conserved	1	1	0	0	0.02526	2.8	0.00000	-1.0	0.00023	1.9	0.00005	-0.6	0.04091	3.2	0.00000	3.2	0.00000	3.2	0.00000	3.2	0.00000	3.2	0.00000	3.2
EF1933		Hypothetical proteins	Conserved	1	1	0	0	0.06350	1.8	0.00000	-1.0	0.00168	1.0	0.00017	-0.5	0.00527	2.0	0.00000	2.0	0.00000	2.0	0.00000	2.0	0.00000	2.0	0.00000	2.0
EF1934		Hypothetical proteins	Conserved	1	1	0	0	0.38750	0.5	0.00694	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1935		Hypothetical proteins	Conserved	1	1	0	0	0.93950	0.2	0.07829	-0.6	0.00042	-0.1	0.72280	0.1	0.75340	NA	0.75340	NA	0.75340	NA	0.75340	NA	0.75340	NA	0.75340	NA
EF1936		Hypothetical proteins	Conserved	1	1	0	0	0.00135	-0.4	0.01517	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF1937		Transport and binding proteins	Cations and iron carrying compounds	1	1	0	0	0.00066	1.7	0.00000	0.9	0.00078	1.4	0.00054	0.6	0.006											

Locus	Gene	Functional category	Subcategory	M	O	T	S	T	M	I	S	M	I	S	O	S	I	S	O	S	I	S	O	q-value	S	I	S	O	q-value
EF2074	<i>e6C</i>	Transport and binding proteins	Unknown substrate	1	1	1	1	1	2.8	0.00000	3.0	0.00000	3.1	0.00000	3.1	0.00000	3.7	0.00000	3.4	0.00000	3.4	0.00000	3.4	0.00000	3.4	0.00000	3.4	0.00000	
EF2075	<i>e6B</i>	Transport and binding proteins	Unknown substrate	1	1	1	1	1	2.7	0.00004	3.5	0.00000	3.0	0.00000	3.0	0.00000	3.6	0.00000	3.4	0.00000	3.4	0.00000	3.4	0.00000	3.4	0.00000	3.4	0.00000	
EF2076	<i>e6A</i>	Cellular processes	Cell adhesion	1	1	1	1	1	2.3	0.00004	2.9	0.00000	2.8	0.00001	2.8	0.00001	3.0	0.00000	3.2	0.00000	3.2	0.00000	3.2	0.00000	3.2	0.00000	3.2	0.00000	
EF2077		Transport and binding proteins	Unknown substrate	1	1	1	1	1	0.2	0.26870	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2078		Hypothetical proteins	Degradation of proteins, peptides, and glycopeptides	1	1	1	1	1	0.2	0.26770	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2079		Protein fate	Other	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2080		Cell envelope	Other	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2081		Transport and binding proteins	Unknown substrate	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2082		Transport and binding proteins	Unknown substrate	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2083		Hypothetical proteins	Unknown substrate	1	1	1	1	1	0.5	0.04413	0.0	0.68480	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2084		Hypothetical proteins	Unknown substrate	1	1	1	1	1	-0.5	0.99860	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2085		Hypothetical proteins	Conserved	1	0	0	0	0	0.0	0.00007	-0.9	0.00040	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2086		Hypothetical proteins	Prophage functions	1	0	0	0	0	-1.3	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2087		Mobile and extrachromosomal element functions	Prophage functions	1	0	0	0	0	-0.4	0.14360	0.0	0.74230	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2088		Hypothetical proteins	Prophage functions	1	0	0	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2089		Hypothetical proteins	Prophage functions	1	0	0	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2090		Hypothetical proteins	Prophage functions	1	0	0	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2091		Hypothetical proteins	Prophage functions	1	0	0	0	0	-0.1	0.73320	-0.2	0.14210	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2092		Hypothetical proteins	Prophage functions	1	0	0	0	0	-0.2	0.18830	-0.6	0.00272	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2093		Unknown function	General	1	0	0	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2094		Hypothetical proteins	General	1	0	0	0	0	-0.2	0.27710	-0.6	0.00323	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2095		Hypothetical proteins	General	1	0	0	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2096		Mobile and extrachromosomal element functions	Prophage functions	1	0	0	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2097		Mobile and extrachromosomal element functions	Prophage functions	1	0	0	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2098		Hypothetical proteins	Prophage functions	1	0	0	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2099		Hypothetical proteins	Prophage functions	1	0	0	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2100		Hypothetical proteins	Prophage functions	1	0	0	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2101		Hypothetical proteins	Prophage functions	1	0	0	0	0	-0.3	0.16500	-0.6	0.00018	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2102		Hypothetical proteins	Prophage functions	1	0	0	0	0	-0.3	0.20880	-0.7	0.00013	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2103		Hypothetical proteins	Prophage functions	1	0	0	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2104		Hypothetical proteins	Prophage functions	1	0	0	0	0	0.1	0.76950	-0.7	0.00575	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2105		Hypothetical proteins	Prophage functions	1	0	0	0	0	0.0	0.68680	-0.8	0.00156	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2106		Hypothetical proteins	Prophage functions	1	0	0	0	0	0.0	0.97030	-0.6	0.00079	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2107		Hypothetical proteins	Prophage functions	1	0	0	0	0	0.0	0.82580	-0.4	0.00579	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2108		Hypothetical proteins	Prophage functions	1	0	0	0	0	-0.1	0.67170	-0.5	0.01507	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2109		Hypothetical proteins	Prophage functions	1	0	0	0	0	-0.1	0.67170	-0.5	0.01507	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2110		Hypothetical proteins	Prophage functions	1	0	0	0	0	-0.2	0.34900	-0.5	0.00104	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2111		Hypothetical proteins	Prophage functions	1	0	0	0	0	0.0	0.95750	-0.1	0.44600	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2112		Hypothetical proteins	Prophage functions	1	0	0	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2113		Hypothetical proteins	Prophage functions	1	0	0	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2114		Mobile and extrachromosomal element functions	Prophage functions	1	0	0	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2115		Regulatory functions	DNA interactions	1	0	0	1	1	-0.1	0.73000	0.2	0.18420	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2116		Hypothetical proteins	Conserved	1	0	0	1	1	0.1	0.90200	0.2	0.00442	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2117		Hypothetical proteins	Conserved	1	0	0	1	1	0.1	0.82120	-0.6	0.00688	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2118		Hypothetical proteins	Conserved	1	0	0	1	1	0.0	0.96790	-0.4	0.00688	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2119		Hypothetical proteins	Conserved	1	0	0	1	1	0.1	0.86970	0.1	0.46620	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2120		Hypothetical proteins	Conserved	1	0	0	1	1	0.1	0.79050	0.2	0.27990	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2121		Hypothetical proteins	Conserved	1	0	0	1	1	0.0	0.98500	-0.2	0.15810	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2122		Hypothetical proteins	Conserved	1	0	0	1	1	0.0	0.94280	-0.5	0.01618	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2123		Hypothetical proteins	Prophage functions	1	0	0	1	1	0.0	0.49860	-0.7	0.00011	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2124		Mobile and extrachromosomal element functions	Prophage functions	1	0	0	1	1	-0.1	0.75690	0.0	0.87650	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2125		Hypothetical proteins	Prophage functions	1	0	0	1	1	0.1	0.75920	0.2	0.06500	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2126		Hypothetical proteins	Prophage functions	1	0	0	1	1	0.1	0.64720	0.1	0.35970</																	

Locus	Gene	Functional category	Subcategory	MF	TO	ST	M	IS	q-value	M_130	q-value	O	IS	q-value	O_130	q-value	S	IS	q-value	S_130	q-value	
EF2333		Hypothetical proteins	Putative function	0	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF2334		Hypothetical proteins	hypothetical protein	0	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF2335		Hypothetical proteins	conserved domain protein	1	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF2336		Hypothetical proteins	conserved	1	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF2337		Hypothetical proteins	Conserved	1	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF2338		Hypothetical proteins	Conserved	1	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF2339		Regulatory functions	hypothetical protein	1	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF2340		Hypothetical proteins	transcriptional regulator, Cro/C1 family	1	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF2341		Hypothetical proteins	hypothetical protein	1	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF2342		Hypothetical proteins	C-5 cytosine-specific DNA methylase	1	0	0	-0.2	0.32840	-0.9	0.0001												
EF2343		Hypothetical proteins	hypothetical protein	1	0	0	-0.1	0.61780	-0.4	0.0376												
EF2344		Cellular processes	hypothetical protein	1	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF2345		Hypothetical proteins	FISKSP01E family protein	1	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF2346		Hypothetical proteins	hypothetical protein	1	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF2347		Hypothetical proteins	conserved hypothetical protein	1	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF2348		Hypothetical proteins	conserved hypothetical protein	1	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF2349		Hypothetical proteins	cell wall surface anchor family protein	1	0	0	0.0	0.99260	0.0	0.80050												
EF2350		Regulatory functions	hypothetical protein	1	0	0	0.1	0.43780	0.5	0.00220												
EF2351		Hypothetical proteins	transcriptional regulator, Cro/C1 family	1	0	0	0.3	0.19230	1.0	0.0002												
EF2352		Unknown function	hypothetical protein	1	0	0	-0.7	0.00235	-1.2	0.00100												
EF2353	<i>lepA</i>	Unknown function	GTP-binding protein LepA	1	1	1	-0.1	0.63630	-0.6	0.01160												
EF2354		Hypothetical proteins	acetyltransferase, GNAT family	1	1	1	-0.2	0.31350	-0.3	0.01411												
EF2355	<i>cipB</i>	Protein fate	conserved hypothetical protein	1	1	1	0.2	0.21830	0.2	0.04382												
EF2357		Hypothetical proteins	Degradation of proteins, peptides, and glycopeptides	1	1	1	1.5	0.00000	2.1	0.00000												
EF2358		Hypothetical proteins	conserved hypothetical protein	1	1	1	-0.4	0.03312	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF2360		Hypothetical proteins	hypothetical protein	1	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF2361	<i>purB</i>	Purines, pyrimidines, nucleosides, and nucleotides	adenylosuccinate lyase	1	1	1	-0.5	0.01875	-0.8	0.00003												
EF2362	<i>purK-2</i>	Purines, pyrimidines, nucleosides, and nucleotides	phosphoribosylaminimidazole carboxylase, ATPase subunit	1	1	1	-0.2	0.23550	0.0	0.81690												
EF2363		Hypothetical proteins	hypothetical protein	1	1	1	0.0	0.88090	0.4	0.01342												
EF2364		Transport and binding proteins	xanthine permease	1	1	1	-0.2	0.4920	-1.1	0.01508												
EF2365	<i>xpt</i>	Purines, pyrimidines, nucleosides, and nucleotides	salvage of nucleosides and nucleotides	1	1	1	0.0	0.91380	0.4	0.02664												
EF2366		Hypothetical proteins	Conserved	1	1	1	0.8	0.01105	0.5	0.00005												
EF2367		Cell envelope	N-acetylmuramoyl-L-alanine amidase, family 4	1	1	1	0.8	0.00056	0.8	0.00389												
EF2368		Hypothetical proteins	hypothetical protein	1	1	1	0.9	0.00175	1.3	0.00000												
EF2369		Hypothetical proteins	hypothetical protein	1	1	1	0.1	0.00032	0.3	0.00000												
EF2370		Unknown function	collodolactase, GfoIyH/MocA family	1	1	1	0.3	0.19740	0.2	0.16980												
EF2371	<i>asgS</i>	Protein synthesis	asparaginyl-HRNA synthetase	1	1	1	-0.1	0.76730	0.6	0.05065												
EF2372	<i>asgB</i>	Amino acid biosynthesis	aspartate aminotransferase	1	1	1	-0.3	0.28100	0.1	0.64160												
EF2373		Hypothetical proteins	conserved	1	1	1	-0.3	0.08223	0.3	0.02719												
EF2374		Hypothetical proteins	DNA replication, recombination, and repair	1	1	1	-0.3	0.09423	-0.6	0.00119												
EF2376		Hypothetical proteins	hypothetical protein	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF2377		Transport and binding proteins	amino acid permease family protein	1	1	1	-0.6	0.00180	-3.3	0.00000												
EF2378		DNA metabolism	DNA replication, recombination, and repair	1	1	1	0.1	0.13040	-0.6	0.00056												
EF2379	<i>proS</i>	Protein synthesis	prolyl-HRNA synthetase, alpha subunit, Gram-positive type	1	1	1	0.1	0.58430	0.3	0.02659												
EF2380		Protein fate	Degradation of proteins, peptides, and glycopeptides	1	1	1	0.4	0.04814	0.2	0.15110												
EF2381		Hypothetical proteins	hypothetical protein	1	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF2382		Energy metabolism	glucose 1-dehydrogenase	1	1	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF2383		Hypothetical proteins	conserved hypothetical protein	1	1	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF2384		Hypothetical proteins	hypothetical protein	1	1	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF2385		Hypothetical proteins	hypothetical protein	1	0	0	0.0	0.82380	-0.2	0.02850												
EF2386		Hypothetical proteins	hypothetical protein	1	0	0	0.3	0.04481	0.3	0.06573												
EF2387		Cellular processes	chromosome partitioning ATPase, Para family	1	0	0	0.3	0.06284	0.0	0.93370												
EF2388		Hypothetical proteins	hypothetical protein	1	1	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF2389		Hypothetical proteins	hypothetical protein	1	1	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF2390		Hypothetical proteins	conserved hypothetical protein	1	1	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF2391		Unknown function	NiU family protein	1	1	0	0.0	0.80400	1.0	0.00003												
EF2392		Unknown function	amino transferase, class V	1	1	1	-0.3	0.20320	0.9	0.00019												
EF2393		Hypothetical proteins	conserved hypothetical protein	1	1	1	-0.2	0.31870	0.4	0.01119												
EF2394		Transport and binding proteins	ABC transporter, ATP-binding protein	1	1	1	-0.2	0.96520	0.1	0.04520												
EF2395	<i>lr</i>	Protein synthesis	ribosome recycling factor	1	1	1	-0.2	0.19280	-2.5	0.00000												
EF2396	<i>pyrH</i>	Purines, pyrimidines, nucleosides, and nucleotides	uridylate kinase	1	1	1	0.0	0.83330	-2.6	0.00000												
EF2397	<i>tsf</i>	Protein synthesis	translation elongation factor Ts	1	1	1	-0.2	0.13730	-3.0	0.00000												
EF2398	<i>rpsB</i>	Protein synthesis	ribosomal protein S2	1	1	1	-0.1	0.69650	-2.1	0.00000												
EF2399		Unknown function	acyltransferase, GNAT family	1	1	1	0.4	0.08774	0.0	0.88340												
EF2400		Protein synthesis	RNA methyltransferase, TrmH family	1	1	1	0.3	0.23120	-0.1	0.66650												
EF2401	<i>aypP</i>	Fatty acid and phospholipid metabolism	acylphosphatase	1	1	1	1.1	0.00013	2.5	0.00000												
EF2404		Hypothetical proteins	hypothetical protein	1	1	1	-0.4	0.01392	0.1	0.25200												
EF2405		Hypothetical proteins	hypothetical protein	1	1	1	0.1	0.74712	-1.5	0.00000												
EF2406	<i>gls</i>	Protein synthesis	glycyl-HRNA synthetase, beta subunit	1	1	1	0.1	0.58630	-1.4	0.00000												
EF2407	<i>gIQ</i>	Protein synthesis	glycyl-HRNA synthetase, alpha subunit	1	1	1	0.0	0.94380	-1.5	0.00001												
EF2408		Hypothetical proteins	hypothetical protein	1	1	1	-0.3	0.07838	-1.0	0.00002												
EF2409	<i>era</i>	DNA metabolism	DNA repair protein RecO, putative	1	1	1	-0.1	0.59200	-0.8	0.00001												
EF2410		Fatty acid and phospholipid metabolism	GTP-binding protein Era	1	1	1	-0.1	0.60450	-0.1	0.63940												
EF2411	<i>dgaA</i>	Hypothetical proteins	diacylglycerol kinase	1	1	1	-0.3	0.05057	-0.7	0.00035												
EF2412		Unknown function	conserved hypothetical protein TIGR00043	1	1	1	-0.2	0.23350	-0.3	0.03315												
EF2413		Unknown function	HD domain protein	1	1	1	-0.2	0.33510	-0.3	0.03151												
EF2414	<i>phoH</i>	Unknown function	phoH-like protein	1	1	1	-0.2	0.18010	-0.3	0.02509												
EF2415		Protein synthesis	conserved hypothetical protein	1	1	1	0.5	0.00445	1.4	0.00001												
EF2416	<i>rpsU</i>	Protein synthesis	ribosomal protein S21	1	1	1	0.0	0.83420	0.9	0.00004												
EF2417	<i>zur</i>	Regulatory functions	transcriptional regulator, Fur family	1	1	1	0.4	0.01875	0.4	0.01702												
EF2419		Hypothetical proteins	conserved hypothetical protein	1	1	1																

Locus	Gene	Functional category	Subcategory	M	O	T	S	T	M	I	S	q-value	M	I	S	q-value	O	I	S	q-value	S	I	S	q-value	I	S	q-value	
EF2423		Regulatory functions	DNA interactions	1	1	1	1	1	0.7	0.00126	0.9	0.00022	0.3	0.07198	0.3	0.07198	0.3	0.23100	0.6	0.00126	1.1	0.00001	0.00001	0.00001	0.00001	0.00001	0.00001	0.00001
EF2424		Amino acid biosynthesis	Glutamate family	1	1	1	1	1	-0.6	0.00186	0.2	0.07024	-0.1	0.25950	-0.3	0.25950	-0.3	0.23100	0.6	0.00126	1.1	0.00001	0.00001	0.00001	0.00001	0.00001	0.00001	0.00001
EF2425		Energy metabolism	Sugars	1	1	1	1	1	-0.3	0.31130	-0.3	0.02174	-0.1	0.96080	-0.3	0.96080	-0.3	0.23100	0.6	0.00126	1.1	0.00001	0.00001	0.00001	0.00001	0.00001	0.00001	0.00001
EF2426		Regulatory functions	DNA interactions	1	1	1	1	1	0.7	0.00469	0.4	0.00469	0.2	0.27660	0.2	0.27660	0.2	0.41750	0.1	0.00001	0.00001	0.00001	0.00001	0.00001	0.00001	0.00001	0.00001	
EF2427		Hypothetical proteins		1	1	1	1	1	-0.2	0.00477	-0.2	0.02699	-0.8	0.00025	-0.4	0.00025	-0.4	0.42820	0.1	0.00001	0.00001	0.00001	0.00001	0.00001	0.00001	0.00001	0.00001	
EF2428		Purines, pyrimidines, nucleosides, and nucleotides	DNA interactions	1	1	1	1	1	0.9	0.00009	1.0	0.00001	1.3	0.00003	1.3	0.00003	1.3	0.00003	0.4	0.07112	1.2	0.00005	0.2	0.00005	0.2	0.00005	0.2	
EF2429	<i>guaC</i>	Transport and binding proteins	Nucleotides and nucleoside interconversions	1	1	1	1	1	1.5	0.00006	0.1	0.04060	0.1	0.00018	0.1	0.00018	0.1	0.81250	2.2	0.00005	0.2	0.00005	0.2	0.00005	0.2	0.00005	0.2	
EF2430		Unknown function	Enzymes of unknown specificity	1	1	1	1	1	1.1	0.00011	-0.3	0.04768	1.1	0.00153	-0.6	0.01630	-0.6	0.16870	1.3	0.00007	-0.1	0.00007	-0.1	0.00007	-0.1	0.00007	-0.1	
EF2431		Unknown function	Enzymes of unknown specificity	1	1	1	1	1	1.2	0.00009	NA	NA	1.3	0.00020	NA	NA	1.3	0.00020	2.1	0.00000	0.1	0.00000	0.1	0.00000	0.1	0.00000	0.1	
EF2432		Energy metabolism	Enzymes of unknown specificity	1	1	1	1	1	0.2	0.30090	0.9	0.00036	0.4	0.09471	0.7	0.09471	0.7	0.16412	0.4	0.03684	0.8	0.00032	0.8	0.00032	0.8	0.00032	0.8	
EF2433		Regulatory functions	Glycolysis/gluconeogenesis	1	1	1	1	1	0.2	0.23310	1.1	0.00000	0.3	0.44370	0.5	0.44370	0.5	0.23370	0.5	0.01867	0.9	0.00703	0.9	0.00703	0.9	0.00703	0.9	
EF2434		Signal transduction	DNA interactions	1	1	1	1	1	0.3	0.18370	0.1	0.49180	0.2	0.47310	0.0	0.47310	0.0	0.84620	0.0	0.03366	0.2	0.28640	0.2	0.28640	0.2	0.28640	0.2	
EF2435		Unknown function	General	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	0.13780	0.2	0.33040	-0.3	0.21580	-0.3	0.21580	-0.3	0.21580	-0.3	
EF2436		Hypothetical proteins	Conserved	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	0.13780	0.2	0.33040	-0.3	0.21580	-0.3	0.21580	-0.3	0.21580	-0.3	
EF2437		Hypothetical proteins	Conserved	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	0.13780	0.2	0.33040	-0.3	0.21580	-0.3	0.21580	-0.3	0.21580	-0.3	
EF2438		Signal transduction	PTS	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	0.13780	0.2	0.33040	-0.3	0.21580	-0.3	0.21580	-0.3	0.21580	-0.3	
EF2439		Cellular processes	Toxin production and resistance	1	1	1	1	1	-1.4	0.00030	-1.3	0.00000	-1.5	0.00009	-1.5	0.00009	-1.5	0.28260	-1.5	0.00000	-1.8	0.00001	-1.8	0.00001	-1.8	0.00001	-1.8	
EF2440		Unknown function	General	1	1	1	1	1	-0.6	0.00822	-2.4	0.00000	-2.4	0.00000	-2.4	0.00000	-2.4	0.24350	-1.6	0.00080	-1.3	0.00080	-1.3	0.00080	-1.3	0.00080	-1.3	
EF2441		Hypothetical proteins	Conserved	1	1	1	1	1	-0.6	0.01123	-2.0	0.00000	-2.0	0.00000	-2.0	0.00000	-2.0	0.05158	-1.3	0.00084	-0.8	0.00084	-0.8	0.00084	-0.8	0.00084	-0.8	
EF2442		Transport and binding proteins	Anions	1	1	1	1	1	-0.6	0.00019	-1.7	0.00000	-1.7	0.00000	-1.7	0.00000	-1.7	0.01199	-1.1	0.00002	-1.5	0.00002	-1.5	0.00002	-1.5	0.00002	-1.5	
EF2443		Protein synthesis	Ribosomal proteins; synthesis and modification	1	1	1	1	1	-0.6	0.002078	-0.8	0.00056	-1.3	0.00001	-1.3	0.00001	-1.3	0.04880	-0.6	0.00008	-0.6	0.00008	-0.6	0.00008	-0.6	0.00008	-0.6	
EF2444	<i>rpsT</i>	Fatty acid and phospholipid metabolism	Other	1	1	1	1	1	-0.6	0.00480	-1.9	0.00000	-1.4	0.00001	-1.4	0.00001	-1.4	0.49650	-0.8	0.00008	-1.6	0.00008	-1.6	0.00008	-1.6	0.00008	-1.6	
EF2445		Biosynthesis of cofactors, prosthetic groups, carriers	Pantothenate and coenzyme A	1	1	1	1	1	0.9	0.00440	1.6	0.00005	1.1	0.00028	1.1	0.00028	1.1	0.00278	0.3	0.23380	0.2	0.00470	0.2	0.00470	0.2	0.00470	0.2	
EF2446		Hypothetical proteins	Conserved	1	1	1	1	1	-0.6	0.00799	-0.4	0.00146	-0.5	0.03692	-0.4	0.03692	-0.4	0.06712	-0.6	0.00488	-0.9	0.00488	-0.9	0.00488	-0.9	0.00488	-0.9	
EF2447		Cellular processes	DNA transformation	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	0.34460	NA	NA	NA	NA	NA	NA	NA	NA	NA	
EF2448		Cellular processes	DNA transformation	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	0.34460	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF2449		Cellular processes	DNA transformation	1	1	1	1	1	-0.7	0.02193	0.4	0.00171	-0.5	0.02217	-0.1	0.02217	-0.1	0.60410	-0.7	0.00232	0.2	0.41100	0.2	0.41100	0.2	0.41100	0.2	
EF2450		Unknown function	General	1	1	1	1	1	-0.4	0.16400	-1.1	0.00016	-1.1	0.50560	-1.1	0.50560	-1.1	0.47510	-0.2	0.24200	-0.3	0.06493	-0.3	0.06493	-0.3	0.06493	-0.3	
EF2451		Biosynthesis of cofactors, prosthetic groups, carriers	Pantothenate and coenzyme A	1	1	1	1	1	-0.4	0.23700	-0.8	0.00003	-0.2	0.26900	-0.3	0.26900	-0.3	0.12630	-0.1	0.47040	0.1	0.96720	0.1	0.96720	0.1	0.96720	0.1	
EF2452		Unknown function	Enzymes of unknown specificity	1	1	1	1	1	-0.5	0.02081	-0.7	0.00019	-0.1	0.75200	0.0	0.75200	0.0	0.96650	-0.3	0.02594	0.1	0.48970	0.1	0.48970	0.1	0.48970	0.1	
EF2453		Hypothetical proteins	Conserved	1	1	1	1	1	-0.5	0.17450	-0.2	0.06155	-0.4	0.04760	-0.4	0.04760	-0.4	0.94760	-0.3	0.00761	0.1	0.89770	0.1	0.89770	0.1	0.89770	0.1	
EF2454		Hypothetical proteins	Conserved	1	1	1	1	1	-0.5	0.52750	-0.3	0.14990	-0.4	0.10701	-0.2	0.10701	-0.2	0.45800	-0.3	0.05902	0.1	0.85340	0.1	0.85340	0.1	0.85340	0.1	
EF2455		Hypothetical proteins	Conserved	1	1	1	1	1	-0.5	0.34040	-0.6	0.01718	-0.1	0.72310	-0.2	0.72310	-0.2	0.62090	-0.4	0.00219	-0.4	0.07519	-0.4	0.07519	-0.4	0.07519	-0.4	
EF2456		Energy metabolism	Glycolysis/gluconeogenesis	1	1	1	1	1	-0.2	0.04310	-1.0	0.00548	-0.1	0.95600	-0.2	0.95600	-0.2	0.49650	-0.4	0.06354	-0.4	0.15700	-0.4	0.15700	-0.4	0.15700	-0.4	
EF2457		Cellular processes	Cell division	1	1	1	1	1	-0.2	0.17900	-0.6	0.00449	-0.3	0.89036	-0.3	0.89036	-0.3	0.07768	-0.3	0.05864	-0.5	0.12040	-0.5	0.12040	-0.5	0.12040	-0.5	
EF2458		Hypothetical proteins	Conserved	1	1	1	1	1	-0.3	0.26430	-0.5	0.00449	-0.3	0.89036	-0.3	0.89036	-0.3	0.07768	-0.3	0.05864	-0.5	0.12040	-0.5	0.12040	-0.5	0.12040	-0.5	
EF2459		Hypothetical proteins	Conserved	1	1	1	1	1	-0.3	0.00093	-0.5	0.00449	-0.3	0.89036	-0.3	0.89036	-0.3	0.07768	-0.3	0.05864	-0.5	0.12040	-0.5	0.12040	-0.5	0.12040	-0.5	
EF2460		Unknown function	General	1	1	1	1	1	-0.8	0.00219	-1.5	0.00001	-1.5	0.00001	-1.5	0.00001	-1.5	0.01754	-1.1	0.00001	-1.9	0.00001	-1.9	0.00001	-1.9	0.00001	-1.9	
EF2461		Unknown function	General	1	1	1	1	1	-0.7	0.00034	-1.3	0.00000	-1.3	0.00000	-1.3	0.00000	-1.3	0.13830	0.5	0.00358	0.1	0.74680	0.1	0.74680	0.1	0.74680	0.1	
EF2462		Hypothetical proteins	Conserved	1	1	1	1	1	-0.2	0.32860	-0.3	0.05390	-0.4	0.02708	-0.4	0.02708	-0.4	0.13830	0.5	0.00358	0.1	0.74680	0.1	0.74680	0.1	0.74680	0.1	
EF2463		Transport and binding proteins	Anions	1	1	1	1	1	-0.2	0.37820	-0.3	0.03618	-0.3	0.32970	-0.3	0.32970	-0.3	0.62430	-0.6	0.00571	-0.3	0.20410	-0.3	0.20410	-0.3	0.20410	-0.3	
EF2464		Hypothetical proteins	Domain	1	0	0	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	0.06020	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF2465		Hypothetical proteins	Domain	1	0	0	0	0	0.1	0.52220	0.2	0.22790	0.2	0.22790	0.2	0.22790	0.2	0.93710	0.9	0.00101	NA	NA	NA	NA	NA	NA	NA	NA
EF2466		Hypothetical proteins	Domain	1	0	0	0	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	0.06020	NA	NA	NA	NA	NA					

Locus	Gene	Functional category	Subcategory	M*	O*	T	S	M	I	S	q-value	M_130	q-value	O_130	q-value	S_130	q-value	S_130	q-value				
EF3132		Hypothetical proteins	Conserved	1	1	1	1	1	1	1	0.8	0.00420	0.4	0.00597	0.3	0.12090	0.0	0.92420	0.4	0.02629	0.6	0.00399	
EF3133		Hypothetical proteins	Conserved	1	1	1	1	1	1	1	1.9	0.00001	NA	NA	NA	0.46210	-0.4	0.03827	0.6	0.00272	-0.2	0.26290	
EF3134	<i>eda-2</i>	Energy metabolism	Entner-Doudoroff	1	1	1	1	1	1	1	1.9	0.00001	NA	NA	0.00013	0.4	0.00013	0.4	0.03827	0.6	0.00272	-0.2	0.26290
EF3135		Energy metabolism	Conserved	1	1	1	1	1	1	1	1.9	0.00001	NA	NA	0.00013	0.4	0.00013	0.4	0.03827	0.6	0.00272	-0.2	0.26290
EF3136		Signal transduction	Sugars	1	1	1	1	1	1	1	2.1	0.00003	NA	NA	1.6	0.00000	NA	NA	NA	0.0	0.00010	-0.8	0.00306
EF3137		Signal transduction	PTS	1	1	1	1	1	1	1	1.8	0.00004	NA	NA	1.6	0.00001	NA	NA	NA	0.0	0.00010	-0.8	0.00306
EF3138		Signal transduction	PTS	1	1	1	1	1	1	1	1.7	0.00069	NA	NA	1.3	0.00001	NA	NA	NA	0.0	0.00021	-0.8	0.01002
EF3139		Signal transduction	PTS	1	1	1	1	1	1	1	1.8	0.00069	NA	NA	1.3	0.00001	NA	NA	NA	0.0	0.00021	-0.8	0.01002
EF3140		Signal transduction	PTS	1	1	1	1	1	1	1	1.6	0.00018	NA	NA	1.1	0.00002	NA	NA	NA	0.0	0.00052	-0.8	0.00750
EF3141		Energy metabolism	Fermentation	1	1	1	1	1	1	1	1.2	0.00061	-0.5	0.01182	1.4	0.00114	NA	NA	NA	0.7	0.00217	-1.0	0.00325
EF3141		Enzymes of unknown specificity	Enzymes of unknown specificity	1	1	1	1	1	1	1	1.2	0.00061	-0.5	0.01182	1.4	0.00114	NA	NA	NA	0.7	0.00217	-1.0	0.00325
EF3142		Energy metabolism	Sugars	1	1	1	1	1	1	1	0.0	0.23120	-1.9	0.00000	1.3	0.00001	-1.5	0.00006	0.5	0.02315	-1.1	0.00043	
EF3144		Regulatory functions	DNA interactions	1	1	1	1	1	1	1	0.0	0.96370	-0.6	0.01470	1.0	0.00018	-0.9	0.00004	0.5	0.02315	-1.1	0.00043	
EF3144		Regulatory functions	DNA interactions	1	1	1	1	1	1	1	0.0	0.96370	-0.6	0.01470	1.0	0.00018	-0.9	0.00004	0.5	0.02315	-1.1	0.00043	
EF3145		Hypothetical proteins		1	1	1	1	1	1	1	0	NA	NA	NA	0.1	0.38080	0.0	0.97590	NA	NA	NA	NA	NA
EF3146		Hypothetical proteins		1	1	1	1	1	1	1	0	NA	NA	NA	0.1	0.38080	0.0	0.97590	NA	NA	NA	NA	NA
EF3146		Fatty acid and phospholipid metabolism	Biosynthesis	1	1	1	1	1	1	1	-0.5	0.02045	-0.4	0.05364	0.2	0.06261	0.2	0.97770	NA	0.40940	-0.3	0.09809	
EF3148	<i>pgsA</i>	Fatty acid and phospholipid metabolism	Degradation of proteins, peptides, and glycopeptides	1	1	1	1	1	1	1	-0.3	0.21900	-0.3	0.35700	-0.2	0.33760	0.2	0.49100	-0.6	0.00320	-0.6	0.09418	
EF3150		Protein fate	Conserved	1	1	1	1	1	1	1	-0.4	0.10310	-0.5	0.00189	0.3	0.22950	0.2	0.40750	-0.3	0.05677	-0.1	0.58830	
EF3151		Hypothetical proteins	Adaptations to atypical conditions	1	1	1	1	1	1	1	0.0	0.95210	0.0	0.72560	0.4	0.03161	-0.2	0.39590	-0.1	0.55750	-0.4	0.04186	
EF3152		Cellular processes	Conserved	1	1	1	1	1	1	1	0.6	0.00117	3.9	0.00000	0.2	0.32070	1.7	0.00004	0.5	0.00182	3.2	0.00000	
EF3153		Hypothetical proteins	Conserved	1	1	1	1	1	1	1	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF3154		Hypothetical proteins	Conserved	1	1	1	1	1	1	1	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF3155		Hypothetical proteins	Conserved	1	1	1	1	1	1	1	0	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
EF3156		Regulatory functions	DNA interactions	1	1	1	1	1	1	1	-0.3	0.05986	-0.8	0.00001	-0.1	0.55810	-0.2	0.35550	-0.1	0.39990	0.0	0.99700	
EF3157		Energy metabolism	Biosynthesis and degradation of polysaccharides	1	1	1	1	1	1	1	-1.0	0.00004	-2.5	0.00000	-0.8	0.00034	-1.1	0.00124	-1.0	0.00001	-2.0	0.00001	
EF3158		Unknown function	Enzymes of unknown specificity	1	1	1	1	1	1	1	0.3	0.10740	-1.5	0.00000	0.4	0.01167	-0.5	0.06831	0.6	0.00307	-2.0	0.00006	
EF3160		Hypothetical proteins	Enzymes of unknown specificity	1	1	1	1	1	1	1	0	NA	NA	NA	NA	NA	NA	NA	NA	0.40160	-0.0	0.96920	
EF3161		Hypothetical proteins		1	1	1	1	1	1	1	0	NA	NA	NA	NA	NA	NA	NA	NA	0.40160	-0.0	0.96920	
EF3163	<i>prxA-2</i>	Purines, pyrimidines, nucleosides, and nucleotides	Purine ribonucleotide biosynthesis	1	1	1	1	1	1	1	-0.3	0.12000	0.5	0.00163	-0.6	0.00437	-0.1	0.52240	-0.2	0.24110	0.1	0.53460	
EF3164	<i>msbB</i>	Cellular processes	General	1	1	1	1	1	1	1	-0.6	0.00795	4.4	0.00000	0.3	0.22430	0.7	0.00005	0.0	0.85760	3.3	0.00000	
EF3165	<i>msf</i>	Cellular processes	Cell division	1	1	1	1	1	1	1	-0.5	0.00530	-0.3	0.04687	0.0	0.91570	0.2	0.42030	-0.1	0.45400	-0.1	0.49550	
EF3166	<i>hevB</i>	DNA metabolism	DNA replication, recombination, and repair	1	1	1	1	1	1	1	-0.6	0.00374	-0.8	0.00002	0.1	0.76420	0.1	0.74150	0.0	0.73440	-0.1	0.45610	
EF3167	<i>hevA</i>	DNA metabolism	DNA replication, recombination, and repair	1	1	1	1	1	1	1	-0.2	0.40130	-0.2	0.34230	0.0	0.92960	0.1	0.76660	0.1	0.61530	-0.2	0.15910	
EF3168		Hypothetical proteins	Conserved	1	1	1	1	1	1	1	0.1	0.58430	-0.3	0.04514	0.0	0.88980	-0.2	0.25550	0.1	0.23680	0.1	0.33930	
EF3169		Hypothetical proteins	Conserved	1	1	1	1	1	1	1	-0.1	0.78370	-0.1	0.63940	0.1	0.81100	0.1	0.70120	0.2	0.14920	-0.3	0.05645	
EF3170		Unknown function	Conserved	1	1	1	1	1	1	1	0.0	0.99240	-0.6	0.00013	-0.8	0.08726	0.3	0.29670	-0.1	0.78840	-0.2	0.21080	
EF3171	<i>recA</i>	DNA metabolism	DNA replication, recombination, and repair	1	1	1	1	1	1	1	-0.2	0.18710	0.9	0.00000	-0.1	0.78540	0.0	0.90910	0.0	0.78500	0.1	0.00006	
EF3172	<i>cinA</i>	DNA metabolism	DNA replication, recombination, and repair	1	1	1	1	1	1	1	0.6	0.00780	1.4	0.00000	0.4	0.65732	1.0	0.00165	0.7	0.00641	1.0	0.00058	
EF3173		Hypothetical proteins	Conserved	1	1	1	1	1	1	1	-0.3	0.21540	-1.7	0.00000	-1.9	0.03580	-1.9	0.00017	0.2	0.19680	-1.0	0.00194	
EF3174		Hypothetical proteins	Conserved	1	1	1	1	1	1	1	0.1	0.63830	0.6	0.00060	0.2	0.14210	0.0	0.84240	-0.5	0.06317	0.6	0.00692	
EF3175		Cell envelope	Other	1	1	1	1	1	1	1	-0.1	0.61590	-0.1	0.63550	0.0	0.87040	-0.6	0.03584	-0.2	0.00193	-0.3	0.13020	
EF3177		Hypothetical proteins	Conserved	1	1	1	1	1	1	1	-0.2	0.55450	1.2	0.00000	0.3	0.86640	0.6	0.01114	0.3	0.44170	0.1	0.00005	
EF3178		Protein fate	Degradation of proteins, peptides, and glycopeptides	1	1	1	1	1	1	1	1.4	0.00004	2.9	0.00000	1.3	0.00008	1.1	0.00153	1.4	0.00000	2.5	0.00000	
EF3179		Hypothetical proteins	Conserved	1	1	1	1	1	1	1	-0.1	0.68210	0.3	0.00644	-0.3	0.80743	0.4	0.15270	-0.2	0.11840	0.5	0.01954	
EF3180	<i>sigV</i>	Transcription	Transcription factors	1	1	1	1	1	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	0.11840	0.5	0.01954	
EF3181		Hypothetical proteins	Conserved	1	1	1	1	1	1	1	0.3	0.12990	1.1	0.00000	-0.2	0.41550	0.3	0.21450	0.2	0.29950	1.1	0.00103	
EF3182		Hypothetical proteins	Conserved	1	1	1	1	1	1	1	-0.2	0.35570	0.7	0.00000	-0.4	0.36240	0.2	0.22280	0.2	0.29950	1.1	0.00103	
EF3183		Cell envelope	Other	1	1	1	1	1	1	1	-0.7	0.00440	-1.1	0.00060	-0.4	0.02083	-0.4	0.07663	NA	NA	-0.1	0.60130	
EF3184		Hypothetical proteins	Conserved	1	1	1	1	1	1	1	-0.6	0.00736	-0.5	0.00049	-0.2	0.18870	-0.2	0.22280	0.0	0.86310	0.0	0.88000	
EF3185		Hypothetical proteins	Conserved	1	1	1	1	1	1	1	-0.6	0.00179	-1.5	0.00000	-0.2	0.59400	-0.2	0.48970	-0.2	0.29140	-0.1	0.71930	
EF3186		Hypothetical proteins	Conserved	1	1	1	1	1	1	1	-0.6	0.00198	-1.0	0.00001	-0.2	0.96000	-0.1	0.51900	NA	NA	-0.1	0.83360	
EF3187		Cell envelope	Other	1	1	1	1	1	1	1	-0.8	0.00139	-0.9	0.00001	0.4	0.04266	0.2	0.22460	-0.3	0.18550	-0.2	0.49170	
EF3188		Hypothetical proteins	Conserved	1	1	1	1	1	1	1	-0.7	0.00221	-0.2	0.05726	0.3	0.04266	0.2	0.22460	-0.3	0.18550	-0.2	0.49170	
EF3189		Hypothetical proteins	Conserved	1	1	1	1	1	1	1	-0.4	0.02491	-0.2	0.26690	0.2	0.19540	0.1	0.49830	-0.5	0.01344	-0.3	0.10410	
EF3191		Fatty acid and phospholipid metabolism	Degradation	1	1	1	1	1	1	1	0.4	0.18040	0.9	0.00061	0.3	0.06230	0.5	0.01428	0.1	0.73340			

