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6 **Extreme short-day induction requirements for flowering strawberry**
7 **cultivar ‘Malwina’**

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16 **ABSTRACT**

17 We studied short-day induction of the strawberry cultivar ‘Malwina’ under both phytotron and
18 field conditions. Flowering was assessed by crown dissection of treated plants and subsequent
19 flowering performance. Serial dissections revealed no visible changes in crown apices during
20 the first 4 weeks of short day (SD) at 18°C in the phytotron, while after 6 weeks, all plants had
21 formed rudimentary flower primordia with visible sepals. At 9°C, the same stages were reached
22 after 8 and 10 weeks of SD, respectively. When subsequently forced in long day (LD) at 20°C,
23 no substantial flowering took place after less than 6 weeks SD treatment at 18°C, while full
24 flowering required 10 weeks of SD induction. At 9°C, full flowering was not obtained even
25 after 10 weeks of SD treatment. Under field conditions, the ‘Malwina’ plants did not reach
26 floral development stage 2 before 22 October, approximately a month after ‘Frida’ and ‘Sonata’
27 which reached this stage on 21 September, and three weeks after ‘Florence’. SD exposure
28 resulted in repeated crown branching in ‘Malwina’ and we suggest that early spontaneous
29 abortion of the emerging floral primordium takes place under unsaturated SD induction
30 conditions, thus causing crown branching and hence, delayed floral initiation and development.

31

32 **KEYWORDS**

33 Floral initiation; ‘Malwina’ strawberry; photoperiod; SD requirement; temperature

34 **Introduction**

35 Because of the economic importance of the cultivated strawberry (*Fragaria x ananassa* Duch.),
36 the flowering physiology of the species has been extensively researched and reviewed. Most
37 seasonal flowering (June-bearing) strawberry cultivars are classified as facultative short day
38 (SD) plants. At temperatures above 18-20°C, they require exposure to SD for induction of
39 flowering, while at lower temperatures, most cultivars also initiate flowers in long days (LD)
40 (Guttridge, 1985; Heide et al., 2013). However, the flower-inducing effect of SD is highly
41 temperature dependent and is optimal at intermediate temperatures, while at temperatures <
42 12°C and > 21°C, short day induction efficiency is progressively declining (Guttridge, 1985;
43 Heide et al., 2013). The critical photoperiod for SD induction is about 14-15 h, depending on
44 the cultivar (Darrow & Waldo, 1934; Konsin et al., 2001). Therefore, under natural
45 environment conditions, floral initiation takes place in late summer and early autumn when
46 photoperiod and temperature become conducive for floral induction (Guttridge, 1985; Heide et
47 al., 2013).

48 The minimum number of SDs required for induction of flowering is between 7 and 14, but
49 can vary considerably in response to temperature conditions, length of the photoperiod, and
50 daily light integral (Guttridge, 1985; Heide et al., 2013). With extension of the SD period
51 beyond the critical length, the number of initiated flowers increases linearly with the additional
52 number of SD cycles, at least up to 49 cycles, while further initiation ceases immediately when
53 the plants are transferred to LD conditions (Konsin et al., 2001; Heide et al., 2013). For
54 commercial greenhouse production, SD periods of 3-5 weeks duration are usually
55 recommended. However, the SD requirement can vary considerably among cultivars, early
56 cultivars generally needing the lowest number of SD cycles (Heide et al., 2013). In an
57 experiment with six cultivars of distant origin, Sønsteby and Heide (2017) found that 4 weeks
58 of 10-h SD at intermediate temperatures induced profuse flowering in all cultivars except the
59 late-flowering and late-maturing 'Malwina' (Stoppel, 2012), which produced only a few
60 flowers in a single plant at 15°C. By comparison, the cultivar 'Florence', which is also known
61 to be slow-responding and late flowering (Opstad et al., 2011), produced profuse flowering
62 with the 4-week induction period at both 15 and 21°C. After autumn-planting and
63 overwintering in the field, flowering and fruit ripening was also delayed by 2-3 weeks in
64 'Malwina' compared with 'Florence' and even more so compared with the other cultivars
65 (Sønsteby & Heide, 2017).

66 Basically, there are two principally different response patterns that can explain such a
67 delayed flowering response: 1) the plants have an exceptionally short critical photoperiod for

68 floral induction which under natural light conditions postpones the date when the critical
69 daylength is reached, or 2) the plants have a normal critical photoperiod but require an
70 exceptionally large number of SD cycles for initiation of flowering. Since the 10-h photoperiod
71 used in the cited experiment by Sønsteby and Heide (2017) is 4-5 h shorter than the critical
72 photoperiod commonly found in seasonal flowering strawberry cultivars (Guttridge, 1985;
73 Heide et al., 2013), the results strongly support the second alternative.

74 In order to learn more about this unusual flowering behaviour, we have studied flower
75 induction in ‘Malwina’ strawberry in more detail under both phytotron and field conditions.

76

77 **Materials and methods**

78 *Plant materials and handling*

79 For the phytotron experiment, stock plants were dug in the field in early August at the NIBIO
80 Experimental Centre Apelsvoll in South East Norway (60°40’N, 10°40’E, 250 m above sea
81 level) and brought into a heated greenhouse maintained at minimum 21°C and 20 h
82 photoperiod. Runners were collected from these plants on 6 September and rooted in 9 cm
83 plastic pots in a peat based potting compost (Gartnerjord, LOG, Oslo, Norway with 10% added
84 granulated perlite) under saturated atmosphere and the same temperature and light conditions.
85 On 4 October, when the plants were well rooted, they were moved into the daylight phytotron
86 of the Norwegian University of Life Sciences at Ås, Norway, where they were exposed to 10-
87 h photoperiod at 9 and 18°C for 4, 6, 8, or 10 weeks. In the phytotron, the plants received
88 natural daylight supplemented by 150 $\mu\text{mol quanta m}^{-2} \text{ s}^{-1}$ artificial light supplied by 400 W
89 Philips HPI-T lamps from 0800 h to 1800 h. Control plants were exposed to 20 h photoperiod
90 established by daylength extension with 80 W incandescent lamps. Temperatures were
91 controlled to $\pm 1.0^\circ\text{C}$, and a water vapour pressure deficit of 530 Pa was maintained at both
92 temperatures throughout day and night.

93 For the field experiment, over-wintered plug plants were received from an authorized
94 producer and planted in the field at Apelsvoll in early June 2018 on raised beds with black
95 polyethylene mulch at a spacing of 25 cm x 40 cm x 160 cm. Flowers were removed as they
96 appeared, while runners were allowed to grow until 3 September when all runners were
97 removed. For comparison, the well-known cultivars ‘Florence’, ‘Frida’ and ‘Sonata’ were
98 included in the experiment. These plants were rooted current year runners which were planted
99 in the field in early August 2018. Otherwise, the plants were treated as described above for the
100 ‘Malwina’ plants.

101 Starting on 17 August, the progress of the floral initiation process of the plants was
102 monitored by sampling and dissection of crowns at approximately 10-day intervals. Starting
103 on 1 October, samples of plants were also dug at monthly intervals and forced in a greenhouse
104 at 21°C and 20 h photoperiod for assessment of flowering performance. Furthermore, two
105 groups of 15 plants were overwintered in the field, of which one group was dug in late April
106 of the following spring for forcing in the greenhouse as ascribed above, while the other group
107 was allowed to flower and fruit in the field for assessment of flowering and yield performance.
108 Temperature conditions at Apelsvoll in the 2018-2019 autumn and winter season are shown in
109 Figure 1.

110

111 **Experimental design, data collection and analyses**

112 The experimental design of the phytotron experiment was a split plot with temperatures as main
113 plots and photoperiods as subplots. Each treatment had three replications, each with 14 plants
114 on a separate trolley (9 plants for dissection and 5 for flowering performance). Plant growth
115 and development during SD treatment was monitored by counting of the numbers of leaves,
116 runners and crowns and by measurement of petiole length of the last fully developed leaf in
117 each plant at completion of each treatment duration. On each occasion, 9 plants in each
118 treatment were dissected for assessment of the flower development status of the main crown
119 as described by Opstad et al. (2011), while 5 plants in each treatment were transplanted to 12
120 cm pots and set to flower in a heated greenhouse at 20°C and 20 h photoperiod. Flower
121 development status of the dissected plants was scored according to a six-stage scale where
122 stage 1 denotes entirely vegetative apices, and stage 2 the first visible sign of transition to
123 generative development, while stage 6 denotes fully differentiated primary flower primordia
124 (cf. Opstad et al., 2011). Flowering performance of the forced plants was recorded after 10
125 weeks of forcing of plants from each treatment.

126 The field experiment had three replicate beds with 60 plants each of each cultivar. At each
127 sampling date, 2 plants from each replicate bed were dissected and examined for flowering
128 status ($n = 6$). The dissections followed the same procedures as described above for the
129 phytotron experiment. For assessment of flowering performance in the greenhouse and in the
130 field, 5 plants from each replicate bed were used ($n = 15$).

131 Experimental data were subjected to analysis of variance (ANOVA) by standard procedures
132 using a MiniTab[®] Statistical Software program package (Release 15, Minitab Inc., State
133 College, PA, USA). Percentage values were always subjected to an arc sin transformation
134 before performance of the ANOVA.

135 **Results**

136 *Phytotron experiment*

137 Runner formation declined sharply when the plants were transferred to SD and ceased almost
138 completely after 4 weeks at both 18 and 9°C, while production of leaves continued at more or
139 less constant, but temperature dependent rates in SD (Figure 2). This was accompanied by a
140 strong and parallel decline in petiole length of the new-formed leaves at both temperatures. On
141 the other hand, little crown branching took place during the first 8 weeks of SD, where after it
142 started to increase at both temperatures.

143 Serial dissections of crowns revealed no visible changes in the crown apices during the first
144 4 weeks of SD at 18°C, while after 6 weeks, all plants had formed rudimentary flower primordia
145 with visible sepal primordia (stage 2) (Figure 3). With continued SD, there was a more or less
146 linear progress in flower primordia development all the way up to stage 6 after 10 weeks of
147 SD. At 9°C, the first visible changes were observed in one half of the plants after 8 weeks,
148 whereas floral stage 2 in all plants was not reached until 10 weeks of SD treatment.

149 The flowering performance of the plants when forced in LD at 20°C is shown in Table 1.
150 The results show that although a single plant from 18°C formed a few flowers after 4 weeks of
151 SD treatment, no substantial flowering took place with less than 6 weeks of SD treatment, while
152 100% flowering required 10 weeks of SD. Among the plants from 9°C, a couple of plants
153 flowered with 6 weeks of SD, while full flowering was not obtained even after 10 weeks of SD
154 treatment. The number of inflorescences and flowers per plant were always higher in plants
155 exposed to SD at 18°C than in those at 9°C, and at both temperatures the numbers increased
156 steadily with increasing length of SD treatment. With marginal SD induction, a few plants from
157 both temperatures developed pronounced phyllody as shown in Figure 4. In plants from both
158 temperatures, the time to anthesis decreased in parallel with increasing length of SD treatment.
159 The trend of change was the same at both temperatures, but with a delay of approximately two
160 weeks at 9 °C. Although the plants at 18°C had twice as many crowns as those at 9°C after 10
161 weeks of SD (Figure 2), the difference had evened out during the forcing period (Table 1).

162

163 *Field experiment*

164 Also under field conditions, 'Malwina' initiated floral primordia very late, and much later than
165 the other cultivars (Figure 5). Thus, visible floral primordia at stage 2 was not observed until
166 22 October in 'Malwina', 3 weeks after 'Florence' and 5 weeks after 'Frida' and 'Sonata'.
167 Further floral differentiation progressed in parallel in the four cultivars, so that at the last
168 sampling on 9 November, 'Malwina' was still at floral stage 3.5 only, whereas 'Frida' and

169 'Sonata' had fully differentiated terminal flowers on their primary inflorescences, a floral stage
170 that was not reached in the 'Malwina' plants until in the following spring. Crown branching
171 increased rapidly in all cultivars during the first and second week of September, where after it
172 gradually levelled off in parallel with the decreasing autumn temperature (Figure 6). The
173 number of crowns was always highest in the 'Malwina' plants.

174 Greenhouse forcing of 'Malwina' plants dug in the field on 1 October produced similar
175 results (Table 2). Although most plants eventually flowered, it took more than 12 weeks of
176 forcing in LD at 20°C and the plants produced only one or two inflorescences each. When dug
177 on 1 November, all 'Malwina' plants flowered, but still only after nearly 11 weeks of forcing,
178 compared with 5-6 weeks in 'Frida' and 'Florence'. However, flowering was still rather sparse
179 in 'Malwina', with only 3 inflorescences and a total of 16 flowers per plant, compared with 6-
180 7 inflorescences and 45-60 flowers per plant in 'Frida' and 'Florence'. Also, in plants that were
181 overwintered in the field and dug and set to forcing on 23 April (when the soil had thawed),
182 flowering was still 10 to 15 days later in 'Malwina' than in the other cultivars. However, while
183 the number of inflorescences and flowers per plant decreased in 'Frida' and 'Florence' plants
184 forced in spring, it increased slightly in 'Malwina', indicating that continued flower initiation
185 had compensated for losses of flower primordia during the winter (Table 2). Furthermore, the
186 losses of flower primordia were largely eliminated in all three cultivars when the plants were
187 allowed to flower in the field under cooler temperature conditions. This response was most
188 pronounced in 'Frida' where greenhouse forcing in spring reduced flowering by nearly 50%
189 compared with November forcing or spring flowering in the field (Table 2). Another marked
190 difference between the cultivars was that while 'Frida' and 'Florence' plants developed 5-10
191 crowns, the 'Malwina' plants on average produced nearly 20 crowns plant⁻¹ by the time of
192 flowering (Table 2).

193 The yield of the field-grown plants presented in Figure 7, show disappointingly low yields
194 of 'Malwina' compared with the other cultivars. Thus, the yield was only 54% of that of the
195 Norwegian cultivar 'Frida', and 62% of that of 'Sonata'. As usual, the date of 50% harvest was
196 delayed by approximately 3 weeks compared with these two cultivars.

197

198 **Discussion**

199 The results confirm our earlier results (Sønsteby & Heide, 2017) showing that the strawberry
200 cultivar 'Malwina' has an extreme SD induction requirement for flower initiation. Whereas
201 most other SD cultivars produced advanced flower primordia and attained saturated flowering

202 with 4 weeks of SD induction under optimal temperature conditions of 18-21°C (Guttridge,
203 1985; Konsin et al., 2001; Heide et al., 2013; Sønsteby & Heide, 2017), ‘Malwina’ required 10
204 weeks of SD under the same conditions for a similar flowering response. At suboptimal
205 temperatures of 9°C, ‘Malwina’ produced only partial flowering even with 10 weeks of 10-h
206 SD treatment (Figure 2, Table 1). Similarly, under field conditions at Apelsvoll in South East
207 Norway, most SD cultivars developed visible flower primordia around mid-September (Opstad
208 et al., 2011), whereas this stage was delayed for another 5 weeks till about 20 October in
209 ‘Malwina’ (Figure 5). Even the relatively late-flowering and late-maturing cultivar ‘Florence’,
210 which is commonly used for extension of the strawberry marketing season, initiated floral
211 primordia 3 weeks ahead of ‘Malwina’. This extreme SD induction requirement is apparently
212 the main reason for the exceptionally late flowering and fruit maturation experienced in
213 ‘Malwina’ under both experimental and commercial production conditions (Sønsteby & Heide,
214 2017). On the other hand, the slow response to the near-optimal SD photoperiod of 10 h is not
215 compatible with the possibility that an exceptionally short critical photoperiod is the reason for
216 the late flowering of the cultivar. Rather, the prompt cessation of runner formation and strong
217 restriction of petiole length after 4 weeks of SD exposure (Figure 2), indicate normal SD
218 signalling.

219 This unusual physiological behaviour may morphologically be associated with the excessive
220 branching of the crown axis of ‘Malwina’ (Tables 1, 2; Figure 6). Normally, crown branching
221 is the result of terminal flower formation and lateral displacement of the leading shoot (cf.
222 Guttridge, 1985). However, in ‘Malwina’ the crowns had been branching repeatedly before the
223 first inflorescence appeared. This suggests the occurrence of an early spontaneous abortion of
224 the emerging floral primordium. Possibly, this could be caused by some sort of cultivar specific
225 malfunction of the apical meristem. This would have the same effect as a soft pinch in causing
226 outgrowth of subtending lateral meristems. In some ornamental SD plants such as poinsettia
227 (*Euphorbia pulcherrima*), a marginal SD induction has in fact been used to bring about
228 symmetrical branching of the stem (Rünger, 1967). The excessive and repeated branching of
229 ‘Malwina’ (Figure 6, Table 2), provides strong support for the hypothesis. It may be argued
230 that the results of the field experiment are not directly comparable due to different planting
231 dates (early June and August, respectively). However, it is not likely that earlier planting of
232 ‘Malwina’ should result in delayed flowering. Furthermore, coincidence in the timing of crown
233 branching in all cultivars (Figure 6), together with several weeks difference in floral initiation
234 (Figure 5) tend to exclude the possibility that different planting dates could be the reason for
235 the differences in flowering time.

236 However, since the strawberry plant in fact appears to be a negative LD-plant rather than a
237 regular SD-plant with a direct response to SD (cf. Guttridge, 1985), an alternative perspective
238 of the results could also be suggested. Thus, by the use of donor-receptor pairs of runner plants
239 connected by the stolon, Guttridge (1959) found that donor plants in LD delayed and sometimes
240 inhibited flower formation in receptor plants in SD, while donors in SD failed to induce
241 flowering in receptors in LD. Further spectral evidence for induction of flowering in strawberry
242 by release from LD inhibition was provided by studies on the sensitivity of strawberry plants
243 to R and FR irradiation indicating the temporal sensitivity of a LD-plant (Vince-Prue &
244 Guttridge, 1973). It might therefore, be argued that the rapid cessation of runner formation and
245 petiole elongation upon transfer to SD indicates that the photoperiodic response involved is a
246 promotion of runnering by LD. However, the repeated branching of the crown in ‘Malwina’
247 plants during SD induction demonstrate that the mechanism involved is an impairment of the
248 apical development taking place downstream of the triggering photoperiodic response.

249 Whatever the explanation, since flower initiation eventually took place also in the
250 ‘Malwina’ plants, it is evident that an extended period of SD exposure is able to trigger and
251 support the normal development of the inflorescence primordium also in this cultivar.

252 In commercial production, the late flowering characteristic of ‘Malwina’ has been of interest
253 mainly for extension of the marketing season. However, the excessive crown branching of the
254 cultivar (Table 2) results in shoot crowdedness and competition for space and light, and
255 possibly constrained yields. Low yields of ‘Malwina’ has in fact been experienced by
256 strawberry growers in both Norway and Finland (J. Haslestad, Norwegian Agricultural
257 Advisory team), as well as in the present experiment, where ‘Malwina’ yielded only 55 to 60%
258 of ‘Frida’ and ‘Sonata’, respectively. The destiny of ‘Malwina’ in commercial production
259 therefore seems rather uncertain at present.

260

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264

265 **Disclosure statement**

266 No potential conflict of interest was reported by the authors.

267

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Table 1. Flowering performance of ‘Malwina’ strawberry plants exposed to varying durations of SD treatment at 9 and 18°C and subsequently forced in LD at 20°C for 10 weeks. The data are means of three replicates with 5 plants each.

Temperature (°C)	Weeks of treatment	Flowering plants (%)	Days to anthesis	Infloresc. plant ⁻¹	Flowers plant ⁻¹	Flowers inflor ⁻¹	Crowns plant ⁻¹
9	0	0.0	>100	0.0	0.0	0.0	-
	4	0.0	>100	0.0	0.0	0.0	4.0
	6	13.3	96.2	0.1	1.0	1.0	6.5
	8	73.3	75.0	1.9	13.9	5.6	7.3
	10	73.3	71.0	2.3	16.5	5.2	8.1
<i>Mean</i>		<i>32.0</i>	<i>88.4</i>	<i>0.9</i>	<i>6.5</i>	<i>2.4</i>	<i>7.5</i>
18	0	0.0	>100	0.0	0.0	0.0	-
	4	6.7	99.8	0.3	0.7	0.2	6.0
	6	80.0	76.9	5.0	12.8	2.0	7.8
	8	86.7	70.8	6.0	23.2	3.3	7.9
	10	100	59.5	7.5	38.1	4.9	6.1
<i>Mean</i>		<i>54.7</i>	<i>81.4</i>	<i>3.8</i>	<i>14.9</i>	<i>2.1</i>	<i>7.3</i>

Probability level of significance (ANOVA)

Source of variation

Temperature (A)	0.059	ns	0.03	ns	ns	Ns
Weks. of treatment (B)	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001
A x B	0.002	0.003	<0.001	0.03	ns	0.001

Table 2. Flowering performance of field-grown strawberry cultivars after lifting at different times and forcing in a greenhouse at 20°C and 20 h photoperiod for 11 (‘Frida’ and ‘Florence’) or 14 weeks (‘Malwina’). The flowering performance of plants flowering in the field in the spring 2019 is also included. The data are means of three replicates, each with 5 plants of each cultivar.

Cultivar	Date of lifting/ start forcing	Flowering plants (%)	Days to flower emergence	Days to anthesis	Infloresc. plant ⁻¹	Flowers plant ⁻¹	Crowns plant ⁻¹
‘Malwina’	1 Oct. 2018	89	77.3	87.7	1.8	14.4	17.6
	1 Nov. 2018	100	63.2	75.9	3.2	15.7	21.3
	23 Apr. 2019	100	27.6	39.2	3.5	21.7	16.7
	Field flowering	100	-	66.1*	4.9	36.9	19.2
‘Frida’	1 Oct. 2018	-	-	-	-	-	-
	1 Nov. 2018	100	23.3	33.2	7.1	44.9	5.1
	23 Apr. 2019	100	13.2	22.5	3.9	23.2	7.8
	Field flowering	100	-	52.3*	7.9	45.3	10.0
‘Florence’	1 Oct. 2018	-	-	-	-	-	-
	1 Nov. 2018	100	31.5	45.3	6.2	59.6	4.5
	23 Apr. 2019	100	18.2	28.5	4.8	31.0	9.7
	Field flowering	100	-	59.1*	5.6	33.1	9.0

300 *Days from 23 April

FIGURE LEGENDS

Figure 1. Temperature conditions at the NIBIO Experimental Centre Apelsvoll during late summer and autumn in 2018 and winter and spring 2019 (1 August 2018 – 1 July 2019).

Figure 2. Plant growth and development of ‘Malwina’ strawberry plants during 10 weeks of SD treatment at 9 and 18°C. The data are means of three replicates with 5 plants each \pm SE.

Figure 3. Time courses of successive floral development stages of ‘Malwina’ strawberry plants as affected by increasing duration of SD treatment in the phytotron at 9 and 18°C. The data are means of three replications with 9 plants each \pm SE.

Figure 4. Abnormal flower development (phyllody) in ‘Malwina’ strawberry plants after marginal SD induction of 6 weeks at 9°C.

Figure 5. Time courses of successive floral development stages of four strawberry cultivars under natural field conditions at Apelsvoll. The data are means of three replicates with two plants each of each cultivar \pm SE.

Figure 6. Time courses of cumulative crown branching of four strawberry cultivars under natural field conditions at Apelsvoll. The data are means of three replicates with two plants each of each cultivar \pm SE.

Figure 7. Time courses of cumulative berry yield in four strawberry cultivars in 2019. Data are the means of 3 replicate plots with 0 plants per plot of each cultivar \pm SE.

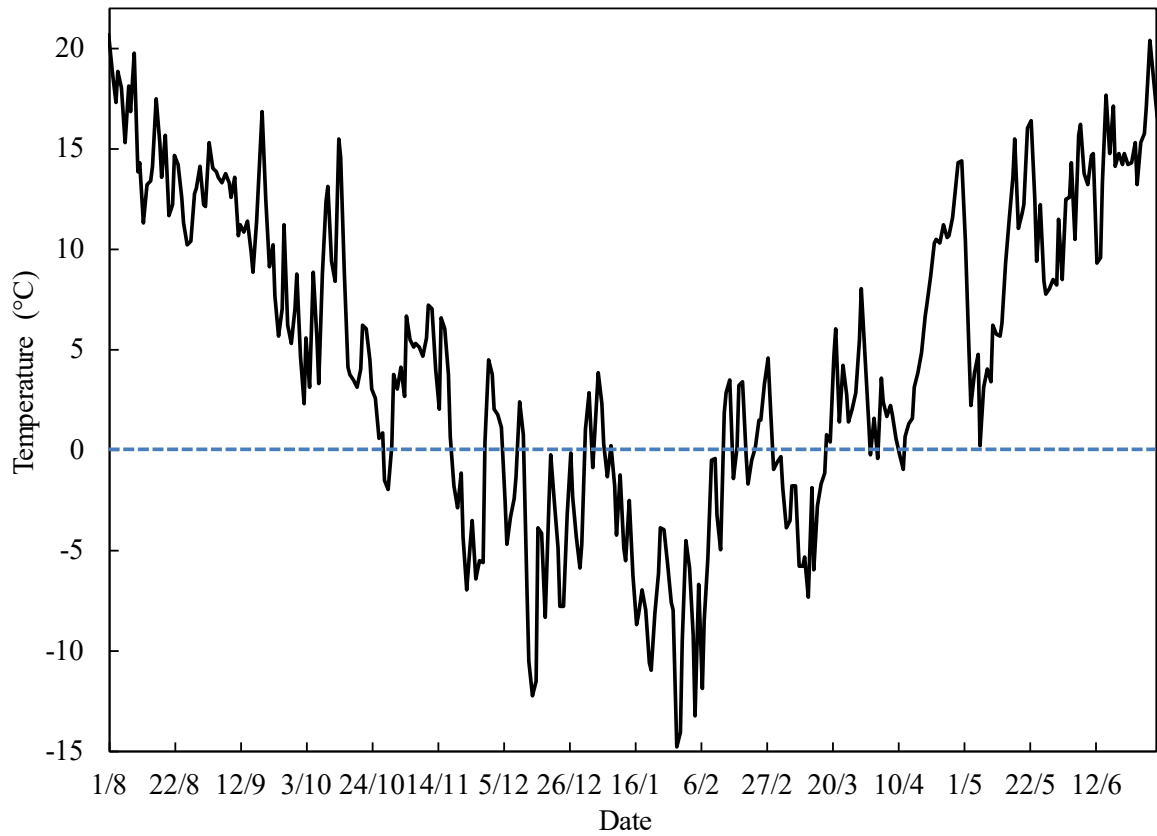


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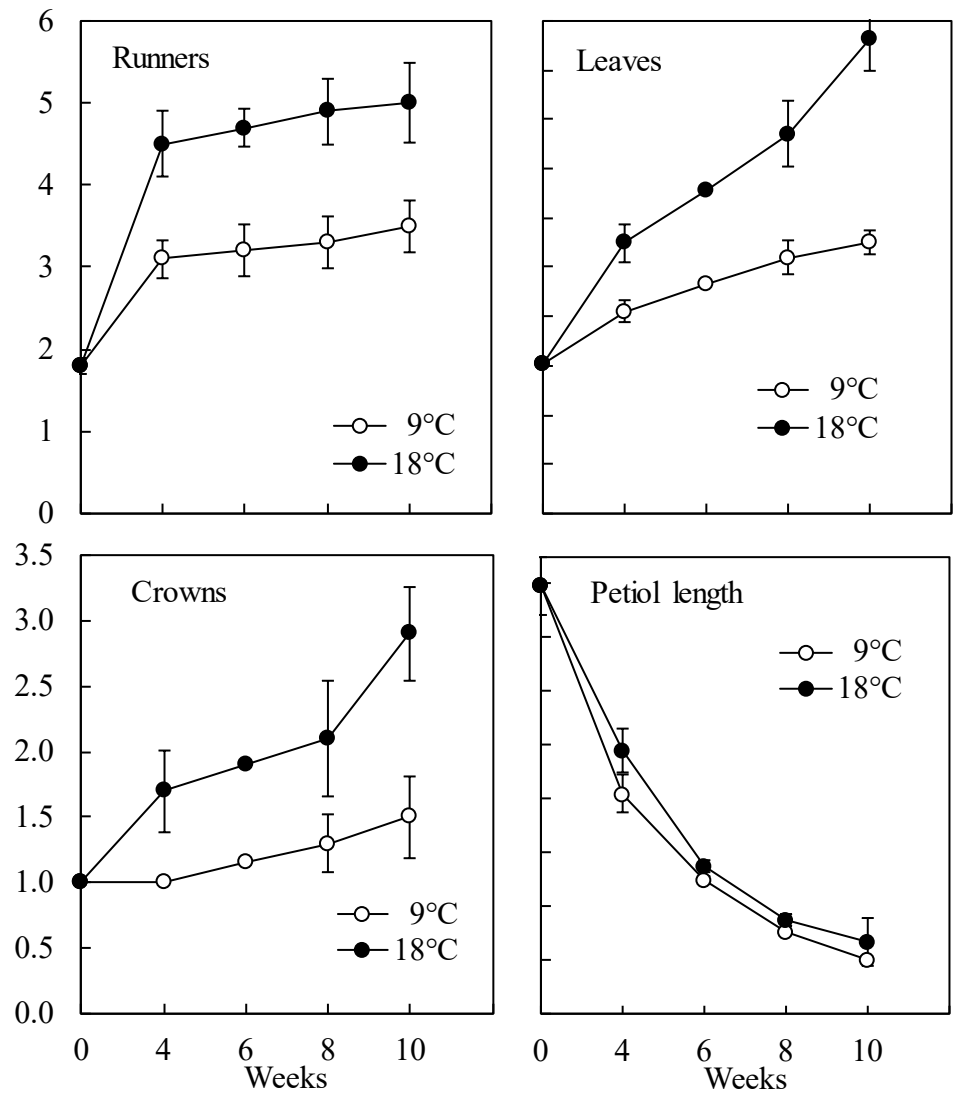


Figure 2. Plant growth and development of 'Malwina' strawberry plants during 10 weeks of SD treatment at 9 and 18°C. The data are means of three replicates with 5 plants each \pm SE.

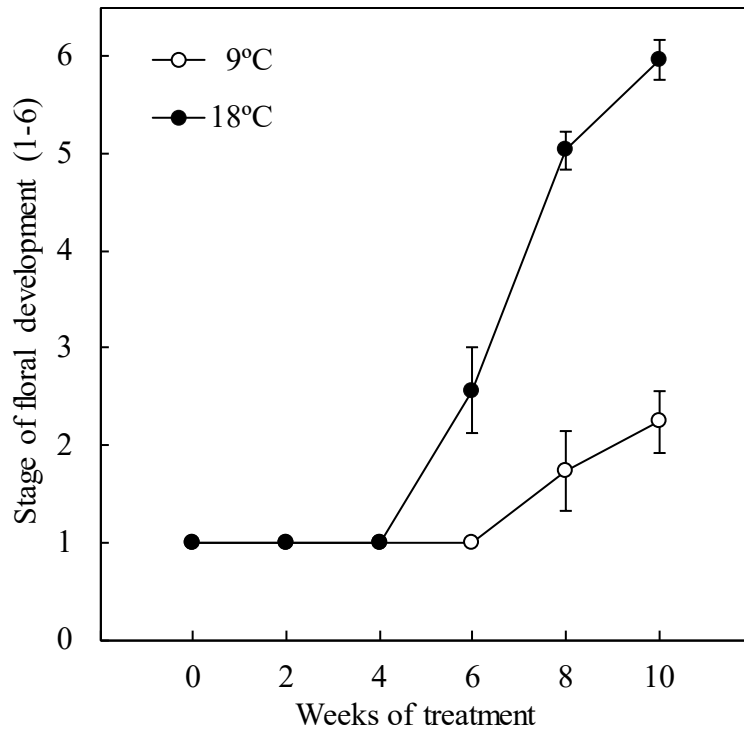


Figure 3. Time courses of successive floral development stages of ‘Malwina’ strawberry plants as affected by increasing duration of SD treatment in the phytotron at 9 and 18°C. The data are means of three replications with 9 plants each \pm SE.



Figure 4. Abnormal flower development (phyllody) in 'Malwina' strawberry plants after marginal SD induction of 6 weeks at 9°C.

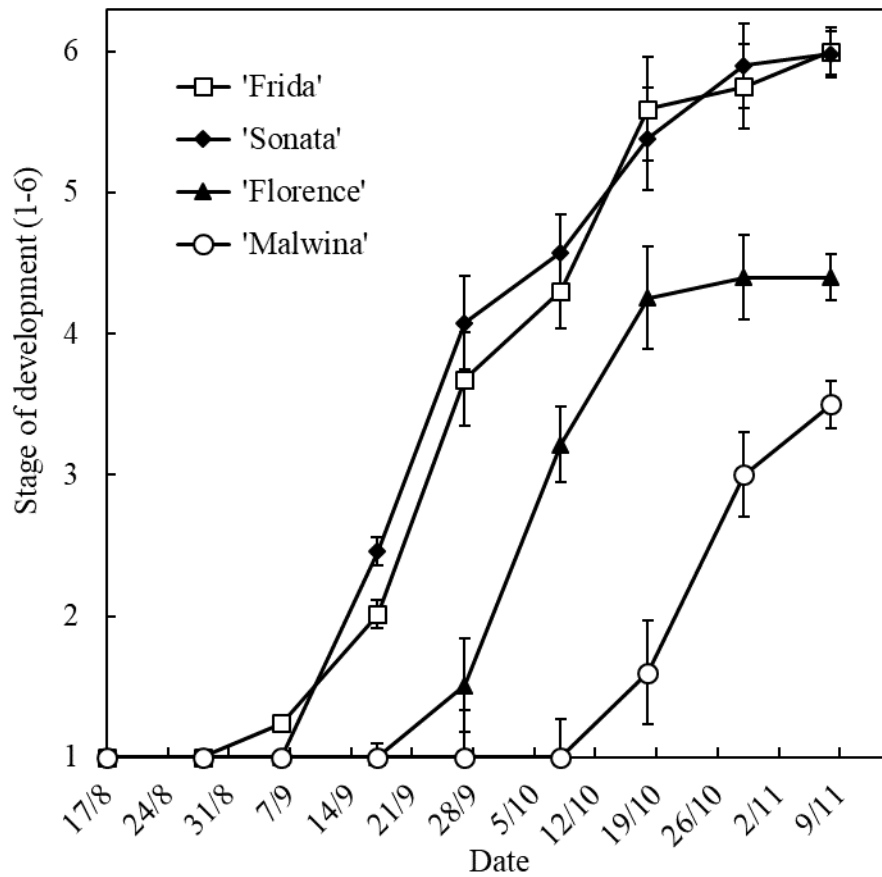


Figure 5. Time courses of successive floral development stages of four strawberry cultivars under natural field conditions at Apelsvoll. The data are means of three replicates with two plants each of each cultivar \pm SE.

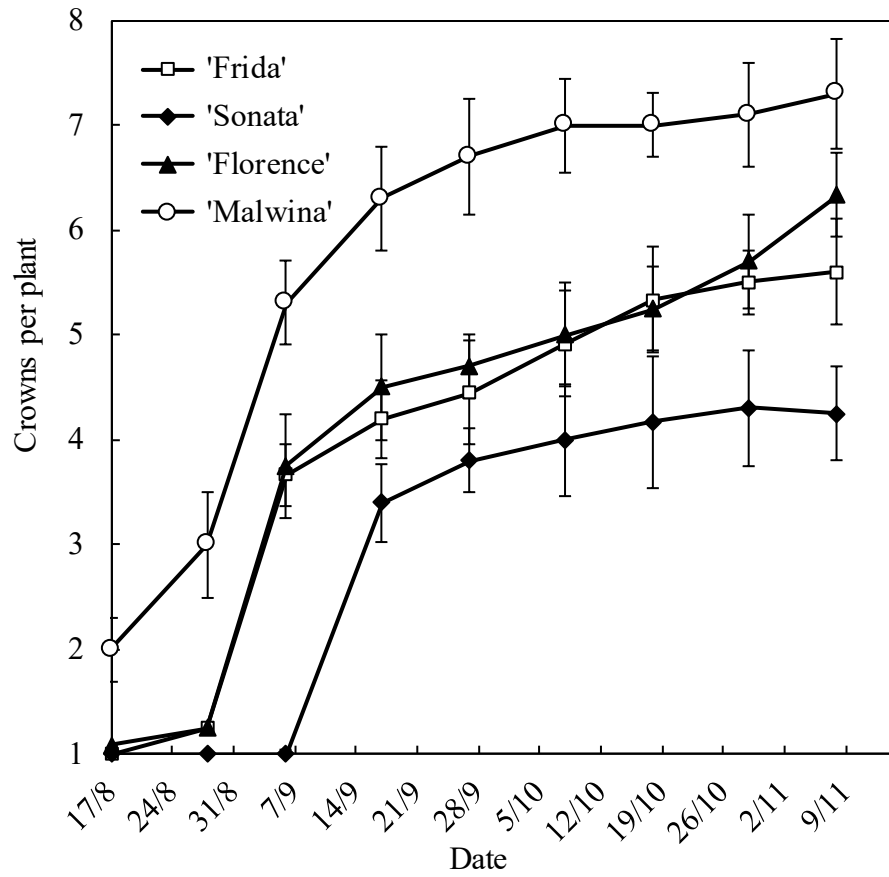


Figure 6. Time courses of cumulative crown branching of four strawberry cultivars under natural field conditions at Apelsvoll. The data are means of three replicates with two plants each of each cultivar \pm SE.

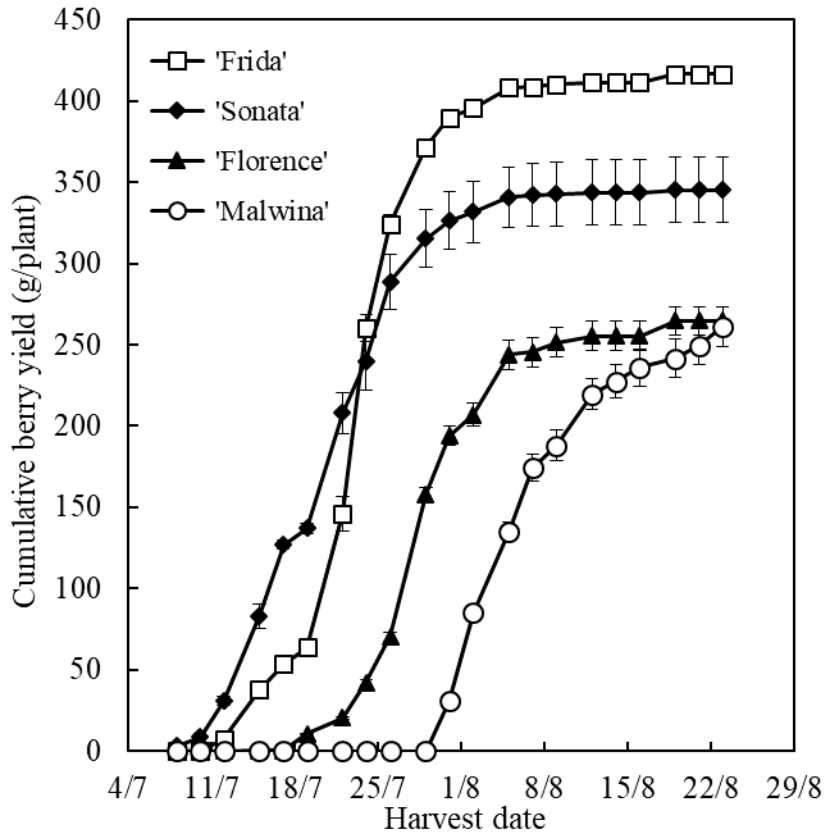


Figure 7. Time courses of cumulative berry yield in four strawberry cultivars in 2019. Data are the means of 3 replicate plots with 20 plants per plot of each cultivar \pm SE.